

**REGIONAL HISTOMORPHOMETRY AND DISTRIBUTION OF ANDROGEN RECEPTOR  
IN THE COMMON CAROTID ARTERIES AMONG KENYANS**

Thesis submitted in partial fulfilment of the requirements for the degree of Master of Science  
in Human Anatomy of the University of Nairobi

By

Dr. Kevin Wangwe Ongeti

MAY 2012

## TABLE OF CONTENTS

TABLE OF CONTENTS.....	i
DECLARATIONS.....	iv
DEDICATION.....	v
ACKNOWLEDGEMENTS.....	vi
ABBREVIATIONS .....	vii
LIST OF FIGURES .....	viii
LIST OF TABLES.....	ix
SUMMARY .....	x
CHAPTER 1: INTRODUCTION AND LITERATURE REVIEW .....	1
1.1 Introduction.....	1
1.2 Literature Review.....	4
1.2.1 Carotid Intimomedial thickness .....	4
1.2.2 Age and gender related changes in carotid histomorphometry.....	4
1.2.3 Androgens and androgen receptors in the carotid artery.....	5
1.3 Justification.....	6
1.4 Significance of the study.....	7
1.5 Study Question.....	8
1.6 Hypothesis.....	9
1.7 Objectives .....	9
1.7.1 Broad objective .....	9
1.7.2 Specific Objectives .....	9
CHAPTER TWO: MATERIALS STUDY METHODS.....	10
2.1 Study design and setting .....	10
2.2 Ethical considerations in the study.....	10
2.3 Sample size and distribution .....	10

2.3.1	Inclusion Criteria.....	11
2.3.2	Exclusion Criteria .....	11
2.4	Controls.....	11
a.	Positive control .....	12
b.	Negative control.....	12
2.5	Accessing the common carotid artery .....	12
2.6	Histological methods used .....	14
2.6.1	Tissue processing for light microscopy.....	14
2.6.2	Staining Techniques.....	14
2.7	Immunohistochemistry technique.....	15
2.8	Morphometry .....	16
2.8.1	Intimal thickness .....	17
2.8.2	Medial thickness .....	18
2.8.3	Intimal-medial thickness .....	18
2.8	Inter observer Error.....	18
2.9	Data handling and management.....	19
CHAPTER THREE: RESULTS .....		20
3.1	Histomorphometry of the common carotid artery.....	27
3.2	Age and gender differences in the structure of the common carotid artery .....	28
3.3	Regional differences in the structure of the common carotid artery .....	29
3.4	Androgen receptor distribution in the common carotid artery .....	33
CHAPTER FOUR: DISCUSSION AND CONCLUSIONS.....		42
4.1	Histomorphology of the Human Common Carotid Artery .....	42
4.2	Histomorphometry of the Common carotid artery.....	45
4.3	Segmental differences in the histomorphometry of the common carotid artery .....	47
4.4	Age and gender differences in structure and morphometry of the common carotid artery... 48	
4.4.1:	Age differences in structure and morphometry of the common carotid artery .....	48

4.4.2: Gender differences in structure and morphometry of the common carotid artery .....	48
4.5 Androgen receptor expression in the common carotid artery .....	49
Conclusions.....	51
REFERENCES .....	52
APPENDIX 1.....	76
CONSENT FORM.....	76
CHETI CHA RUHUSA.....	77

DECLARATIONS

I hereby declare that this thesis is my original work and has not been presented elsewhere for approval and examination.

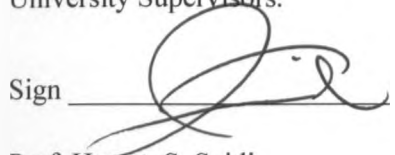
Sign 

Date 5<sup>th</sup> July, 2012

Dr. Kevin W. Ongeti (candidate)

B.Sc. Anat (Hons), M.B.Ch.B.

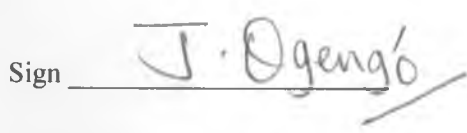
University Supervisors.

Sign 

Date 13/7/12

Prof. Hassan S. Saidi

B.Sc. Anat (Hons), M.B.Ch.B., M.Med. (Surg), FCS (ECSA), FACS.

Sign 

Date 18/7/2012

Prof. Julius A. Ogengó

B.Sc. Anat (Hons), M.B.Ch.B., PhD.

## DEDICATION

To my wife Catherine Nyokabi

Daughter Casey Namukonga

To my mum Catherine Khasiala

To my friends and colleagues

## ACKNOWLEDGEMENTS

I acknowledge my supervisors and mentors, Prof. Julius Ogeng'o and Prof. Hassan Saidi for their continued support and inspiration from conceptualisation and proposal to realisation of this thesis. I am sincerely grateful to Prof. Julius Ogeng'o for his close interest in my work, regular supervision and inculcating in me the virtues of dedication and perseverance. I am greatly indebted to Prof. Hassan Saidi for his concise critical appraisal, insisting on focus, his contribution, ideas and incredible support throughout this subject.

I also acknowledge the support and contributions of the members of staff of the Department of Human Anatomy, University of Nairobi especially Dr. Pamela Mandela, Dr. Peter Gichangi, Dr. Moses Obimbo, Dr. Anne Pulei, Dr. Wycliff Kaisha and Dr. Kirsten Awori in the development of this thesis.

I acknowledge the assistance offered by Mr Acleus Murunga in specimen collection, Ms Esther Mburu, Mr Martin Inyimili and Mr Samuel Kasera in histology processing of the tissues. I also acknowledge the support offered by the Pathologist Lancet Laboratory in immunohistochemistry and the Chiromo Funeral Parlour for allowing me to collect the post-mortem specimen.

Great thanks go to my family members for being understanding and supportive. My thanks and appreciations go to the families who selflessly donated the arteries of their beloved to this study. I am grateful to the University of Nairobi for the institutional support and fee waiver. Finally I wish to thank the other people not mentioned above, who directly or indirectly contributed to the success of this thesis.

## ABBREVIATIONS

AR.....	Androgen receptor
AR+.....	Androgen receptor positive cells
CA.....	Carotid Artery
CIMT.....	Carotid Intimomedial thickness
CVD.....	Cardiovascular Disease
EC.....	Endothelial cells
EEL.....	External elastic lamella
IEL.....	Internal elastic lamella
IgG.....	Immunoglobulin Gamma
IMT.....	Intimomedial Thickness.
Is.....	Intimal segment
KNH.....	Kenyatta National Hospital
Ms.....	Medial segment
MT.....	Masons Trichrome
SPSS.....	Statistical Program for social sciences
TCC.....	Total cell count
VSMC.....	Vascular Smooth Muscle cells
WE.....	Weigert Elastin



## LIST OF FIGURES

Figure 1: Reflected skin to expose the CCA.....	13
Figure 2: Scion Image analyser .....	16
Figure 3: Segments of the vessel wall that were measured.....	17
Figure 4: Photomicrograph of the general structure and tunica intima of the CCA.....	23
Figure 5: Photomicrograph of the tunica media of the common carotid artery.....	25
Figure 6: Photomicrograph of the adventitia of the common carotid artery.....	27
Figure 7: Intimomedial thickness in different age groups.....	28
Figure 8: A photomicrograph showing gender differences in the tunica media .....	31
Figure 9: A photomicrograph showing regional differences in the structure of the tunica adventitia .....	35
Figure 10: A photomicrograph showing the age and gender differences the carotid tunica intima.....	37
Figure 11: A photomicrograph showing the non-expression of androgen receptors in the carotid arterial wall.....	39

## LIST OF TABLES

Table 1: Carotid intimomedial thickness across the segments, age and gender groups.....	32
Table 2: Distribution of androgen receptors in the common carotid artery.....	33
Table 3: Carotid Intimomedial thickness in various populations.....	40
Table 4: Age differences in the carotid intimomedial thickness.....	40
Table 5: Gender differences in the carotid intimomedial thickness.....	41

## SUMMARY

**Background:** The carotid intimomedial thickness is a predictor of atherosclerosis in other vascular beds and is useful in cardiovascular risk stratification. Atherosclerosis is reported to be worse in males and seems to increase with age. There have been reports that androgens are protective while other reports suggest they are bane. Expression distribution of androgen receptors in the carotid intimomedial thickness may help explain the role of androgens in development and progression of atherosclerosis. The relationship between the androgen receptors and the carotid intimomedial thickness is unknown.

**Hypothesis:** There are segmental, age and gender differences in the carotid intimomedial thickness which are influenced by presence of androgen receptors.

**Objectives:** To describe the distribution of androgen receptors in the common carotid artery intima and media and relate them with the carotid intimomedial thickness among Kenyans.

**Study design:** Descriptive crosssectional study

**Setting:** Department of Human Anatomy, University of Nairobi.

**Materials and Methods:** Materials for this study were obtained from twelve (6 males and 6 females) common carotid arteries during autopsy within 48 hours of demise at the Chiromo funeral parlour. Three millimetre samples from the proximal, middle and distal thirds of artery segments were collected. They were fixed in 10% formaldehyde solution immediately and routinely processed for light microscopy. Another set of slides were stained using anti-androgen receptor monoclonal antibody (AR 318). All prostate samples used as positive controls were immunopositive for androgen receptors. The total cell count and the number of stained cells for each designated vessel zone were assessed in the three sections in four visual fields per section at 400 x magnification using a Zeiss® photomicroscope. The carotid intimomedial thickness was measured. The presence and distribution of androgen receptors

were observed and described. The intimomedial thickness was measured using the Scion Image Multiscan® software. The data collected was analysed using SPSS® version 18 for Windows® for means and variances.

**Results:** The mean age of the cases was  $28 \pm 19$  yrs. Mean carotid intimomedial thickness was  $0.86 \pm 0.22$  mm (Male:  $0.97 \pm 0.22$ ; Females  $0.77 \pm 0.06$ ),  $p=0.05$ . Mean proximal, middle and distal intimomedial thickness were  $0.86 \pm 0.26$  mm,  $0.84 \pm 0.28$  mm and  $0.90 \pm 0.35$  mm respectively. Carotid intimomedial thickness increased with age;  $0.5 \pm 0.16$  mm,  $0.87 \pm 0.24$  mm and  $1.21 \pm 0.36$  mm for the age groups 0-20 yrs, 21-40 yrs and 41-60 yrs respectively ( $p=0.035$ ). Androgen receptors were not detected in the 12 samples tested.

**Conclusions:**

The common carotid artery is an elastic artery with a well-developed tunica intima and does not display androgen receptors. Carotid intimomedial thickness increases distally, with age and is higher in males. The carotid intimomedial thickness is not influenced by the presence of androgen receptors.

## CHAPTER 1: INTRODUCTION AND LITERATURE REVIEW

### 1.1 Introduction

The common carotid artery (CCA) originates from the brachiocephalic trunk and aortic arch on the right and left respectively (Sinnathamby, 1999; Standring et al., 2004). This artery supplies the brain, extracranial structures in the head, and neck region and it can be divided arbitrarily into three parts: proximal, middle and distal (Watson and Silverstone, 1939; Skandalakis et al., 2004). The proximal region extends from the origin to the sternoclavicular joint, while the middle section is located in the neck between the sternoclavicular joint and the 2<sup>nd</sup> cervical vertebra. The distal part extends from the 2<sup>nd</sup> cervical vertebra to the carotid bifurcation at C3/4. It is a common site of atherosclerosis (Bo et al., 1989). Further, it increases with age and it is more common in males than premenopausal females (Hansen et al., 1995; Samijob et al., 1998). The reason for this regional gender and age preference are largely unclear. Intrinsic vascular factors that may explain these differences remain largely underreported. These include carotid intimomedial thickness and expression of androgen receptors (Tell et al., 1989).

The structure of the mammalian CCA varies according to species some revealing a muscular media (Kimani, 1983; Gabella et al., 2005), while others reveal an elastic structure (Pinto et al., 1998; Carallo et al., 1999; Orsi et al., 2006; Parchami et al., 2009; Parchami and Dehkordi, 2011). There are segmental differences in structure of the sheep, goat and giraffe carotid artery (Kimani, 1983; Parchami et al., 2009). These regional changes are influenced by the hemodynamic differences due to the length of the neck and body posture (Kimani, 1983). All artery tunics show age related changes in structure (Toda et al., 1980). These include intimal thickening, fragmentation of the internal elastic lamina, elastic tissue

fragmentation, collagen deposition in the tunica media (Futterman and Lemberg, 2003; Lee and Oh, 2010; Kotsis et al., 2011). Although these changes occur in virtually every artery, there are differences in the age of onset and degree of change experienced by different arteries (Toda et al., 1980). Generally, males have a thick intima and media and usually age faster than females (Lakatta and Levy, 2003). These gender differences in the vessel structure have been related to the effects of androgens (Sader and Celermajer, 2002).

The exact role of androgens in development and progression of atherosclerosis is controversial; with some studies showing protection (Bonnel et al., 1941; Alexandersen et al., 1999) while others demonstrate detriment (Liu et al., 2001; Lawlor et al., 2004; Ramirez et al., 2007). Effects of androgens are mediated through androgen receptors (AR) (Liu et al., 2003). These receptors have been demonstrated in cultured vascular macrophages, endothelial cells (EC), fibroblasts and smooth muscle cells (Kimura et al., 1993; Liu et al., 2005; Rexrode et al., 2008; Sieveking et al., 2010), and show gender dichotomy in expression. Males exhibit a four-fold higher expression AR than females (McCrohon et al., 2000; Ng et al., 2003).

