academicJournals

Vol. 5(7), pp. 166-171, July, 2013 DOI: 10.5897/IJFA2013.0333 ISSN 1996 - 0840 © 2013 Academic Journals http://www.academicjournals.org/IJFA

Full Length Research Paper

Nutritive value of cultured white leg shrimp Litopenaeus vannamei

Gunalan B¹*, Nina Tabitha S.¹, Soundarapandian P.¹ and T. Anand²

¹CAS in Marine Biology, Faculty of Marine Sciences, Annamalai University, Parangipettai, Tamil Nadu, India. ²Department of Fisheries Environment, TANUVAS, Fisheries College and Research Institute, Tuticorin, Tamil Nadu, India.

Accepted 21 January, 2013

Biochemical assays play a major role in recent years. The biochemical composition is the yardstick to measure and assess the nutritional quality of food sources. In the present study, protein, carbohydrate, lipid, moisture and ash in Litopenaeus vannamei was 35.69, 3.20, 19, 76.2 and 1.2% respectively. Calcium content was maximum (154.5 mg) followed by sodium (67.7 mg) and potassium (56.7 mg). Manganese was reported to be minimum (0.898 mg). Copper and chromium were observed in trace. Totally 18 amino acids were detected, among these, argentine, histamine, isoleucine, leucine, methionine, phenylalanine, tryptophan, lysine and valine are essential amino acids and alanine, asparagine, aspartic acid, cysteine, glutamic acid, glycine, proline, serine and tyrosine are nonessential amino acids. In individual essential amino acids, valine (23.72%) was maximum, followed by lysine (13.42%) and methionine (13.06%). Histidine was minimum (1.08%). Glycine (9.8%), cystine (5.56%) and proline (4.26%) contributed as major non-essential amino acids. Ten individual fatty acids were identified, which includes both unsaturated and saturated fatty acids. Three saturated fatty acids (SFA) were recorded (Palmitic acid, Margaric acid and Stearic acid). The polyunsaturated fatty acids (PUFA) were the most dominant common fatty acids (38.5%) with the higher levels of linoleic acid (16.3%) and alpha-linolenic acid (11.2%). Oleic acid is the only monounsaturated fatty acid (MUFA) contributed 12.48% of total fatty acids. At the same time the Omega – 6 and omega - 3 fatty acids accounted for 16.3 and 35.4% of the total PUFA (51.7%). the present study confirming that based on the results, L.vannamei species can be considered as a good source of fatty acid as well as protein.

Key words: Litopenaeus vannamei, proximate composition, fatty acids, amino acids, minerals, shrimp nutrition.

INTRODUCTION

Shrimp is one of the world's most popular shellfish. It provides high quality rich protein, calcium and various extractable compounds and minerals for human body, while low in calorie and fat (Abdullah et al., 2009). Lipid of shrimp contains mostly polyunsaturated fatty acids (essential fatty acids). These essential fatty acids are available in shrimp provides health benefits for human e.g., eye (retina) and brain development and function (Conner et al., 1992).

There are many inorganic elements in the body of shrimp that support associated vital physiological functions. Although shrimps are capable of extracting some of the elements from water, they do respond to dietary sources (Deshimaru and Yone, 1978; Kanazawa

*Corresponding author. E-mail: aquagunal@gmail.com. Tel: 04144- 243223. Fax: 04144- 243553.

et al., 1984; Davis et al., 1992. Since these micronutrients are essential, their absence in the diet may lead to deficiency disease. Some elements, such as copper, zinc, manganese, iron and chromium have useful biological function and are found in shrimp at acceptable levels are very useful for human health (Abdullah et al., 2009). In India, there is no published report on biochemical composition of *Litopenaeus vannamei*. Therefore, the present investigation is the first of its kind on proximate composition of basic biochemical constituents, such as total protein, carbohydrate, lipid, amino acids, fatty acids, moisture and ash in the muscle of *L. vannamei*.