Post-mortem studies have elucidated different numbers of AR in the coronary arterial wall, inversely relating them to early atherosclerosis (Liu et al., 2005). Relating the distribution and number of androgen receptors to the carotid structure and CIMT would give some insight into the role of androgens in the gender differences in development and progression of atherosclerosis. The pattern of expression of androgen receptors in the carotid arterial wall is unknown. Expression of androgen receptors if related to the vessel segment, age and gender may explain the gender and regional differences in atherogenesis (McGill and Sheridan, 1981; McGrath et al., 2008). This study therefore describes the segmental structure,

histomorphometry and androgen receptor expression in the carotid artery in males and females of different age groups.

## 1.2 Literature Review

The common carotid artery is a common site of atherosclerosis. This preponderance is related to the changes in its intimomedial thickness (Sader et al., 2001; Liu et al., 2003). Furthermore, the intimomedial thickness may show age, gender, segmental differences related to the distribution of androgen receptors in the different mural layers. Mammalian CCA wall is composed of three histological layers namely the tunica intima, tunica media and tunica adventitia (Kimani, 1983; Parchami et al., 2009). The tunica intima is composed of a single layer of endothelial cells and the subendothelial zone (SEZ), a region just below the endothelial surface but on the adluminal side of the IEL (Timmins et al., 2010). The tunica media in some mammals is preponderantly elastic, with concentric elastic lamellae and interposed smooth muscle cells and collagen (Orsi et al., 2006; Parchami et al., 2009). Other mammals have muscular CCA tunica media (Kimani, 1983). Arterial adventitia on the other hand is fibroelastic (Wang et al., 2010).

### 1.2.1 Carotid Intimomedial thickness

Although CIMT is not yet routinely measured in clinical practice, its predictive value regarding cardiovascular complications has been established, giving it a potential role in future for cardiovascular disease risk stratification and primary prevention (Hodis et al., 1998; Greenland et al., 2000; Jarauta et al., 2010). The human carotid intimomedial thickness is about 0.8 to 0.91mm, measured using ultrasound (Jadhav and Kadam, 2001; Okeahialam et al., 2011). It is 1.049mm and 0.55mm in Americans and Nepalese respectively (Crouse et al., 1995; Sharma et al., 2009).

### 1.2.2 Age and gender related changes in carotid histomorphometry.

Aging changes in muscular arteries include; intimal thickening, elastic tissue fragmentation, and smooth muscle hyperplasia (Roach, 1970; Nakagami and Morishita, 2008). The internal



elastic lamina (IEL) becomes less organized with aging (Gross et al., 1934). Notable changes in the tunica media include increased collagen and smooth muscle deposition, thickening and subsequent stiffness (Cameron et al., 2003). The external elastic lamina (EEL) thickens with age (Kawasaki et al., 1987). The tunica adventitia ages by collagen deposition and elastic tissue loss (Gutterman, 1999; Wang et al., 2010). These changes occur earlier in males than in females before menopause, levelling afterwards (Kannel et al., 1976; Colditz et al., 1987; English et al., 1997; Futterman and Lemberg, 2003). This gender specific age changes have therefore been related to the effects of sex hormones (Juul and Skakkebaek, 2002; Anderson and Pepine, 2007). The age of onset of these changes in the common carotid artery is not described. Segmental intimomedial thickness in the carotid artery, aging changes and the relationship with the number and distribution of vascular AR, in the common carotid artery in both genders is unknown.

### 1.2.3 Androgens and androgen receptors in the carotid artery

The marked sexual dimorphism that exists in IMT and human cardiovascular diseases has led to the concept that androgens have deleterious effects and exacerbates the development of cardiovascular disease in males (Perusquia and Stallone, 2010; Parchami and Dehkordi, 2011). The carotid luminal diameter is wider in females than males; the converse is true for wall thickness (Parchami and Dehkordi, 2011). Androgens have traditionally been regarded as the proximate cause underlying this male disadvantage (Wu and Eckardstein, 2003). Although the CIMT and atherosclerosis increase with age, serum levels of endogenous androgens decrease with age (Bernini et al., 1999; Jarauta et al., 2010). If the effects of androgens persist despite their lower levels in aging, it is probable that there is up regulation of androgen receptors in the elderly. The older individuals with wider CIMT must have more androgen receptors in the carotid mural layers. The role of androgens in the age related

worsening of intimomedial thickness and subsequent development of atherosclerosis is however unknown. Androgen receptors are expressed in the arterial wall of mammals including man (Fujimoto et al., 1994; Hanke et al., 2001; Rexrode et al., 2008; Sieveking et al., 2010). Expression and numbers of these receptors in the coronary artery have a linear correlation with protection from atherosclerosis (Liu et al., 2005). Furthermore, treatment of rats with testosterone is associated with inhibition of neointimal plaque formation and increased expression of the AR mRNA after endothelial injury (Hanke et al., 2001; Li et al., 2008). Androgens have also been shown to stimulate proteoglycan proliferation and elongation of vascular smooth muscles, promoting atherosclerosis (Hashimura et al., 2005). However, other studies (Somjen et al., 1998) have shown that androgens inhibit vascular smooth muscle cell (VSMC) proliferation, thereby protecting against atherosclerosis. Independently, androgens have also been found to up regulate their receptors (Ma et al., 2005). Over expression of AR in endothelial cells confers them with increased androgen sensitivity (Sieveking et al., 2010). Vascular AR may have a cellular or zonal preference and their expression may be related to the histomorphometry of the artery, especially the intimomedial thickness. This information could predict the role these receptors in increased intimomedial thickness and subsequent development of atherosclerosis.

### **1.3 Justification**

The common carotid artery is prone to atherosclerosis and thromboembolic phenomena which are a common cause of stroke and mortality especially in males (Bösel et al., 2010; Hart and Benavente, 1999). When exposed to the same lifestyle, the male predominant atherosclerosis, thromboembolic phenomena, arterial wall stiffness, dissection and aneurysm in the CCA may be related to the carotid intimomedial thickness and effects of sexual hormones (Lerner and Kannel, 1986; Ahimastos et al., 2003). While the benefits of

oestrogens in sparing arteries from atherosclerosis are largely known (Tracy et al., 1966; Sack et al., 1994), controversy still exists as to the role of androgens in increasing CIMT and subsequent development of atherosclerosis (Bernini et al., 1999; McGrath et al., 2008). This knowledge is relevant now in an attempt to resolve the role of androgens in atherogenesis in both genders, as current research is reviewing complementary theories of atherogenesis and management of atherosclerosis.

#### **1.4 Significance of the study**

The common carotid artery is the major source of blood to the brain, head and neck region (Sinnathamby, 1999). It is frequently plagued by atherosclerosis (Duvall and Vorchheimer, 2004). Mural factors predisposing the common carotid artery to atherosclerosis include its carotid intimomedial thickness and androgen receptor distribution (Ramirez et al., 2007; Jarauta et al., 2010). Carotid intimomedial thickness in sheep varies with age and shows segmental and age differences (Parchami and Dehkordi, 2011). The relationship between CIMT and androgen receptor distribution in the human carotid artery is unknown. This study therefore describes the segmental histomorphology and dimensions of the carotid artery in humans. It also relates the expression of androgen receptors to the carotid intimomedial thickness. This information is useful in understanding aging of the carotid arteries, patterns of atherogenesis in both gender and the role of androgens in atherogenesis.

## 1.5 Study Question

Epidemiological studies have shown there is a striking gender difference in cardiovascular disease with men having higher rates of clinical events than women (Kalin and Zumoff, 1990). Gender patterns in the prevalence and onset of carotid atherosclerosis may be related to its histological structure and dimensions (Rosfors et al., 1998). Less is known regarding the relationship between CIMT, an established cardiovascular risk factor and the presence of AR in the carotid artery. Prior to this, no study has been designed to evaluate the morphologic appearance of the non-pathologic human carotid and its relationship to the presence and distribution of androgen receptors.

## **1.6 Hypothesis**

### **Alternate hypothesis**

There are segmental, age and gender differences in the carotid intimomedial thickness which are influenced by presence of androgen receptors.

## **1.7 Objectives**

### **1.7.1 Broad objective**

To describe distribution of androgen receptors in the common carotid artery intima and media and relate them with the carotid intimomedial thickness among Kenyans.

### **1.7.2 Specific Objectives**

With respect to the common carotid artery among Kenyans;

1. To describe the histomorphometry
2. To describe the age and gender differences in histomorphometry.
3. To describe the segmental differences in carotid intimal medial thickness.
4. To demonstrate androgen receptors in different mural layers.

## CHAPTER TWO: MATERIALS STUDY METHODS

### 2.1 Study design and setting

A descriptive cross-sectional study carried out at the Department of Human Anatomy University of Nairobi, between March and June 2011.

### 2.2 Ethical considerations in the study

All protocols were carried out according to a study proposal approved by the UON/KNH-Ethical and Research committee. Relatives of the deceased gave consent for use of the carotid artery samples for this particular study (Appendix 1). The names and identities of the deceased were not used during sample collection. Three millimetre sections from the proximal, middle and distal segments of the common carotid artery were harvested for histology during autopsy. Remnants of the histologic material that were not used in the study were decently buried at Langata cemetery, Nairobi.

### 2.3 Sample size and distribution

The number of samples was calculated according to formula provided by Eng (2003).

$$N = \frac{4\sigma^2 (Z_{crit})^2}{D^2}$$

In the above formula, N was the sample size required. It was assumed that there was a standard deviation ( $\sigma$ ) of 2.5 for each variable in each group, and a minimum expected difference (D) between the means was 2.5 (McRobb et al., 2009). Zcrit of 1.960 based on significance criteria (P-value) of 0.05 was used.

Accordingly;

$$N = 4 \times 2.5 \times 2.5 (1.96)^2$$

$$2.5^2$$

$$N = 12.36 \text{ Approx. } 12$$

Twelve common carotid arteries from 6 males and 6 females were used in this study. The samples were divided into two groups; male and female; in three age groups (0-20, 21- 40 and 41-60 years). Among females, the first two age groups correspond to the reproductive age groups while the last were postmenopausal.

### Selection criteria

#### 2.3.1 Inclusion Criteria

Common carotid artery samples that were used met all listed conditions;

1. Cases whose relatives gave consent to participate in the study.
2. Individuals without gross defects and atherosclerosis.

#### 2.3.2 Exclusion Criteria

The following individuals were excluded from the current study

1. Individuals with major neck trauma damaging the carotid arteries.
2. Patients who died from hypertension or diabetes.

### 2.4 Controls

Control samples were used during the immunohistochemical staining of androgen receptors for quality control. Both positive and negative controls were used.

#### **a. Positive control**

Three post-mortem prostates samples from 3 different males were harvested within 48hrs of demise were used as positive controls for androgen receptor immunohistochemical staining (Bayer-Garner et al., 1999; Rocha et al., 2000).

#### **b. Negative control**

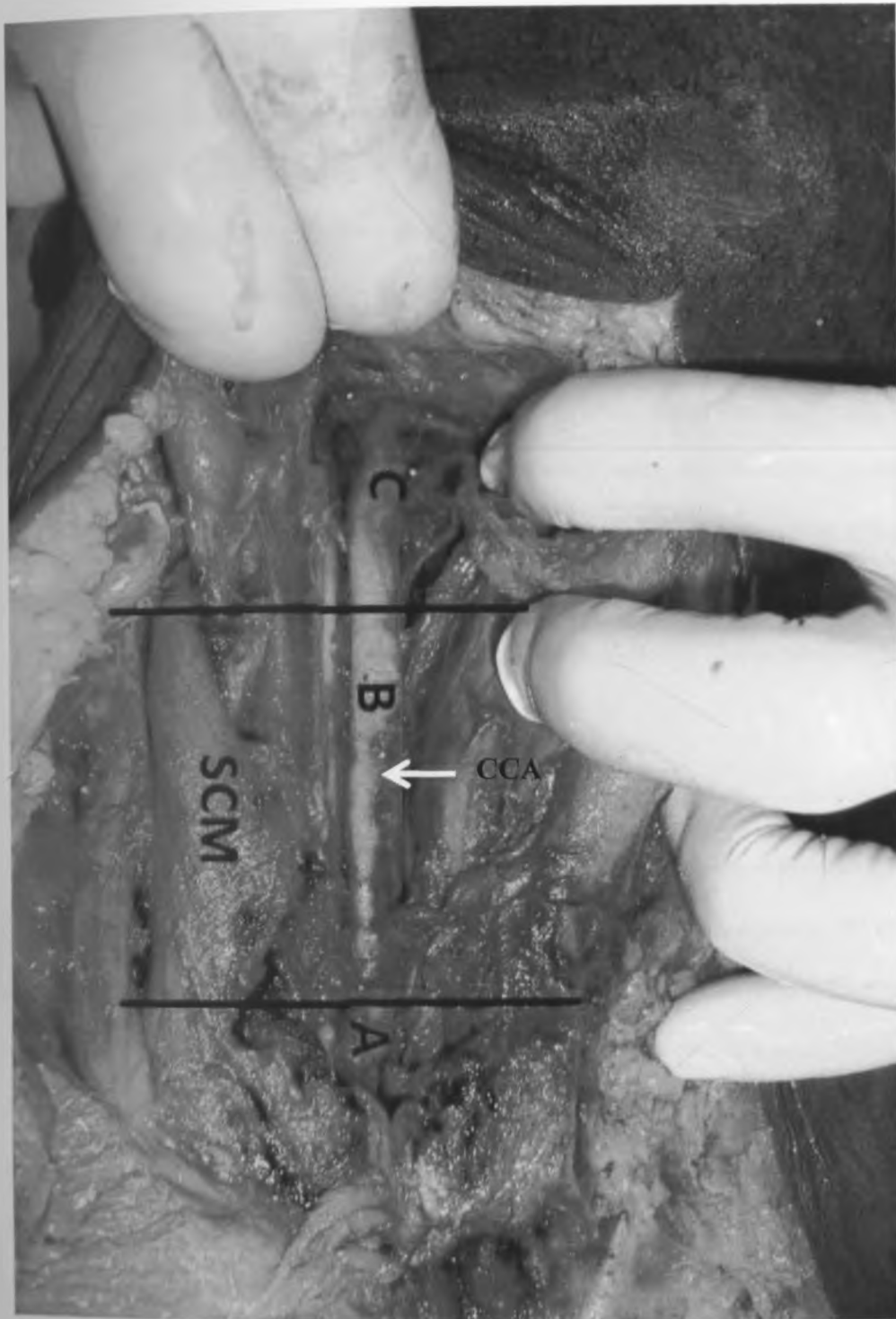
The omission of the primary antibody and substitution with dilution solution alone served as a negative control (Gallardo et al., 2009).