MATERIALS AND METHODS

The shrimps (*L. vannamei*) were collected from the Suriya Marine shrimp farm in Bhimavaram, West Godavari district, Andrapradesh, India. Their lengths ranging from 125 to 130 mm. Samples were washed with deionized water to remove any adhering contamination and drained using filter paper. Samples were brought to the laboratory in ice using insulated containers. The shrimp exoskeletons were pealed out and the meat was homogenized. The grounded samples were oven dried at 70°C and ground into a fine powder.

Analysis of proximate composition

The protein, carbohydrate and lipid contents were estimated by adopting the standard methods of (Lowry et al., 1951; Dubois et al., 1956; Folch et al., 1956) respectively. 1 g of powdered tissue in a porcelain crucible was kept in a muffle furnace at 60°C for 4 h. The residue ash content was weighed and the percentage was calculated. Moisture content was estimated by hot air oven method and minerals were analyzed by following the method of A.O.A.C. (1990). Triplicate reading were taken.

Analysis of fatty acids

The samples were oven dried at 70°C for 24 h until no more weight reduction was observed. After that the samples were grounded with pestle and mortar. To the 100 to 200 mg of finely ground tissue samples 1:1 ratio of chloroform: methanol (2 ml) was added and kept for 30 s. The residual matter was removed by filtering through Whatman No.1filter paper (125 mm). This was washed with 1 ml of chloroform: methanol (2:1vol) to remove the inorganic substances from the combined extract by partition and treated with chloroform: methanol: water (8:4:3) where the lower phase evaporated to dryness. The dried matter was subjected in a sealed test tube with 3% Methanolic HCL at 80°C for 18 h. To this 2 ml of hexane was added to extract the fatty acid methyl esters (FAME) obtained from methanol phase in Hexane. Top 1 ml of the hexane phase was collected in a micro vial. The residual fraction was dissolved in 10/µl of ethyl acetate and injected 1/µl aliquot into a gas chromatograph equipped with flame identification detector and column (HP ULTRA - 225 m, 0.2 mm ID) by gas chromatograph (Kashiwagi et al., 1997).

Estimation of amino acids

The samples were dried at 60°C for 24 h in an oven and packed in

Table 1. Proximate composition in the flesh of L. vannamei.

Parameter	%
Crude protein	35.69 ± 0.5
Crude carbohydrate	3.20 ± 0.3
Crude lipid	19.00 ± 0.6
Moisture	76.2 0± 0.5
Ash	1.20 ± 0.6

Table 2. Mineral composition in the flesh of L. vannamei.	ral composition in the flesh of L. vannan	nei.
---	---	------

Minerals	mg/g
Calcium (Ca)	154.5
Magnesium (Mg)	13.41
Sodium (Na)	67.7
Potassium (K)	56.7
Phosphorus (P)	6.98
Manganese (Mn)	0.898
Iron (Fe)	4.54
Copper (Cu)	in trace
Chromium (Cr)	in trace

airtight polyethylene covers and kept in desiccators. The oven-dried samples were finely grounded before estimating amino acid profile. Amino acids were estimated by using HPLC – Lachromemerck in SPD- 10A VP Detector.

RESULTS

Proximate composition

The percentage of protein, carbohydrate, lipid, moisture and ash in *L. vannamei* was 35.69, 3.20, 19, 76.2 and 1.2% respectively (Table 1).

Mineral composition (mg/g)

The minerals of the *L. vannamei* flesh are shown in Table 2. Calcium content was maximum (154.5 mg) followed by sodium (67.7 mg) and potassium (56.7 mg). Manganese was reported to be minimum (0.898 mg). Copper and chromium were observed in trace amounts. The mineral composition in *L. vannamei* flesh was in the following order; Ca > Na >K>Mg>P>Fe>Mn>Cu, Cr.

Amino acid profile

The amino acid profiles detected from the flesh of the *L. vannamei* is presented in Table 3. Totally 18 amino acids were detected, among these, arginine, histidine, isoleucine, leucine, methionine, phenylalanine, tryptophan,

Table 3. Essential and non essential amino acids in the flesh of
L. vannamei.

Essential amino acids (EAA)	EAA (%)
Arginine	1.2
Histidine	1.08
Isoleucine	12.3
Leucine	5.63
Lysine	13.42
Methionine	13.06
Phenylalanine	1.27
Tryptophan	1.3
Valine	23.72
Total	72.98
Non essential amino acids (NEAA)	NEAA (%)
Alanine	1
Asparagine	0.056
Aspartic acid	1.46
Cystine	5.56
Glutamic acid	2.51
Glycine	9.8
Proline	4.26
Serine	2.66
Tyrosine	2.51
Total	29.816

lysine and valine are essential amino acids and alanine, asparagine, aspartic acid, cysteine, glutamic acid, glycine, proline, serine and tyrosine are non-essential amino acids. In individual essential amino acids, valine (23.72%) was maximum, followed by lysine (13.42%) and methionine (13.06%). Histidine was minimum (1.08%). Glycine (9.8%), cystine (5.56%) and proline (4.26%) contributed as major non-essential amino acids. Asparagine (0.05%) showed the lowest concentration among the non-essential amino acids.

Fatty acid profile

The fatty acid profile from the flesh of the *L. vannamei* (Table 4) show the presence of ten individual fatty acids, which includes both unsaturated and saturated fatty acids. Three saturated fatty acids (SFA) were recorded (Palmitic acid, margaric acid and stearic acid). Among three saturated fatty acids, stearic acid was maximum (12.88%) followed by palmitic and margaric acid. The polyunsaturated fatty acids (PUFA) were the most dominant common fatty acids (38.5%) with the higher levels of linoleic acid (16.3%) and alpha-linolenic acid (11.2%). Oleic acid is the only monounsaturated fatty acids. At the same time the Omega – 6 and omega - 3 fatty acids

accounted for 16.3 and 35.4% of the total PUFA (51.7%).

DISCUSSION

Shrimp is considered as a high-range protein containing nutrient like fish, which contain 8 to 20% protein. It has been reported that protein content of shrimp ranged between 17 and 21% depending on shrimp species (Sriket et al., 2007; Yanar and Celik, 2006). According to Sambhu and Jayaprakash (1994), the protein level in *Penaeus indicus* was varied from 44.62 to 80.87%. The high protein content in the lowest size groups may be attributed to increased protein synthesis during the active growth phase as it has been observed elsewhere in shrimps and mantis shrimps (Achuthan Kutty and Parulekar, 1984; Ajith kumar, 1990; Tanuja, 1996; Pedrazzoli et al., 1998).

According to the study of Sriraman (1978), the protein content of crustaceans and mollusks were around 20%. In the present investigation, the protein content of L. vannamei showed 35.69%. Carbohydrate content exhibited an inverse relationship with protein content. Similar findings were recorded by Silva and Chamul (2000), Sriraman (1978), Nair and Prabhu (1990), Reddy and Shanbhogue (1994), Ravichandran (2000). In general, lipid act as major food reserves along with protein and subjected to periodic fluctuations influenced by environmental variables like temperature (Johnstene, 1917; Pillay and Nair, 1973). But this does not affect the lipid composition of muscle tissue to any great extent. In this investigation, the proportion of protein was greater followed by lipid and carbohydrate in the muscle of L. vannamei. Similar difference has been already reported by (Nargis, 2006).

Moisture of fresh shrimp is generally reported as 75 to 80% (Yanar and Celik, 2006; Sambhu and Jayaprakash, 1994). In the present study, 76.2% moisture was recorded. The ash content of the *L. vannamei* was calculated as 1.2% in the present study. Ash content of shrimp is generally 1 to 1.5%. Gokoglu et al. (2008) and Yanar and Celik (2006) calculated the amount of ash in black tiger and white shrimps were 0.95 and 1.47%, respectively. These values are very close to the findings of the present study

Determination of mineral composition of shrimp is important for both checking raw material quality and labeling requirement in nutritional point of view hence aiding health. The main functions of essential minerals are to maintain colloidal systems and a-acid-base equilibrium. Fish and shellfish contain considerable amounts of minerals such as, calcium, magnesium, phosphorus, potassium and sodium (Attar et al., 1992; Abdullah et al., 2009).