### **2.5 Accessing the common carotid artery**

A skin incision was made in the neck along the impression of the anterior border of sternocleidomastoid (SCM) from the angle of the mandible to the sternoclavicular joint (Figure 1). The skin, investing cervical fascia were separated and retracted exposing the contents of the carotid sheath. The common carotid artery was dissected from the root of the neck to the bifurcation point and divided into three proportional segments; A (Proximal), B (Middle) and C (Distal) (Figure 1). Three millimetre tissue samples of the whole vessel wall were measured and harvested from the proximal, middle and distal segment including the bifurcation in cases less than 72 hours from the time of demise.



Figure 1: Reflected skin to expose the CCA



A macrograph showing the reflected skin, a separated sternocleidomastoid (SCM) muscle exposing the common carotid artery. Arterial segments A, B and C were taken, A (Proximal); B (Middle); C (Terminal).

## 2.6 Histological methods used

### 2.6.1 Tissue processing for light microscopy

The 3mm sections harvested from the three regions described above and 3mm prostatic tissue were used. They were fixed in 10% formal saline for 24 hours, immediately after harvesting and dehydrated in increasing concentrations of ethyl alcohol starting with 70% ethanol up to absolute alcohol for one hour each. Toluene was used as a clearing agent for 1 hour before infiltrating them with paraffin wax for 12 hours at 56<sup>0</sup>c. The tissues were blocked into paraffin waxed blocks. Seven micrometer thick sections were then cut using a Leitz Wetzlar® sledge microtome. They were floated in warm water and thereafter mounted and dried in hot air oven at 40<sup>0</sup>C for 12 hours. Dewaxing was then done using xylene for 5 minutes after which the tissues were rehydrated using ethyl alcohol for 5 minutes and then with decreasing concentrations of alcohol from absolute alcohol to 70% alcohol for 5 minutes each.

### 2.6.2 Staining Techniques

Haematoxylin and Eosin was used to study the general tissue characteristics. The Haematoxylin stains the nucleus while the Eosin counter stains it by staining the cytoplasm and connective tissue with varying intensities (Drury et al., 1967). Masson's Trichrome was used to study the cytoarchitecture and connective tissue of the carotid tunics. Some sections were stained with Weigert's Resorcin Fuschin then counterstained with van Gieson stain to demonstrate the elastic component of the vessels. The slides were examined under Leica® light microscope at magnification x40, x100 and x400. The structure of the tunica intima (TI), media (TM) and adventitia (TA) were noted. The information made was recorded on data sheets.

## 2.7 Immunohistochemistry technique

Paraffin sections 3µm were cut and mounted on previously charged and coded microscope slides. These included a positive control (prostate sample) and the test section (carotid sample). All prostate samples were immunopositive for androgen receptors. The sections were fixed in the bond-max covertiles. Respective labels were then affixed to the slides and the racks slotted into the machine for autostaining. A high amplification, biotin free detection system optimized for use on the bond system was used for staining. Autoimmunostaining occurred as follows; dewaxing was done by a commercially available dewax solution® for 15 minutes. Specimens were incubated with hydrogen peroxide for 5 minutes to quench endogenous peroxidase activity. Antigen retrieval was done by microwaves at 98<sup>o</sup>c for 20 minutes at a Ph of 9.0 (Janssen et al., 1994). They were then incubated with the primary antibody (AR – 318) for 15 minutes then rinsed using bond wash buffer (Sajjad et al., 2004). Post primary IgG linker reagent was applied to the sections for 8 minutes to localize the antibody, followed by a buffer wash. Sections were then incubated with bond polymer for 8 minutes then rinsed using bond wash buffer. Sections were incubated for 10 minutes in 3, 3'-diaminobenzidine tetra hydrochloride (DAB) (Sigma) which forms a brown precipitate with the complex so as to aid visualization. Harris haematoxylin counterstaining was done for 5 minutes to allow visualization of the nuclei (Merck, Poole, Dorset, UK). At the end of the staining the racks were dislodged from the staining chamber and the stained slides fitted into a staining rack. The sections were dehydrated into two changes of alcohol, and cleared in three changes of xylene. The slides were then mounted in DPX. Slides were examined by the principal investigator and a pathologist who were unaware of the gender and ages of the individuals. The total cell count (TCC) and the number of AR stained (AR+) cells for each

designated vessel zone were assessed in the three sections in three visual fields per section at 400 x magnification using a Zeiss® photomicroscope.

## 2.8 Morphometry

Slides were photographed using a Zeiss® digital photomicroscope at magnifications of x40 and x100 for analysis. Every fifth slide from each gender and age group were picked and measurements taken as described below. Analysis of the photographs was done using Scion Image™ Multiscan software (Scion Corporation, Frederick, Maryland) after calibration accurate to 0.01 mm, using ruler measurements of the histological slides [Figure 2] (Nakashima et al., 2002).

Figure 2: Scion Image analyser



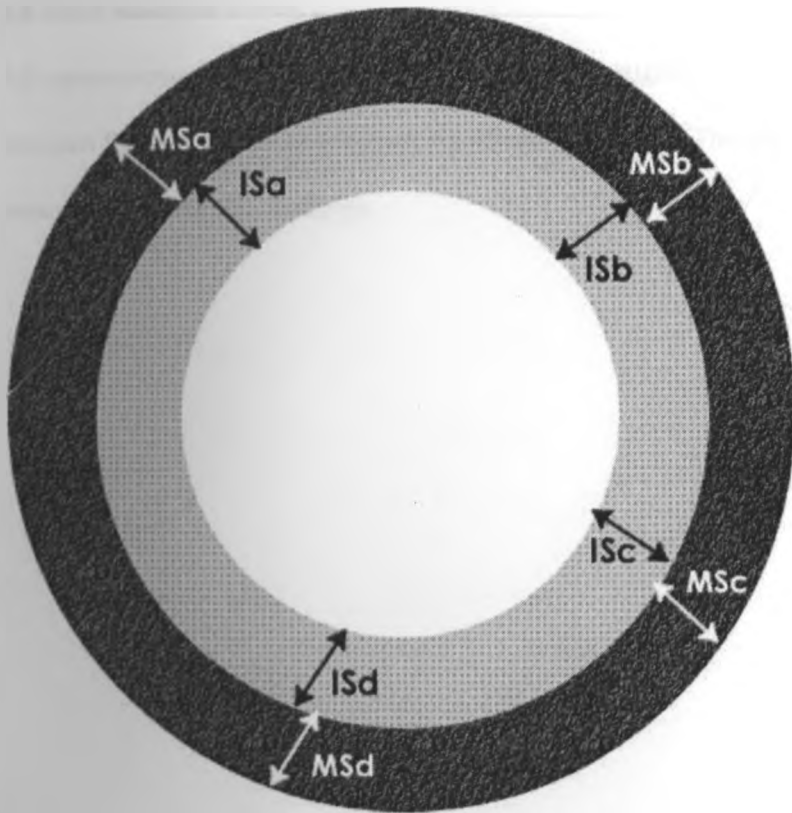
A digital ruler used to measure the various parameters of the CCA. Adopted from Nakashima et al., 2002.

The following parameters were determined;

### 2.8.1 Intimal thickness

Intimal extent was taken as the region extending from the lumen to the internal elastic lamina separating the tunica intima and the tunica media (Fernie and Lamb, 1985). Four random points were selected and the average length was calculated (Figure 3).

Figure 3: Segments of the vessel wall that were measured.



Intimal sections (IS), medial section (MS) of the vessel wall were measured at four random points (a-d).

Adopted from Nakashima et al., 2002.

### **2.8.2 Medial thickness**

Medial extent was taken as the region extending from beneath the thin tunica intima to the elastic fibres separating the medial and adventitial tunics. Four random points were selected and the average length was calculated.

### **2.8.3 Intimal-medial thickness**

Carotid intimomedial thickness was taken as the region covered by both the tunica intima and the tunica media.

### **2.8 Inter observer Error**

All measurements were done by the principal investigator and an assistant. The difference between the two measures were not significant ( $p=0.656$ ). The true measure was taken as the mean between the two readings.

## **2.9 Data handling and management.**

The data collected was tabulated, coded and analysed using a statistical program SPSS® version 18 for Windows® 7 (SPSS Inc. Chicago Illinois, 2010). The intimomedial thickness was determined for the different age and gender groups. Tables and charts were used to illustrate the findings. The F test was used to determine significance. A p value of  $P < 0.05$  was considered significant (Eng, 2003).

## CHAPTER THREE: RESULTS

There were 6 males and 6 females with a mean age of  $28 \pm 19$  yrs ( $28 \pm 20$  yrs for females;  $29 \pm 20$  males). The youngest sample was from a 1 yr old while the oldest was 60 yrs old.

The mean sample collection time after demise was 18 hrs. The common carotid arterial wall comprised of three histological layers namely; tunica intima, tunica media and tunica adventitia (Fig. 4A).

The tunica intima comprises of an endothelium of elongated flattened cells with prominent, darkly stained nuclei, pale cytoplasm and the SEZ (Fig. 4B). The SEZ comprises of loosely arranged collagen interspersed with elastic and spindle shaped oval nucleated cells akin to smooth muscle cells (Fig. 4A-D). Smooth muscle cells were oriented axially (Fig. 5 D). In some places it thickened to form focal intimal thickening (Fig. 4C). A wavy internal elastic lamella (IEL) separated the intima from media (Fig. 4E).

The tunica media was the thickest layer constituting about 80% of the carotid wall (5D). It was predominantly elastic, with 20 to 35 concentric wavy elastic lamella interspersed with collagen and smooth muscle cells (Fig. 5A, Fig 5B). The elastic lamellae decreased to 25-30 in the middle and 20-22 in the distal sections. In some sections the media displayed a sub-intimal zone of smooth muscle cells densely packed in the luminal part of the tunica media (Fig 5C). Smooth muscle cells were arranged in layers; circumferential and longitudinal patterns between the elastic lamellae (5A). Smooth muscle cells increased in the tunica media distally. Collagen fibres were loosely arranged in circular, oblique and transverse directions around the vessel wall (Fig 5C). The amount of collagen fibers in the tunica media increased with age. About eighty percent of the vessel wall volume was occupied by the tunica media.

The tunica adventitia is predominantly fibroelastic, with interspersed smooth muscle cells



(Fig. 6A), in two layers, the inner compact and the outer loose zone (Fig 6B). Zonation of the tunica adventitia was observed in the middle and distal sections of the artery. The elastic fibers in the adventitia were arranged in a circumferential pattern. It also had prominent vasa vasora (vv) and nervi vasora (nv) [Fig. 6C]. The vasa vasora extended into the media and intima in some sections (Fig 10C).

Figure 4: General structure and tunica intima of the CCA

A: A photomicrograph of a section of the CCA showing three layers namely; tunica Intima (TI), Tunica Media (TM) and Tunica Adventitia (TA) (Magnification x40, Masons trichrome stain).

B: A photomicrograph of a section of the CCA showing the tunica intima comprised of squamous endothelial cells (EC) and the SEZ. The SEZ had collagen fibers (starred) and spindle shaped oval nucleated cells akin to SMC (Magnification X400, Masons trichrome).

C: A photomicrograph of a section of the CCA showing Focal intimal thickening (DIT) (Arrowed) in the SEZ. This zone was composed of loosely arranged collagen interspersed with smooth muscle cells (Magnification X40, Masons trichrome).