The mineral content, including Ca, Mg, K, P, Mn, Na and Fe in the edible part of *L. vannamei* were investigated in the present study. Calcium levels of

Saturated fatty acids	Position of the carbon atom	%
Palmitic acid	C 16	7.06
Margaric acid	C 17	1.42
Stearic acid	C 18	12.88
	Total	23.36
Monounsaturated fatty acids		
Oleic acid	18:1 (n-9)	12.48
	Total	12.48
Polyunsaturated fatty acids		
Omega-6 fatty acids		
Linoleic Acid	18:2 (n-6)	16.3
Omega-3 fatty acids		
Alpha-linolenic acid (ALA)	18:3 (n-3)	11.2
Stearidonic acid (SDA)	18:4 (n−3)	in trace
Eicosatrienoic acid (ETE)	20:3 (n-3)	4
Eicosapentaenoic acid (EPA)	20:5 (n-3)	9
Docosahexaenoic acid (DHA)	22:6 (n-3)	11.2
	Total	51.7

Table 4. Saturated fatty acid and unsaturated fatty acids in the flesh of *L. vannamei*.

L. vannamei were found to be 154.5 mg, which is higher (59.5 mg) than green tiger shrimp (Sambhu and Jayaprakash, 1994), sea bass (63.6 mg) and sea bream (19.2 mg) (Erkan and Ozden, 2007). Calcium is essential for hard tissue structure, blood clotting, muscle contraction, nerve transmission and osmoregulation as a cofactor for enzymatic procession. Calcium recommendation is (RDA) 800 mg/day. In addition, many Ca supplements contain lead, which impairs health in numerous ways. Fortunately, Ca interferes with the absorption and action of lead in the body system (Whithney and Rolfes, 2008). Sodium is the principal caution of the extra cellular fluid, aids acid-base balance and is essential for nervous system (Whithney and Rolfes, 2008). The level of Na in flesh of L. vannamei was found as 67.7 mg. Potassium assists in maintaining fluid, electrolyte balance and cell integrity. During nerve transmission and muscle contraction, potassium and calcium briefly exchange places across the cell membrane. Potassium requirement for human is about 2 g day⁻¹. The average K contents of L. vannamei were found to be 56.7 mg, which is lesser compared to that reported by Sambhu and Jayaprakash (1994) for green tiger shrimp and Erkan and Ozden (2007) for sea bass and sea bream.

Magnesium content of *L. vannamei* was about 13.41 mg. Magnesium has some useful roles in the body. Magnesium is required for the body's enzyme system, bone health; it is a major part of protein synthesis in soft tissues and energy metabolism (Whithney and Rolfes, 2008). Iron is an essential trace element since. It serves as a carrier of oxygen to tissues from the lungs by red

blood cell. Exceeding level of 10 mg kg⁻¹ for iron is not permitted (ITS, 2000). Iron content of shrimps used for this study was 4.54 mg. However, the value stated for L. vannamei was higher than those stated by ITS (2000). Compared to green tiger shrimp (1.48 mg), speckled shrimp (1.55 mg) (Gokoglu et al., 2008, Sambhu and Jayaprakash 1994) and Aristeus antennatus (0.9 mg) (Karakoltsidis et al., 1995). Amino acids are the building blocks of proteins (Babsky et al., 1989). Crustacean muscles contain high concentration of free amino acids, such as arginine, glycine, proline, glutamine and alanine (Cobb et al., 1975), and support osmoregulatory functions (Fang et al., 1992). The amino acid, tryptophan plays an important role in the brain as a precursor of the neurotransmitter: Serotonin, which has a major effect on the feeding behavior of animals (Mullen and Martin, 1992). Valine is involved in many metabolic pathways, protein synthesis and optimal growth (Wilson, 2002). Histidine is also involved in many metabolic functions including the production of histamines, which take part in inflammatory allergic and reactions. lt aids osmoregulation and metabolic pathways during certain emergencies/ harsh conditions (Abe and Ohmama, 1987).