D: A photomicrograph of a section of the tunica intima of the CCA with intimal hyperplasia (IH), showing a hyperplastic SEZ comprised of loosely arranged elastic fibers and smooth muscles (Arrowed). The smooth muscle cells are oriented longitudinally away from the endothelium.

E: A photomicrograph of a section of the CCA showing a wavy IEL (Black arrow), separating the intima from media. Concentric circumferential elastic lamellae in the tunica intima are also shown by the green arrows (Magnification X400, Weigert Elastin).

Figure 4: Photomicrograph of the general structure and tunica intima of the CCA

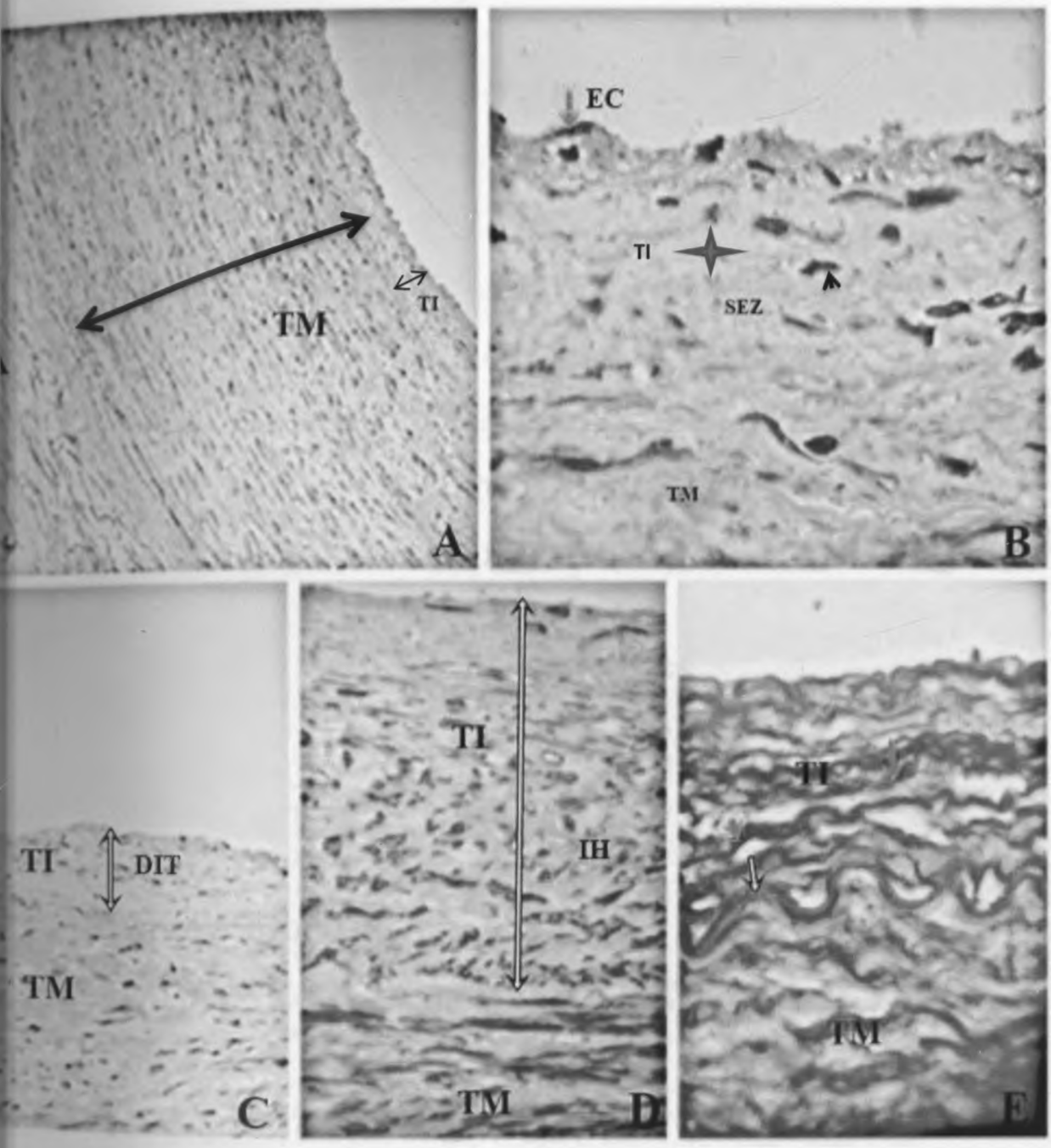


Figure 5: The tunica media of the common carotid artery

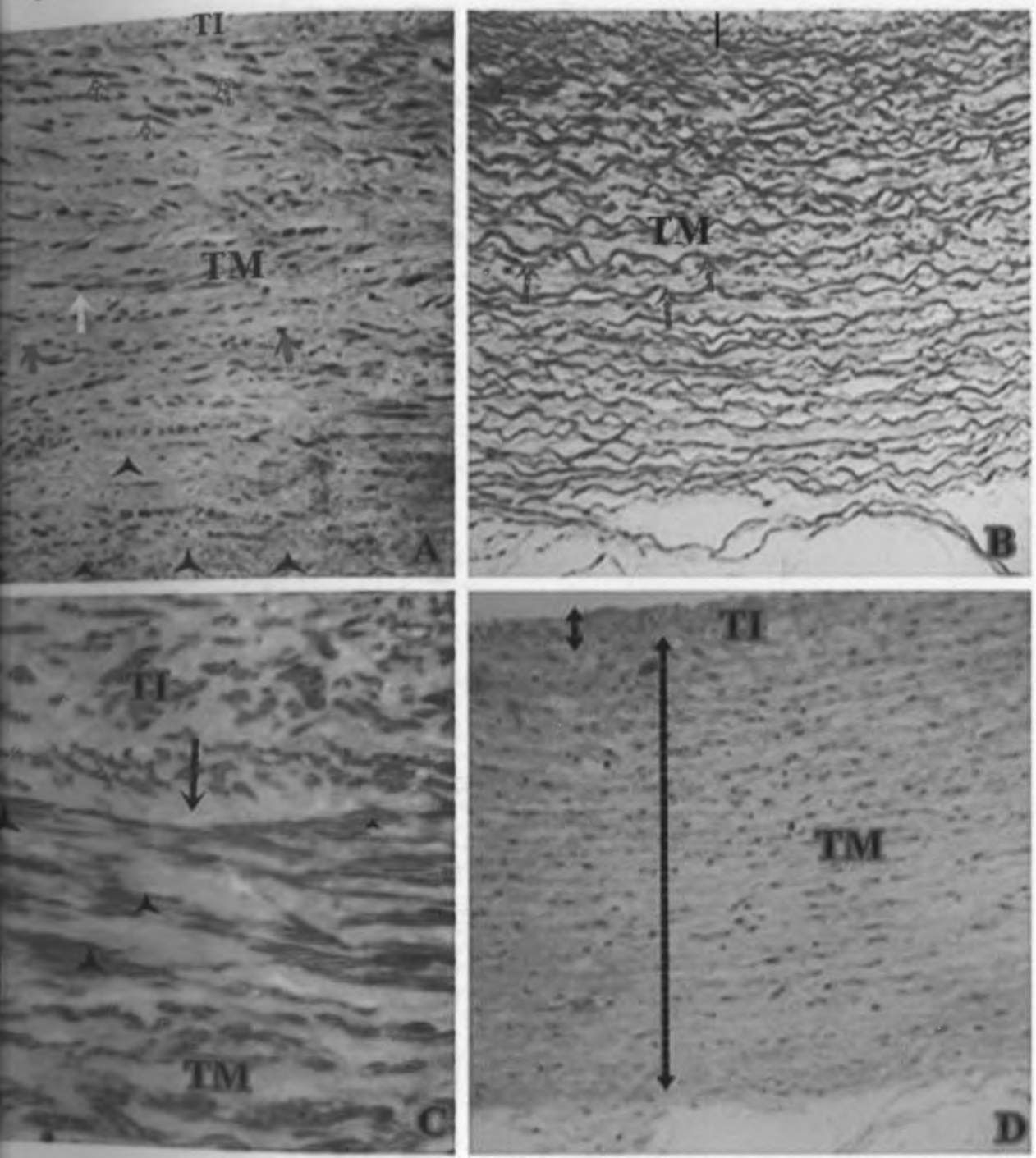
A: Photomicrograph of the common carotid artery tunica media. Concentric elastic lamellae are shown (white arrow), they are interspersed with collagen (starred) and circumferential smooth muscle cells [Red arrows]. Some smooth muscle cells are arranged longitudinally [blue arrow] (Magnification X100, Mason Trichrome).

B: Photomicrograph of the CCA media showing the concentric elastic lamellae (red arrow) in the tunica media (Magnification X100, Weigert Elastin).

C: Photomicrograph of the CCA showing circumferential smooth muscle cells (Starred) densely packed in the inner part of the tunica media to form the sub intimal zone (Magnification X400, Masson Trichrome).

D: The tunica media comprised of about 80% of the vessel wall (Magnification X40, Haematoxylin Eosin).

Figure 5: Photomicrograph of the tunica media of the common carotid artery



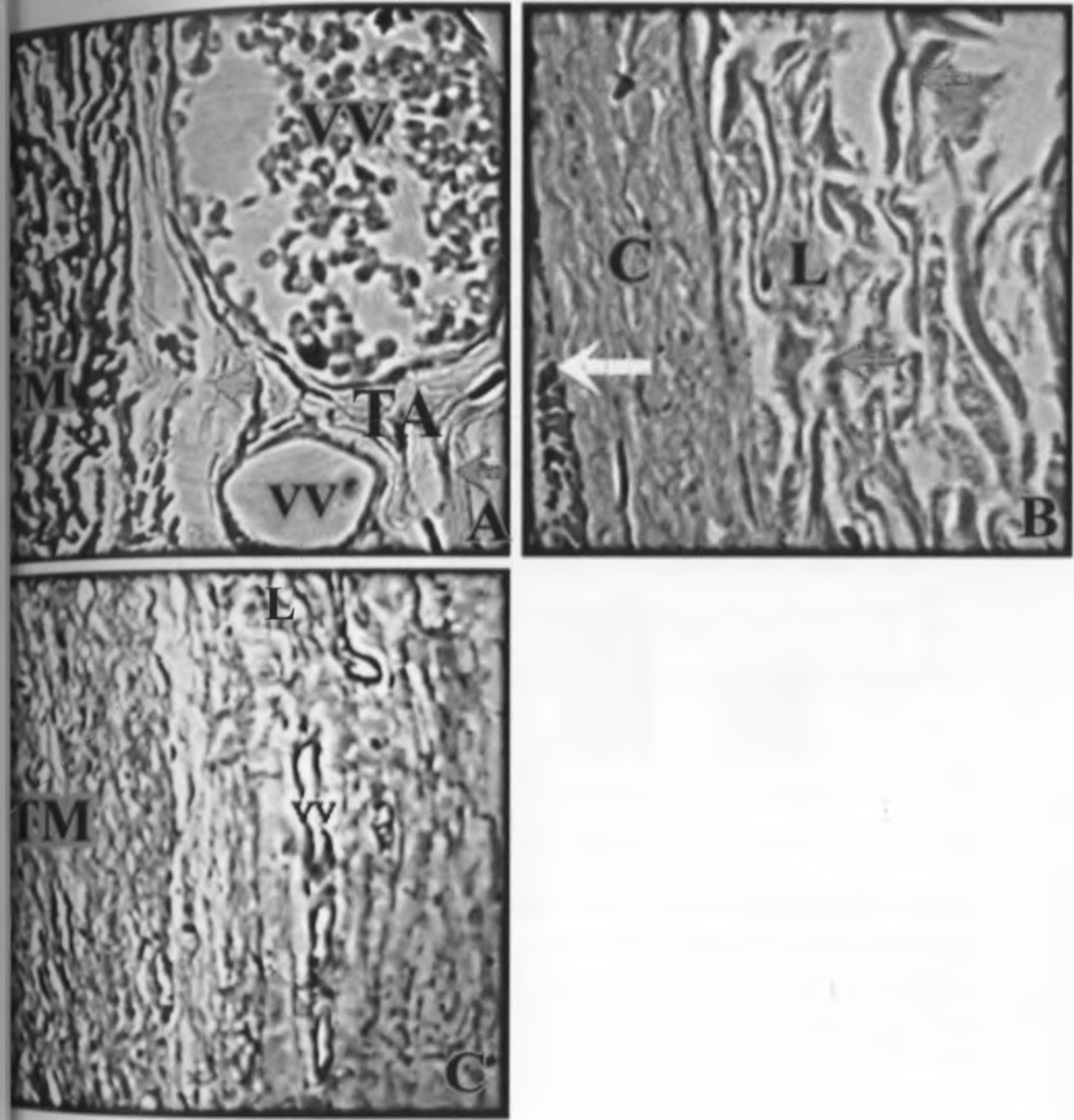
**Figure 6: The adventitia of the common carotid artery**

**A:** The tunica adventitia is predominantly fibroelastic, with interspersed prominent vessels (vv) (Magnification X400, Weigert Elastic).

**B:** It has two layers, the inner compact (C) and the outer loose (L) zone. The EEL (White arrow), and collagen (red arrow) are shown (Magnification X100, Mason Trichrome).

**C:** It also has prominent vasa vasora (vv) in the loose zone (L) (Magnification X100, Mason Trichrome).

Figure 6: Photomicrograph of the adventitia of the common carotid artery

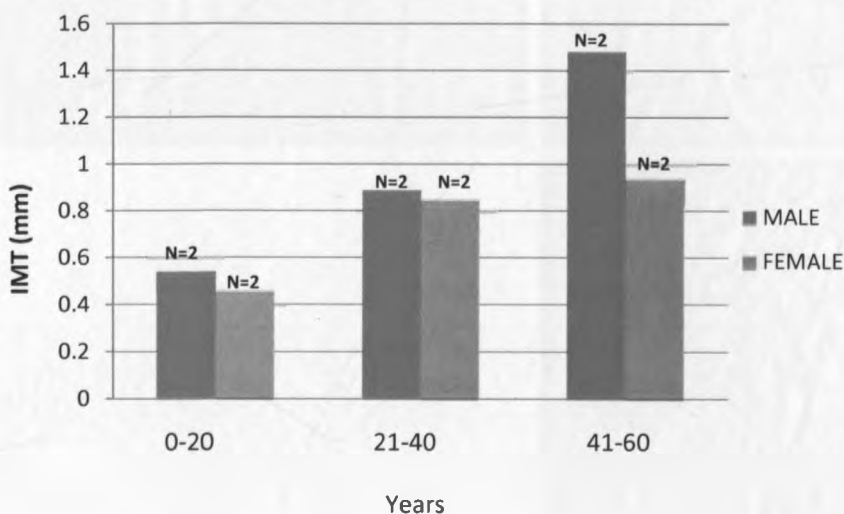


### 3.1 Histomorphometry of the common carotid artery

Mean carotid intimomedial thickness was  $0.86 \pm 0.22$  mm. The mean proximal, middle and distal carotid intimomedial thickness were  $0.86 \pm 0.26$  mm,  $0.84 \pm 0.28$  mm and  $0.90 \pm 0.35$  mm respectively (Table 1) [ $p=0.08$ ]. Males ( $0.97 \pm 0.22$ ) had a thicker intimomedial thickness when compared to females ( $0.77 \pm 0.06$ ), [ $p=0.05$ ].

Carotid intimomedial thickness increased with age;  $0.5 \pm 0.16$  mm,  $0.87 \pm 0.24$  mm and  $1.21 \pm 0.36$  mm for the age groups 0-20, 21-40 and >40 respectively [ $p=0.035$ ] (Figure 7).

Figure 7: Intimomedial thickness in different age groups.



### 3.2 Age and gender differences in the structure of the common carotid artery

There were no obvious gender differences in the histological organization of the carotid artery. Gender differences in the arterial structure became manifest in the age group 21 to 40 years. Males had a thicker arterial wall compared to females (Table 1, Figure 10). Males had a thicker subendothelial zone when compared to females (Figure 10). The largest subendothelial zone was composed of larger smooth muscle cells and abundant collagen with some elastic fibres. Features of early atherosclerosis such as focal intimal thickening, Intimal hyperplasia and increased intimomedial thickening occurred from the 3<sup>rd</sup> decade in males and



the 4<sup>th</sup> decade in females. Males had a less prominent internal elastic lamella when compared to females. The tunica media was thicker among males as compared to females. The media in males had about 30 - 35 thick circumferential elastic fibre lamellae. In comparison, females had 27 - 34 concentric lamellae (Fig. 8). There were no gender differences in the structure of the adventitia. The males had more prominent vasa vasora when compared to females.

### **3.3 Regional differences in the structure of the common carotid artery**

The common carotid artery showed segmental differences in the structure of the three tunics. The tunica intima in the proximal segment comprises of a single layer of endothelial cells and the SEZ and was  $0.86 \pm 0.26$  mm in thickness. These findings decrease distally such that in the middle it was  $0.84 \pm 0.28$  mm while the distal point it increased to  $0.90 \pm 0.35$  (Table 1).

The tunica media in the proximal segment comprises of elastic lamella with interspersed collagen and SMC and was  $0.77 \pm 0.31$  mm (Table 1). Its size reduced relatively from proximal to distal such that it was  $0.59 \pm 0.34$  mm in the middle segment and  $0.59 \pm 0.41$  mm in the distal segment. The content of smooth muscle increased distally, developing an inner muscular medial layer and an outer fibro elastic medial layer. The number of elastic lamellae decreased distally from 35 in the proximal segment, 28 in the middle segment and 25 in the distal segment.

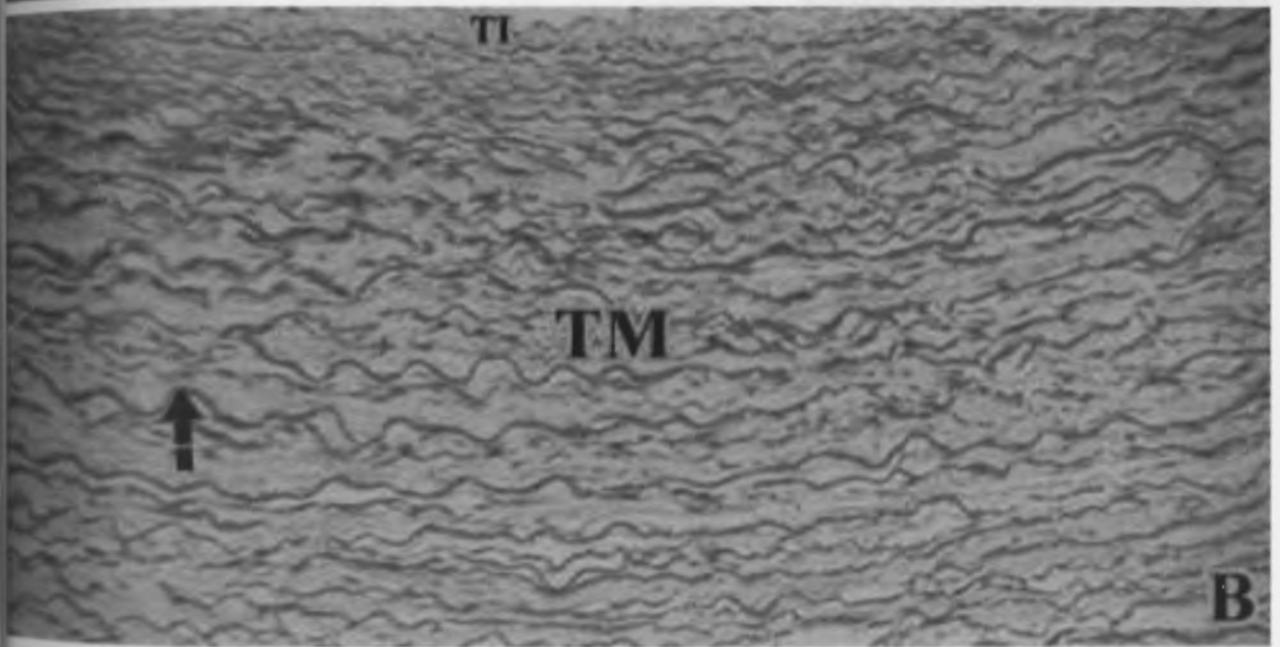
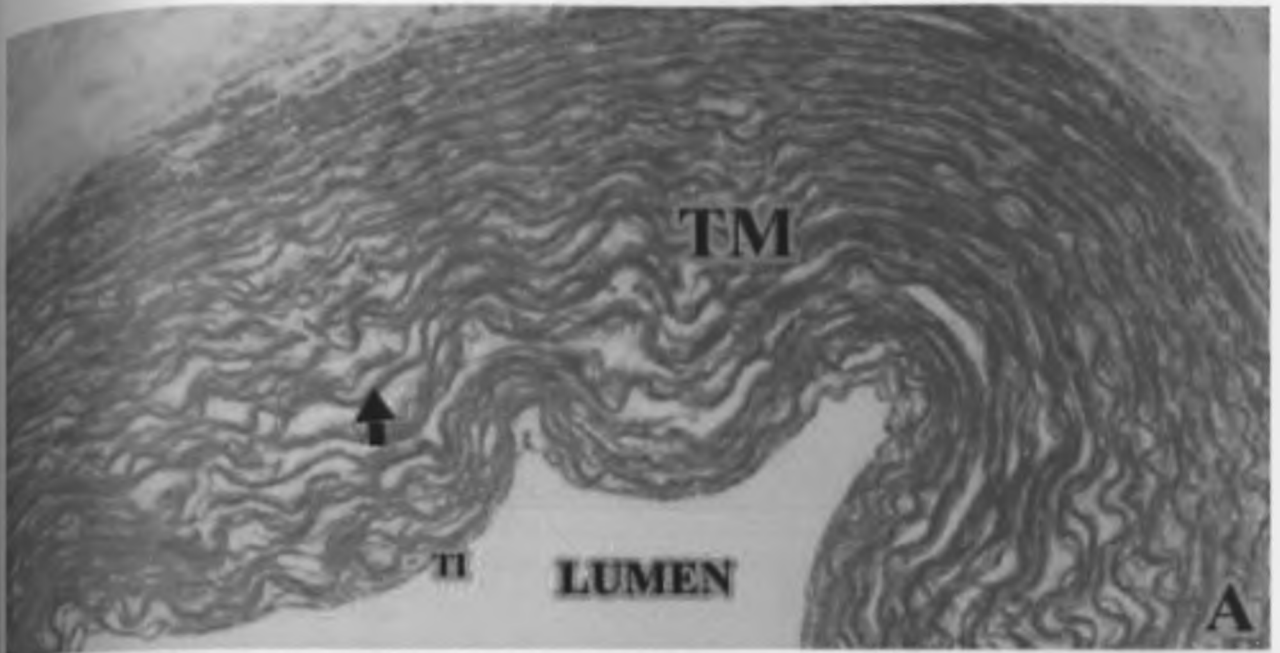
The thickness of the size of the tunica adventitia increased distally, developing from a single layered tunic to a bilayered structure distally, with an inner compact and an outer loose zone (Fig. 6, Fig. 8A-C). The diameter and number of the vasa vasora increased distally.

**Figure 8: Gender differences in the tunica media**

**A:** Photomicrograph showing the tunica media (TM) in a 25 year old female. The tunica media had 20 concentric elastic lamellae. Magnification X 40, Weigert Elastic.

**B:** Photomicrograph showing the tunica media (TM) in a 26 year old male. The tunica media had 30 concentric elastic lamellae. Magnification X 40, Weigert Elastic.

Figure 8: A photomicrograph showing gender differences in the tunica media



**Table 1: Carotid intimomedial thickness across the segments, age and gender groups**

Age group	Gender	SEGMENT		
		IMT A (mm)	IMT B (mm)	IMT C (mm)
0-10	Male (n=1)	0.64	0.47	0.52
	Female (n=1)	0.65	0.38	0.23
11-20	Male (n=1)	0.60	0.51	0.53
	Female (n=1)	0.78	0.40	0.51
21-30	Male (n=1)	0.81	0.70	0.82
	Female (n=1)	0.72	0.65	0.73
31-40	Male (n=1)	0.90	1.00	1.16
	Female (n=1)	0.89	1.03	0.89
41-50	Male (n=1)	1.33	1.32	1.34
	Female (n=1)	0.68	0.90	1.0
51-60	Male (n=1)	1.39	1.50	2.00
	Female (n=1)	0.72	1.15	1.20

### 3.4 Androgen receptor distribution in the common carotid artery

Androgen receptors were not detected in all the segments, age and gender groups of the common carotid artery (Table 2; Figure 11 A-F).

Table 2: Distribution of androgen receptors in the common carotid artery

AGE GROUP		SECTION		
		Proximal	Middle	Distal
A	TCC	27	25	29
	AR+	0	0	0
B	TCC	28	26	30
	AR+	0	0	0
C	TCC	29	29	32
	AR+	0	0	0

TCC – Total cell count per field.

AR+ - Total number of immunopositive cells for AR.