In the present study among saturated fatty acids stearic acid was dominant that followed by palmitic and margaric acids. The amount of palmitic acid in *L. vannamei* was 7.06%. However, Yanar and Celik (2006) reported 22.2% in black tiger shrimp and 21.8% in white tiger shrimp respectively. Sargent et al. (1999) showed reports of 17.3% in golden shrimp, 18.0% in pink shrimp and 17.6% in Norway shrimp. It has been reported that incorporation of essential fatty acids in the diet produced better growth rate and survival in aquaculture. Bell and Sargent (2003), Osborn and Akoh (2002) and Watanabe et al. (1989) mentioned in their review article that n-9 fatty acids, found as oleic acids (C18:1 n-9) plays a moderate role in the body. Moreover, n-6 fatty acids cannot be synthesized by humans and are therefore considered as essential fatty acids.

In the present study, the presence of n-3 PUFA, particularly, linoleic, EPA and DHA indicates better growth and survival of L. vannamei in the culture pond. The higher levels of EPA and DHA would increase stress tolerance and membrane permeability (Watanbe, 1993; Lin et al., 2003). Findings of C18:1 n-9 of L. vannamei species in this study were similar to previously reported findings of white and black tiger shrimps (Yanar and Celik, 2006). In addition, DHA and EPA, belonging to n-3 fatty acids family, are considered as essential (Feliz et al., 2002). DHA and EPA, two of the major PUFA were found as 11.2 and 9% in L. vannamei. Results of this present study also showed that the value of EPA was lower than that of DHA in L.vannamei although Ackman (1989) reported that, the shellfish tend to have EPA greater than DHA. This previous findings disagrees with the present findings of L. vannamei. Furthermore, n-3 fatty acids are essential in growth and development throughout the human life cycle and should be included in the diet.

The n-3 fatty acids have anti-inflammatory and anticoagulant properties as well as many other important health .The ω -9 fatty acids help to reduce the risk of arteriosclerosis, cardiovascular disease and stroke (Since *L. vannamei* contains considerable amounts of PUFA). Based on the results, *L. vannamei* species can be considered as a good source of fatty acid as well as protein.

REFERENCES

- Abdullah O, Ayse O, Mevlut A, Gozde G, Jelena M (2009). A comparative study on proximate, mineral and fatty acid compositions of deep seawater rose shrimp (Parapenaeus longirostris, Lucas, 1846) and red shrimp (Plesionika martia, A. Milne-Edwards, 1883). J. Anim. Vet. Adv. 8(1):183-189.
- Abe H, Ohmama S (1987). Effect of starvation, and seawater acclimation on the concentration and free L-histidine and related dipeptides in the muscle of eel, rainbow trout and Japanese dace. Comp. Biochem. Physiol. 88B:507-511.
- Ackman RG (1989). Nutritional composition of fats in seafood. Progress Food Nutr. Sci. 13:161-241.
- Achuthan Kutty CT, Parulekar AH (1984). Mahasagar-Bul, Natn. Inst. Oceanogr. 17(4):239-242.
- Ajith kumar M (1990). Studies on the proximate composition of the prawn Macrobrachium idella (Hilgendorf). M. phil Thesis, Annamalai University. pp. 1-28.
- Attar KM, El-Fair MZ, Rawdeh TN,Tawabini BS (1992). Levels of arsenic in fish from the Arabian Gulf. Mar. Pollut. Bull. 24:94-97.
- A.O.A.C. (1990). Official methods of analysis 15. Edition. Washington DC. pp. 222-245.
- Babsky EB, Khodorov BI, Kositsky GI, Zubkov AA (1989). In Babsky, E. B. (Ed.), Human Physiology, Mir Publishers, Moscow.