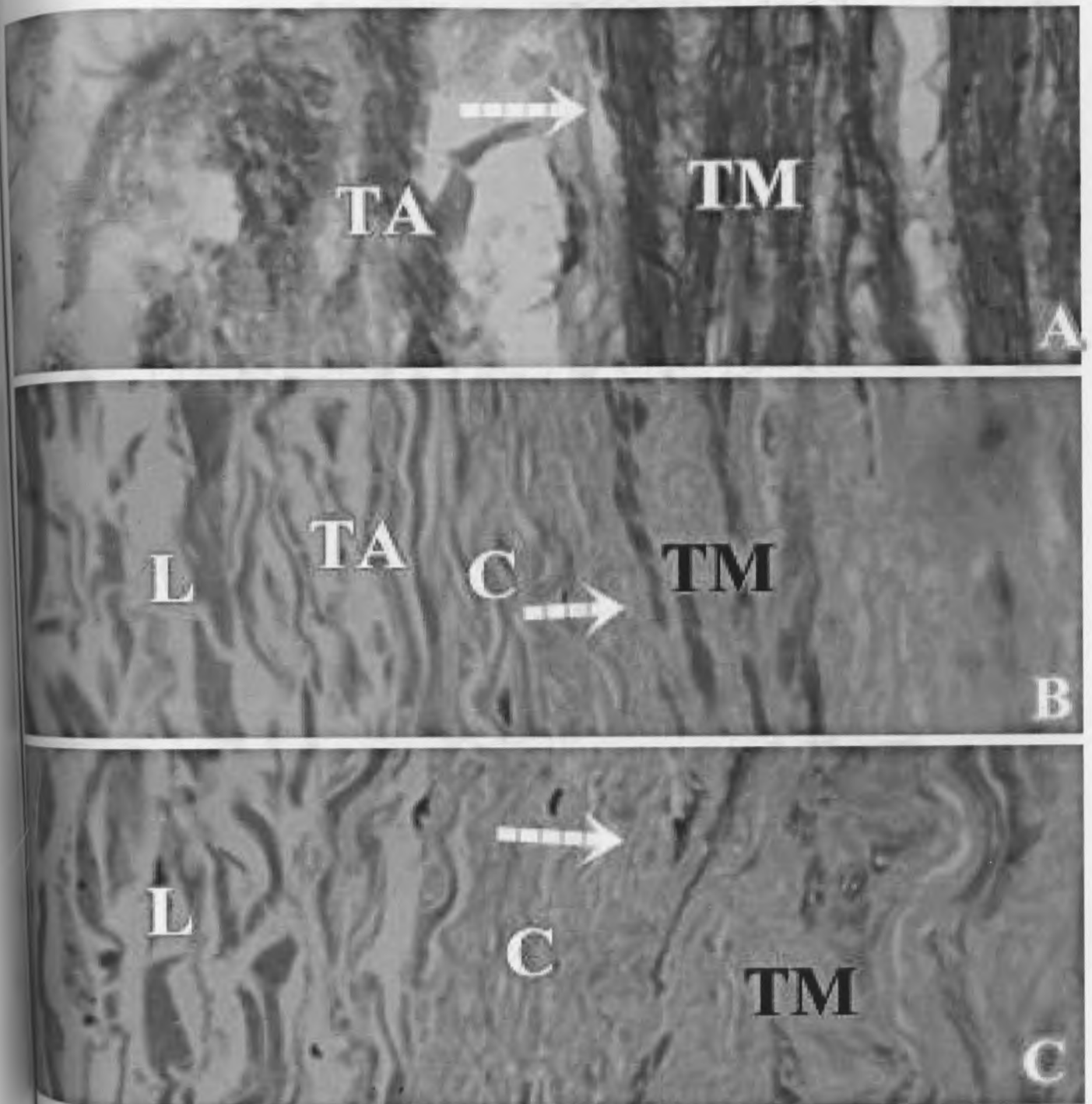
**Figure 9: Regional differences in the structure of the tunica adventitia**

**A:** Photomicrograph of a proximal section of the CCA showing the tunica Adventitia (TA) and Tunica Media (TM), separated by an external elastic lamella (Arrowed). (Magnification x100, MT stain).

**B:** Photomicrograph of a middle section of the CCA showing the tunica Adventitia (TA) and Tunica Media (TM), separated by an external elastic lamella (Arrowed). The tunica adventitia has two layers compact (C) and loose (L). (Magnification x100, MT stain).

**C:** Photomicrograph of a distal section of the CCA showing the tunica Adventitia (TA) and Tunica Media (TM). The tunica adventitia has two layers compact (C) and loose (L). (Magnification x100, MT stain).

Figure 9: A photomicrograph showing regional differences in the structure of the tunica adventitia



**Figure 10: Age and gender differences the carotid tunica intima**

**A:** Photomicrograph of the tunica intima of the proximal segment of the CCA in a 12 year old female. The endothelial cells are flat and short. The SEZ is composed of some SMCs, abundant collagen fibers and elastic fibers. The size of smooth muscle cells is relatively small (Magnification X100, Masson Trichrome).

**B:** Photomicrograph of the tunica intima of the proximal segment of the CCA in a 30 year old male. Endothelial cells are larger and elongated. The SEZ is composed of more SMCs with relatively reduced collagen fibers. (Magnification x100, Masson trichrome).

**C:** Photomicrograph of the tunica intima of the proximal segment of the CCA in a 60 year old female. The SEZ is composed of large SMC between collagen and elastic lamellae. VV are noted in this section (Magnification X100, Masson trichrome).



Figure 10: A photomicrograph showing the age and gender differences the carotid tunica intima

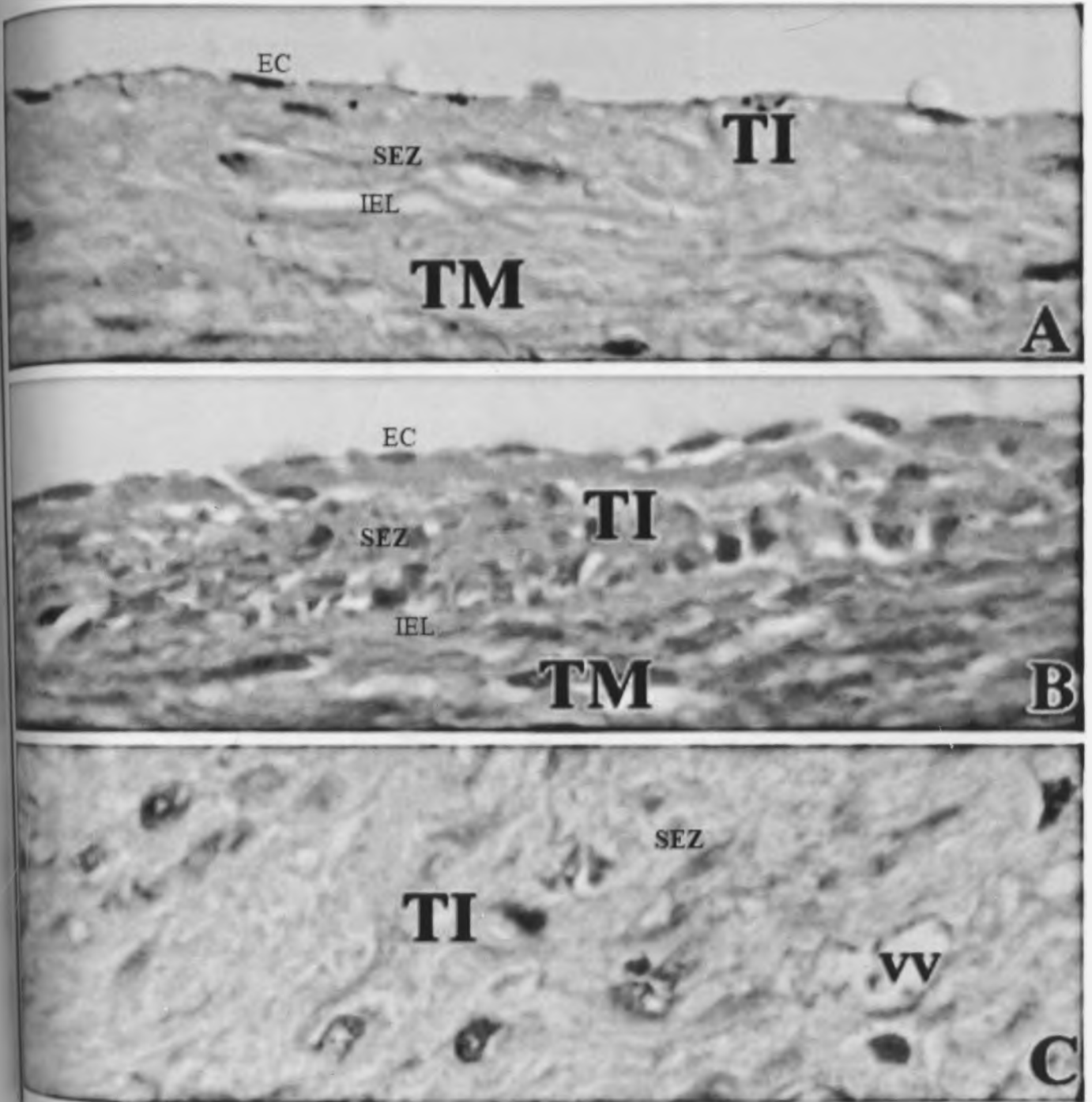


Figure 11: Non expression of androgen receptors in the carotid arterial wall

A: Subendothelial zone of the proximal segment of the carotid artery in a 25 year old female. Smooth muscle cell nuclei without androgen receptors are stained (Arrowed white).

B: The carotid artery tunica media in a 12 year old female is shown. There was no positive staining of androgen receptors in the smooth muscle cells arrowed white.

C: The common carotid artery tunica media in a 30 year old male is shown. Androgen receptors are not expressed in the smooth muscle cells (White arrow).

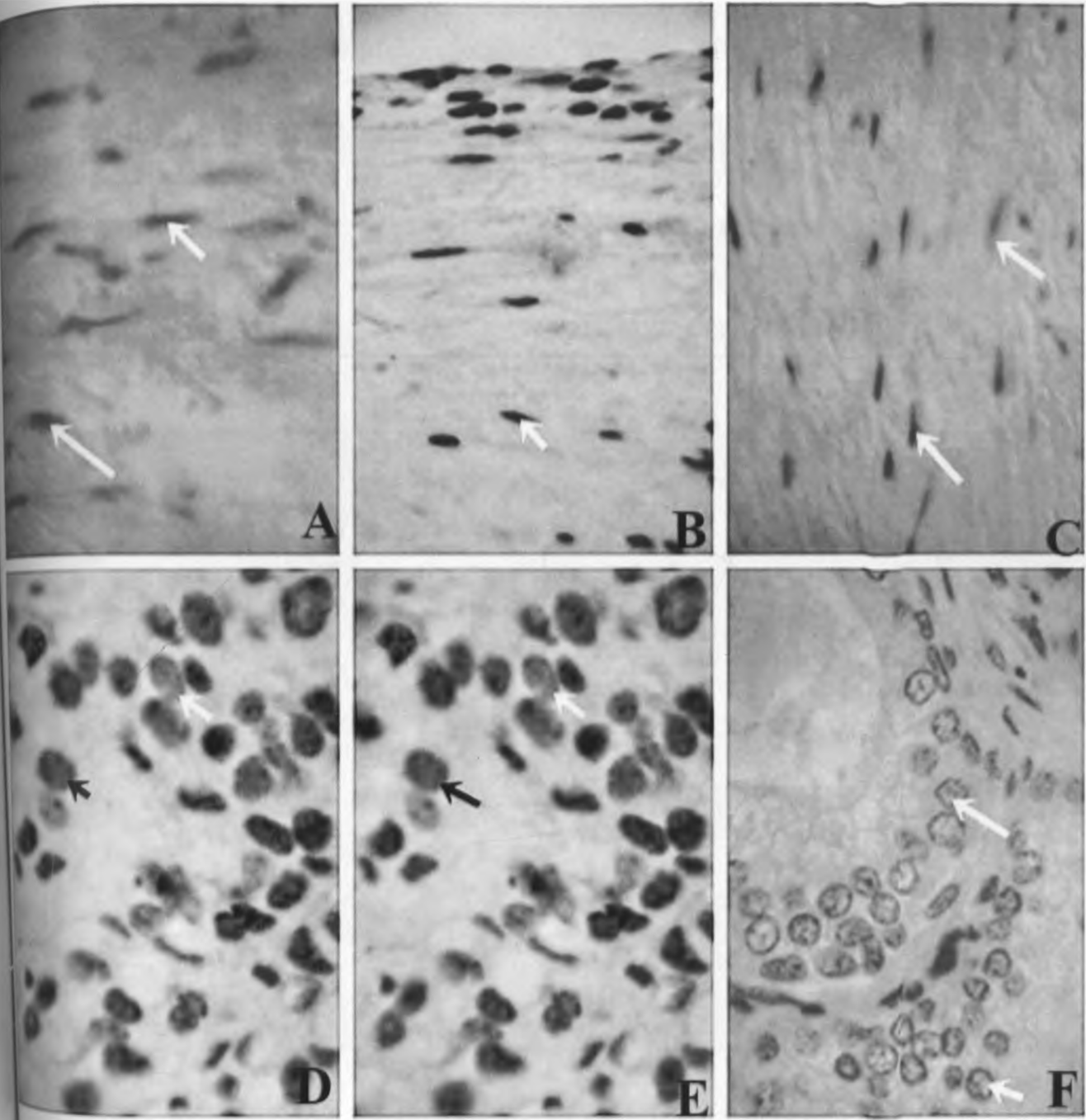
D: A positive control (post-mortem prostate) expressing brown stained nuclei with androgen receptors (Black arrow).

E: A positive control (post-mortem prostate) expressing brown stained nuclei with androgen receptors (AR- 318 positive cells black Arrow).

F: A negative control did not stain any androgen receptors (White arrow).