- Bell JG, Sargent JR (2003). Arachidonic acid in aquaculture feeds: Current status and future opportunities. Aquaculture 218:491-499.
- Cobb BF, Conte FS, Edwards MA (1975). Free amino acids and osmoregulation in penaeid shrimp. J. Agric. Food Chem. 23:1172-1174.
- Conner WE, Neuringer M, Reisbick S (1992). Essential fatty acids: The importance of n-3 fatty acids in the retina and brain. Nutr. Rev. 50:21-29.
- Davis DA, Lawrence AL, Gatlin DM (1992). Mineral requirement of *Penaeus vannamei*: A preliminary examination of the dietary essentiality for thirteen minerals. J. World. Agric. Soc. 23:8-11.
- Deshimaru O, Yone Y (1978). Requirement of prawn for dietary minerals. Bull. Jpn. Soc. Sci. Fish. 44: 907-910.
- Dubois M, Giles KA, Hamilton JK, Rebors PA, Smith F (1956). Calorimetric method for determination of sugar and related substances. Anal. Chem. 28:350-356.
- Erkan N, Ozden O (2007). Proximate composition and mineral contents in aqua cultured sea bass (*Dicantrarchus labrax*), sea bream (*Sparus aurata*) analyzed by ICP-MS. Food Chem. 102:721-725.
- Fang LS, Tang CK, Lee DL, Chen IM (1992). Free amino acid composition in muscle and hemolymph of the prawn *Penaeus monodon* in different salinities. Nippon Suisan Gakkaishi 58:1095-1102.
- Folch J, Lee M, Sloane-Stanley GH (1956). A Simple method for the isolation and purification of total lipids from animal tissues. J. Biol. Chem. 226:497-509.
- Feliz GLA, Gatlin MD, Lawrence LA, Velazquez PM (2002). Effect of dietary phospholipid on essential fatty acid requirements and tissue lipid composition of *Litopenaeus vannamei* juveniles. Aquacult. 207:151-167.
- Gokoglu N, Yerlikaya P, Gokoglu M (2008). Mini-review. Trace elements in edible tissue of three shrimp species (*Penaeus semisulcatus, Parapenaeus longirostris* and *Paleomon serratus*). J. Sci. Food Agric. 88:175-178.
- ITS (2000). The Ministry of the Agriculture of Turkey, Rep. 5.
- Johnstene J (1917). The dietic value of hearing. Rep. Laucas Sea Fish. Lab. pp. 32-85.
- Kashiwagi H, Schwartz MA, EigenthalerM, Davis KA, Ginsberg MH, Shattil SJ (1997). Affinity modulation of platelet integrin alphallbbeta3 by beta3-endonexin, a selective binding partner of the beta3 integrin cytoplasmic tail. J. Cell Biol.137:1433-1443.
- Karakoltsidis PA, Zotos A, Constantinides SM (1995).Composition of the commercially important mediterranean finfish: Crustaceans and molluscs. J. Food Comp. Anal. P. 8.
- Kanazawa A, Teshima S, Sasaki M (1984). Requirements of potassium, copper, manganese and iron. Mem. Faculty Fish. Kagoshima Univ. 33:63-71.
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ (1951). Protein measurement with the tolin phenol reagent. J. Biol. Chem. 193:265-273.
- Lin RY, Huang LS, Huang HC (2003). Characteristric of NADH-depend lipid peroxidation in sarcoplasmic reticulum of white shrimp, *Litopenaeus vannamei* and freshwater prawn, *Macrobrachium rosenbergii*. Comp. Biochem. Physiol. 135:683-687.
- Mullen BJ, Martin RJ (1992). The effect of dietary fat on diet selection may involve central serotonin. Am. J. Physiol. Regul. Integr. Comp. Physiol. 263: 559-563.
- Nair ÅL, Prabhu PV (1990). Protein concentrates from tiny prawns. J. Mar. boil. Assoc. India 32(1-2):198-200.
- Nargis R (2006). Seasonal variation in the chemical composition of body flesh of Koi fish Anabas testudineus (Block) (Anabantidae, Perciformes). Bangladesh J. Sci. Ind. Res. 41:219-226.
- Osborn HT, Akoh CC (2002). Structured lipids-novel fats with medical, nutraceutical and food applications. Comp. Rev. Food Sci. 3:93-103.
- Pedrazzoli A, Molina C, Montoya N, Townsend S, Leon- Hing A, Parades Y, Calderon J (1998). Recent Advances on Nutrition Research of *Penaeus vannamei* in Ecuador. Rev. Fish. Sci. 6:1-2.
- Pillay KK, Nair NB (1973). Observation on the biochemical changes in the gonads and other organs of *Uca annulipes*, *Portunus pelagicus* and *Metapenaes affinis* during reproductive cycles. Mar. Biol. 18:167-198.