Magnification X 400 for figures A-F

Figure 11: A photomicrograph showing the non-expression of androgen receptors in the carotid arterial wall



**Table 3: Carotid Intimomedial thickness in various populations**

Study	Population	Method	IMT (mm)
Crouse et al., 1995	US	ultrasound	1.049
Simons et al., 1999	Caucasian	Ultrasound	0.94
Okeahialam et al., 2011	Nigeria	Ultrasound	0.91
Bots et al., 2005	Netherlands		0.82
Adaikkappan et al., 2002	Indian	Ultrasound	0.73
Özdemir et al., 2006	Turkey	Ultrasound	0.62
Sharma et al., 2009	Nepalese	Ultrasound	0.55
Pignoli et al., 1986	Italian	Histology	0.48
Present study	Kenya	Histology	0.86

**Table 4: Age differences in the carotid intimomedial thickness**

Study	Population	IMT (mm)		
		0-20	21-40	41-60
Ludwig et al., 2006	US	0.5	-	0.9
Ma et al., 2011	Taiwan			0.77
Pignoli et al., 1986	Italian		0.48	
Present study	Kenya	0.77	0.87	1.21

Table 5: Gender differences in the carotid intimomedial thickness

Study	Population	IMT (mm)	
		Males	Females
Crouse et al., 1996	US	0.83	0.76
Ziembicka et al., 2005	Poland	1.05	0.93
Lawlor et al., 2004	British	1.54	1.36
Ma et al., 2011	Taiwan	0.77	0.72
Present study	Kenya	0.97	0.77

## CHAPTER FOUR: DISCUSSION AND CONCLUSIONS

### 4.1 Histomorphology of the Human Common Carotid Artery

The common carotid artery comprises of three histological layers which had also been described in other arteries. The tunica intima comprises of a single layer of flat endothelial cells and the sub endothelial zone, consistent with reports from sheep, goats, giraffe and dogs (Kimani, 1983; Orsi et al., 2006; Parchami et al. 2009). Flat endothelial cells have better adaptation to the shear stress of a conduit vessel like the common carotid artery (Fisher et al., 2001; Fisher et al., 2002). In concurrence with observations made in other mammals, the sub endothelial zone was composed of collagen, elastic fibres and smooth muscle cells in different proportions (Kimani, 1981; Parke et al., 1995; Parchami et al., 2009). Sub endothelial smooth muscle cells were arranged axially as observed in the bovine CCA and pig aorta (Stergiopoulos et al., 2001; Timmins et al., 2010). The axial orientation of subendothelial vascular smooth muscle cells is useful in axial loading of pressure from the endothelial cells to the inner medial layer (Clark and Glagov, 1985). Subendothelial smooth muscle cells synthesise extracellular matrix and are prone to fatty infiltration (Schwartz et al., 1986; Aidinian et al., 2006). The subendothelial layer supports loads that are in the axial direction (Clark and Glagov, 1985), and is prone to remodeling related to the stresses exposed to it (Kimani, 1983) leading to thickening.

The CCA intima displayed focal thickening and intimal hyperplasia. This thickened intima was composed of large smooth muscle cells, collagen and elastic tissue, consistent with previous observations (Virmani et al., 2000). Intimal thickening is a reservoir for lipid retention and is a surrogate marker of atherosclerosis (Wilens, 1951; Nakashima et al., 2007). The observed wavy internal elastic lamella allows for distension of the artery and acts as a

macromolecular barrier between the two layers (Penn et al., 1994; Laurent et al., 2005). In some sections, and areas of intimal hyperplasia, the IEL was split into two. This feature has been seen in muscular arteries and in pathological states such as hypertension and intimal cushions (Todd and Friedman, 1972; Yamazoe et al., 1990). The wavy folding in the IEL observed in CCA is attributed to subendothelial smooth muscle insertion (Krsti, 2009).

The tunica media was the widest zone of the carotid artery wall. It was predominantly elastic as seen in dogs and rats (Pinto et al., 1998; Orsi et al., 2006). This layer is muscular in goats, sheep and giraffe (Kimani, 1983; Parchami et al., 2009). Elastic fibers ensure uniform tension distribution throughout vessel walls and are primarily responsible for the elastic recoil properties of large elastic arteries (Wagenseil et al., 2005). Elastic recoil imparts a hydraulic advantage to the circulatory system by converting intermittent cardiac output into steady flow, thereby reducing cardiac workload (Shadwick, 1999; Faury, 2001; Safar et al., 2003).

The tunica media was composed of 30-35 concentric elastic lamellae, more than the nine seen in rats (Pinto et al., 1998) and fifteen reported in dogs (Orsi et al., 2006). The number of elastic lamellae in the tunica media is related to the size of the mammal (Wolinsky and Glagov, 1967). Elastic lamellae were oriented circumferentially to allow for vascular distension of this conduit artery (Timmins et al., 2010). The propensity of the carotid artery to develop atherosclerosis is not to be anchored in the elastic nature of its tunica media because muscular arteries such as the coronaries are also predisposed to atherosclerosis (Hansson, 2005). It has however been noted that a decline in the amount and quality of elastic tissue in the tunica media makes elastic arteries unable to tightly set stress force values thus creating local wall shear stress reductions that predispose individuals to intimal thickening,

which also contributes to vascular tensile support (Masawa et al., 1994; Caralo et al., 1999). In support of the observations made by some authors, medial smooth muscle cells were oriented circumferentially (Wolinsky and Glagov, 1967; Arner and Uvelius, 1982; Dingemans et al., 2000), contrary to the observations made by Bierring and Kobayashi, who observed oblique orientation of medial SMC (Bierring and Kobayashi, 1963). In some sections, a thin subintimal smooth muscle zone was observed. Subintimal smooth muscle cells are early features of atherosclerosis (Hartman, 1977). The presence of these cells explains why the carotid artery is at risk of atherosclerosis.

The tunica adventitia of the carotid arteries was composed of elastic fibers, collagen fibres with longitudinal, transverse and oblique arrangements. This composition and arrangement of fibers is also observed in femoral and renal arteries (Gutterman, 1999). In middle and distal sections, the tunica adventitia had two zones, inner compact and outer loose. There are no previous reports on the zonation of the tunica adventitia. The compact inner layer of the adventitia may offer more support to the tunica media, when compared to the generally loose tunica media especially in elevated blood pressure. Also, the adventitial elastic fibres and lamellae showed a variable pattern, with a predominance of circumferential dispositions. This arrangement was similarly reported for the CCA of the dog (Orsi et al. 2006). The different spatial arrangements of the connective fibres observed in the adventitial layer perhaps help to guarantee the vascular wall integrity and protect the arterial wall.

The human CCA had prominent vasa vasora extending into the tunica media and intima. This was consistent with the increase in the thickness of the vessel wall from 0.5mm to 1.21mm in the young and the elderly respectively. Bo et al (1989) observed that when the thickness of an artery exceeds the ability of simple diffusion of nutrients from the lumen, vasa vasora extend



to supply the media and intima. This extensive network of vasa vasora in the adventitia arises from branch points of parent arteries. Vasa vasora in the media and intima arise predominantly from adventitial vasa, but can arise from the lumen (Williams and Heistad, 1996; Bayer et al., 2002). The large size of the CCA vasa vasora is of clinical significance as they are related to the size of the vessel wall (Gossl et al., 2003) and are a marker of vessel remodelling to meet its metabolic needs.

#### **4.2 Histomorphometry of the Common carotid artery**

The carotid intimomedial thickness among Kenyans of 0.86mm found by histology closely compares with ultrasound results observed in Netherlands and Nigerians [Table 3] (Bots et al., 2005; Okealahim et al., 2011). Although there is a difference in the methods used, intimomedial thickness measured by ultrasound correlates significantly with the intimomedial thickness determined by histology (Pignoli et al., 1986; Choi et al., 2009). The difference between the intimal medial thickness in the present sample and the 0.48mm observed by Pignoli et al (1986) on microscopy is remarkable. Although there were similarities in sample preparation, differences exist in the sample size, segment, age and measurement method used. Pignoli evaluated 44 male as opposed to the 12 (6 male, 6 female) carotid samples used in the present study. Their samples aged between 20-25yrs while our samples ranged from 1-60 yrs. When adjusted for age and gender, carotid intimal medial thickness in samples between 21-30yrs in the present study were 0.74, still higher than the observations of Pignoli. Furthermore the CCA segments measured by Pignoli were unclear. Finally, Pignoli et al (1986) used a graduated ocular piece for measurement, while the present study used the Scion Image™ Multiscan software for intimal medial thickness assessment. It is however unclear as

to whether methodology, ethnic, or lifestyle differences could explain this remarkable difference in IMT.

This thickness was lower than 1.049mm reported in Americans and it was higher than 0.73mm reported in Indians (Crouse et al., 1995; Adaikkappan et al., 2002). Intimomedial thickness in Kenyans was also lower than the 0.94mm reported in Caucasians, in contrast with earlier reports that IMT is significantly higher in blacks than Caucasians (D'Agostino et al., 1996; Chambless et al., 1997; Urbina et al., 2002). Carotid intimomedial thickness shows population related differences, which are to be attributed to the distribution of cardiovascular risk factors in these populations. The IMT of the CCA is a good marker for both the presence of early arteriosclerosis (Pignoli et al., 1986; Grobbee and Bots, 1994) and the degree of arteriosclerosis of an individual (Ebrahim et al., 1999; Simons et al., 1999; Frauchiger et al., 2001). Increases in the thickness of the tunica intima and media of the carotid artery are directly associated with an increased risk of atherosclerosis in other vascular beds such as the coronary arteries (Riley et al., 1986; Arnet et al., 1994; Salonen and Salonen, 1991; O'Leary et al., 1999). The critical measure of carotid intimomedial thickness which imperatively predicts atherosclerosis is however still unclear, even in the present study. The utility of the current measure in a Kenyan population is limited by tissue shrinkage, sample size and selection challenges, which affected the measured intimomedial thickness. These pilot observations in the Kenyan sample can be anchored by follow-up in vivo studies using doppler ultrasound.

An increased CIMT may reflect a nonatherosclerotic adaptive arterial response (Bots et al., 1997a; Glagov et al., 1987). Common carotid IMT is related to changes in local shear stress and tensile stress (Bots et al., 1997b). It has also been reported that increased IMT and atherosclerotic plaques are overcompensated by an accompanied increase in lumen diameter

of the common carotid arteries (Bonithon-Kopp et al., 1996). When arterial enlargement accompanies increased wall thickness, less lumen constriction results than expected (Crouse et al., 1994; Crouse et al., 1996).

#### **4.3 Segmental differences in the histomorphometry of the common carotid artery**

Observations of the present study reveal that there are segmental changes in the structure of the carotid artery. These include increase in the intimomedial thickness; reduction in the number of elastic lamellae, increase in the amount of tunica media vascular smooth muscle cells, development of two layers of the tunica adventitia and increase in size of the vasa vasora along the vessel profile. The increase in the thickness of the intimomedial complex along the vessel profile observed in the CCA could be theoretically related to the sliding of the mural coats and to the local hemodynamic stress on the vascular wall layers at the origin of the CCA and towards the carotid bifurcation (Kimani, 1983; Milner et al., 1998; Willekes et al., 1999). Observations of the current study imply that the distal segment of the carotid artery is prone to atherosclerosis as compared to the middle or proximal segment. These observations also suggest that the distal carotid artery is more adapted to the hemodynamic stresses exposed to it (Kamiya and Togawa 1980; Glagov, 1994). Nevertheless, distal thickening of the carotid intimomedial thickness occurs earlier than any other segment of the artery (Persson et al., 1994). This observation also explain the conclusions by Ebrahim et al (1999), that IMT measurements in vivo should focus on the distal segment of the carotid artery to give a more clinically useful parameter. The reduction of the number of elastic lamellae with concurrent increase in the amount of smooth muscle cells in the tunica media along the vessel profile is to be attributed to the gradual transition of the arterial structure from elastic to muscular. This transition is however completed in the more distal branches of the carotid artery (Schievink et al., 1994).

#### **4.4 Age and gender differences in structure and morphometry of the common carotid artery**

The carotid artery displayed age and gender difference in its dimensions.

##### **4.4.1: Age differences in structure and morphometry of the common carotid artery**

Carotid intimomedial thickness increased with age, supporting observations made in previous studies [Table 4] (Hort et al., 1982; Labropoulos et al., 1998; Osika et al., 2009; Cobble and Bale, 2010; Jarauta et al., 2010). The increased intimomedial thickness comprised more elastic lamellae, smooth muscle cells and size, and increased collagen deposition. Age changes in the structure of the carotid artery including diffuse intimal thickening has been reported to develop from an early age in human arteries before atherosclerosis evolves (Movat et al., 1958; Nakashima et al., 2002; Nilsson et al., 2008). Changes in the structure of the carotid artery with age could be attributed to hemodynamic differences related to the variability of the carotid bifurcation with age (Goubergrits et al., 2002), as well as lifestyle changes (Okada et al., 2004). Individuals from adolescent age should be encouraged to adopt cardiovascular friendly lifestyles to slow the effects of aging on the carotid artery which would otherwise complicate into atherosclerosis later in life.