- Ravichandran R (2000). Biodiversity, Litter processing, Leaf preference and growth, biochemical and microbial aspects in crabs of Pichavaram mangroves. Ph.D. Thesis, Annamalai University, India.
- Reddy HRV, Shanbhogue SL (1994). Biochemical changes in different tissues of the mantis shrimp, *Oratosq uilla neppa* (*Stomato poda*) during reproductive cycle. Indian J. Mar. Sci. 23:247-249.
- Sambhu C, Jayaprakash V (1994). Effect of hormones on growth, Food conversion and proximate composition of the white prawn, Penaeus indicus (Milne Edwards). Indian J. Mar.Sci. 23:232-235.
- Sargent J, McEvoy L, Estevez A, Bell G, Bell M, Henderson J, Tocher D (1999). Lipid nutrition of marine fish during early development: current status and future directions. Aquaculture 179: 217-229.
- Silva JJ, Chamul RS (2000). Composition of marine and freshwater Finfish and shellfish species and their products. In: Maritin, R.E., E.J. Carter and L.M. Davis (Eds.), Marine and freshwater product handbook, USA: Technomic Publishing Company, Inc. pp. 31-46.
- Sriraman K (1978). Biological and biochemical studies on the prawns of Portonova coast (Crustacea: Decapoda: Macrura). Ph.D. Thesis, Annamalai University, India. P. 69.
- Sriket S, Benjakul P, Visessanguan W, Kijroongrojana K (2007). Comparative studies on chemical composition and thermal properties of black tiger shrimp (*Penaeus monodon*) and white shrimp (*Penaeus vannamei*) meats. Food Chem. 103:1199-1207.
- Tanuja R (1996). Some aspects of biology and utilization of the mantis shrimp Oratosquilla neppa from Cochin waters. Ph.D. Thesis, Cochin University of Science and technology, India. P. 87.

- Watanabe T, Arakawa T, Takeuchi T, Satoh S (1989). Comparison between eico sapentaenoic and docosahexaenoic acids in terms of essential fatty acid efficiency in juvenile striped jack *Pseudocaranx dentex*. Nippon Suisan Gakkasihi, 55:1989-1995.
- Watanabe T (1993). Importance of docosahexaenoic acid in marine larval fish. J. World Aquacult. Soc. 24:152-161.
- Whithney R (2008). Understanding Nutrition (11th Edn. For international student adition), USA. P. 410.
- Wilson (2002). Amino acids and Protein. In J. E. Halver and R.W. Hardy (Eds.), Fish Nutrition, Academic Press, San Diego. CA, USA. pp. 143-179.
- Yanar Y, Celik M (2006). Seasonal amino acid profiles and mineral content of green tiger shrimp (*Penaus semisulcatus*, De Haan, 1844) and speckled shrimp (*Metapenaus monoceros*, Fabricus 1789) from the Eastern Mediterranean sea. Food Chem. 94:33-36.