##### **4.4.2: Gender differences in structure and morphometry of the common carotid artery**

Carotid intimomedial thickness was thicker in males than females, consistent with previous reports in literature [Table 5] (Crouse et al., 1996; Ebrahim et al., 1999) that IMT shows gender dimorphism. The observed wider intimomedial thickness in males begins from adolescence and partly explains why males are more prone to atherosclerosis when compared to females (Bohm et al., 2009). Contrary to previous observations (Sinning et al., 2011), the

gender differences in the dimensions of the carotid intima and media were pronounced in the third age group especially among males. Males in the third age group had unusually thick vessel walls compared to the females. This disparity could be in part related to the differences in the ethnic groups and age groups considered and our small sample size. Nonetheless postmenopausal had increased intimomedial thickness, significantly more than their premenopausal counterparts. The age difference in the intimomedial thickness between the females in the third age group and the females in the second age group was comparable to the gender difference in the CIMT. This suggests that menopause is accompanied with significant increase in carotid intimomedial thickness in support of the observations made by Espeland et al (1995). The similarity in pattern of difference in the intimomedial thickness between the genders, and between the post-menopausal premenopausal females generally also implies that females are masculinized after menopause. The gender difference in the structure of the vessel seems to be related to the effects of oestrogens and androgens.

#### **4.5 Androgen receptor expression in the common carotid artery**

In the present study, androgen receptors were not expressed in any of the layers, segments or age groups of the common carotid artery. Findings of the present study are at variance with the observations of Liu et al (2005) who immunolocalised androgen receptors in the post-mortem coronary arterial wall, inversely relating their numbers to early atherosclerosis. The absence of these receptors in the carotid wall suggests that androgen receptors have a limited genomic role in the carotid arterial structure and subsequent development of atherosclerosis. A similar conclusion was made by Christian et al., (2006), who did not localise AR in the coronary arterial wall. On the contrary, McRobb et al (2009) immunolocalised AR in the rat innominate artery and the aortic sinus and positively associated them with increased

calcification of the atherosclerotic plaques. While androgens play a role in the cardiovascular system, the present study supports the observations that some androgen effects (non-genomic) in arteries are not mediated through androgen receptors (Chou et al., 1996; Costarella et al., 1996; Rubio et al., 1998; Reckelhoff et al., 1999; Williams et al., 2002). In addition, the present observations suggest that the perceived gender disparity in the structure of the carotid artery with worsening atherosclerosis is largely unrelated to the effects of androgens. Since androgen receptors were not localised in the common carotid arterial wall, it is has therefore likely that the view that androgens are solely 'harmful' or 'beneficial' is simplistic as regards the understanding of the effects of androgen receptors on atherosclerosis. Findings of the present study also failed to link androgen receptor expression to the perceived gender dichotomy in the structure of the carotid artery as previously suggested (Parchami and Dehkordi, 2011). Despite this, gender disparity in cardiovascular disease persists with androgen use and abuse is increasing in our population, either for therapeutic or recreational reasons. Whether androgens adversely affect CVD in either men or women remains a contentious issue that is in desperate need of more research.

The absence of the AR in the CCA could also be partly attributed to post-mortem loss or low numbers limiting detection (Mainwaring and Mangan, 1973), since Fodor et al (2002) found that the post-mortem stay before fixation and the duration formalin fixation affects the expression of steroid receptors in post-mortem material. Steroid receptors deteriorate gradually after 24hrs before fixation and their retrieval is reduced after 20 days of formalin fixation. In this regard, there should have been residual cells stained even if most of them were hydrolysed post-mortem. In addition, the prostate controls which were harvested at the same post-mortem duration expressed the receptors, supporting the thought that these receptors were in few numbers that could not be detected. Fodor et al (2002) also observed

that microwave retrieval androgen receptor epitopes from formalin fixed samples occurs even after 20 days of fixation. This suggests that the receptors should have been extracted if they were present in the artery. Lastly, the vascular androgen receptors could have different epitopes when compared to prostatic carcinoma ones. This is supported by the observed heterogeneity of the androgen receptors expressed in prostatic cancer tissue (De winter et al., 2005). Monoclonal (MAb) androgen receptor marker used in the present study may not suitably stain the carotid androgen receptors because a small change in the structure of an epitope (e.g., as a consequence of genetic polymorphism, glycosylation, and sumoylation) can markedly affect the function of a MAb (Thomas et al., 2004; Lipman et al., 2005) in staining the vascular AR, leading to a false absence of this receptors.

### **Conclusions**

The human common carotid is an elastic artery and its structure shows regional and age differences in dimensions. The distal carotid has the largest intimomedial thickness. The carotid artery does not display gender differences in the histological organization although males have a thicker intimomedial thickness than females. Postmenopausal women have a masculinized CIMT. There was no immunostaining of AR-318 in the carotid specimens. Histological features of early atherosclerosis are manifest from the 3<sup>rd</sup> decade in males and 4<sup>th</sup> decade in female.

## REFERENCES

1. Adaikkappan M, Sampath R, Felix A JW, Sethupathy S. 2002. Evaluation of carotid atherosclerosis by B'mode ultrasonographic study in hypertensive patients compared with normotensive patients. *Indian J Radiol Imag.* 12; 365–368.
2. Ahimastos AA, Formosa M, Dart AM, Kingwell BA. 2003. Gender Differences in Large Artery Stiffness Pre- and Post-Puberty. *J Clin End Met.* 88: 5375-5380.
3. Aidinian G, Weiswasser JM, Arora S, Abularrage CJ, Singh N, Sidawy AN. 2006. Carotid Plaque Morphologic Characteristics. *Perspect Vasc Surg Endovasc Ther.* 18: 63-70.
4. Alexandersen P, Haarbo J, Byrjalsen I, Lawaetz H, Christiansen C. 1999. Natural Androgens Inhibit Male Atherosclerosis. *Circ Res.* 84:813-819
5. Anderson DR, Pepine CJ. 2007. Gender Differences in the Treatment for Acute Myocardial Infarction; Bias or Biology? *Circulation.* 115: 823-826.
6. Arner A, Uvelius B. 1982. Force-velocity characteristics and active tension in relation to content and orientation of smooth muscle cells in aortas from normotensive and spontaneous hypertensive rats. *Circ Res* 50: 812–821.
7. Arnet DK, Evans G, Riley WA. 1994. Arterial stiffness: a new cardiovascular risk factor. *Am J Epidemiol.* 140: 669–682.
8. Bayer IM, Caniggia I, Adamson SL, Langille BL. 2002. Experimental angiogenesis of arterial vasa vasorum. *Cell Tissue Res.* 307:303-13.



9. Bayer-Garner I, Givens V, Smoller B. 1999. Immunohistochemical Staining for Androgen Receptors: A Sensitive Marker of Sebaceous Differentiation. *Am J of Dermatopathol.* 21: 426
10. Bernini GP, Sgro' M, Moretti A, Argenio GF, Barlascini CO, Cristofani R, Salvetti A. 1999. Endogenous Androgens and Carotid Intimal-Medial Thickness in Women. *J Clin Endo and Met.* 84: 2008-2012.
11. Bernini GP, Moretti A, Sgró M, Argenio GF, Barlascini CO, Cristofani R, Salvetti A. 2001. Influence of endogenous androgens on carotid wall in postmenopausal women. *Menopause.* 8:43-50.
12. Bierring F, Kobayasi T. 1963. Electron microscopy of the normal rabbit aorta. *Acta Pathol Microbiol Scand* 57: 154–168.
13. Bo WJ, McKinney WM, Bowden RL. 1989. The origin and distribution of vasa vasorum at the bifurcation of the common carotid artery with atherosclerosis. *Stroke.* 20:1484-7.
14. Bohm B, Hartmann K, Buck M, Oberhoffe R. 2009. Sex differences of carotid intima-media thickness in healthy children and adolescents. *Atherosclerosis.* 206: 458-463.
15. Bonithon-Kopp C, Touboul PJ, Berr C, Magne C, Ducimetiere P. 1996. Factors of carotid arterial enlargement in a population aged 59 to 71 years: the EVA study. *Stroke.* 27:654–660.
16. Bonnel RW, Pritchett CP, Rardin T. 1941. Treatment of angina pectoris and coronary artery disease with sex hormones. *Ohio State Med J.* 37:554

17. Bösel J, Kasper AS, Weichert W, Bohner G, Schreiber S J, Endres M. 2010. Surgery after Anticoagulation in Stroke Patients with Nonatherosclerotic Internal Carotid Artery Thrombus: A Clinicopathological Case Series. *Cerebrovasc Dis.* 29: 304-307.
18. Bots ML, Hofman A, Grobbee DE. 1997 a. Increased common carotid intima-media thickness. Adaptive response or a reflection of atherosclerosis? Findings from the Rotterdam Study. *Stroke.* 28:2442–2447.
19. Bots MI, Hoes AW, Koudstaal PJ, Hofman A, Grobbee DE. 1997 b. The Rotterdam Study: Common Carotid Intima-Media Thickness and Risk of Stroke and Myocardial Infarction. *Circulation.* 96: 1432-1437.
20. Bots ML, Grobbee DE, Hofman A, Witteman JCM. 2005. Common carotid intima medial thickness and risk factor for myocardial infarction: the role of lumen diameter. *Stroke.* 36: 762-767.
21. Boyd JD. 1934. Absence of the Right Common Carotid Artery. *J Anat.* 68: 551–557.
22. Cameron JD, Bulpitt JC, Pinto ES, Rajkumar C. 2003. The Aging of Elastic and Muscular Arteries, A comparison of diabetic and non-diabetic subjects. *Diabetes Care.* 26:2133-2138.
23. Carallo C, Irace C, Pujia A, De Franceschi MS, Crescenzo A, Motti C, Cortese C, Mattioli PL, Gnasso A. 1999. Evaluation of Common Carotid Hemodynamic Forces Relations With Wall Thickening. *Hypertension.* 34:217-221.
24. Celermajer DS, Sorensen KE, Gooch VM. Non-invasive detection of endothelial dysfunction in children and adults at risk of atherosclerosis. *Lancet* 1992; 340:1111–1115.

25. Chambless LE, Heiss G, Folsom AR, Rosamond W, Szklo M, Sharrett AR, Clegg LX. 1997. Association of coronary heart disease incidence with carotid arterial wall thickness and major risk factors: the Atherosclerosis Risk In Communities (ARIC) Study, 1987–1993. *Am J Epidemiol.* 146: 483–494.
26. Channer KS, Jones TH. 2003. Cardiovascular effects of testosterone: implications of the “male menopause”? *Heart.* 89: 121–122
27. Choi YS, Youn HJ, You JS, Park CS, Oh YS, Chung WS. 2009. Measurement of the Intimal Thickness of the Carotid Artery: Comparison Between 40 MHz Ultrasound and Histology in Rats. *Ultrasound Med Biol.* 35: 962-966.
28. Chou TM, Sudhir K, Hutchinson SJ, Ko E, Amidon TM, Collins P, Chatterjee K. 1996. Testosterone induces dilation of canine coronary conductance and resistance arteries in vivo. *Circulation.* 94: 2614–2619.
29. Christian RC, Liu PY, Harrington S, Ruan M, Miller VM, Fitzpatrick LA. 2006. Intimal estrogen receptor (ER) beta, but not ER alpha expression, is correlated with coronary calcification and atherosclerosis in pre- and postmenopausal women. *J Clin Endocrinol Metab.* 91:2713-20.
30. Clark JM, Glagov S. 1985. Transmural organization of the arterial media. The lamellar unit revisited. *Arteriosclerosis* 5: 19–34.
31. Cobble M, Bale B. 2010. Carotid intima-media thickness: knowledge and application to everyday practice. *Postgrad Med.* 122:10-8.
32. Colditz CA, Willett MD, Stampfer MJ, Rosner B, Speizer FE, Hennekens CH. Menopause and the risk of coronary heart disease in women. *N Eng J Med* 1987; 316:1105-10.

33. Costarella CE, Stallone JN, Rutecki GW, Whittier FC. 1996. Testosterone causes direct relaxation of rat thoracic aorta. *J Pharmacol Exp Ther.* 277:34–39.
34. Crouse JR, Goldbourt U, Evans G, Pinsky J, Sharrett AR, Sorlie P, Riley W, Heiss G. 1994. Arterial enlargement in the Atherosclerosis Risk in Communities (ARIC) cohort. In vivo quantification of carotid arterial enlargement. The ARIC Investigators. *Stroke.* 25:1354–1359.
35. Crouse JR, Goldbourt U, Evans G, Pinsky J, Sharrett AR, Sorlie P, Riley W, Heiss G. 1996. Risk factors and segment-specific carotid arterial enlargement in the Atherosclerosis Risk in Communities (ARIC) cohort. *Stroke.* 27:69-75.
36. Crouse JR, Craven TE, Hagaman AP, Bond GM. 1995. Association of Coronary Disease With Segment-Specific Intimal-Medial Thickening of the Extracranial Carotid Artery. *Circulation.* 92:1141-1147.
37. D'Agostino RB Jr, Burke G, O'Leary D, Rewers M, Selby J, Savage PJ, Saad MF, Bergman RN, Howard G, Wagenknecht L, Haffner SM. 1996. Ethnic differences in carotid wall thickness. The Insulin Resistance Atherosclerosis Study. *Stroke.* 27: 1744–1749.
38. De Groot E, Hovingh GK, Wiegman A, Duriez P, Smit AJ, Fruchart JC, Kastelein JJP. 2004. Measurement of Arterial Wall Thickness as a Surrogate Marker for Atherosclerosis. *Circulation.* 109: III-33 – III-38
39. De Winter JAR, Trapman J, Vermeij M, Mulder E, Zegers ND, van der Kwast TH. 1991. Androgen receptor expression in human tissues: an immunohistochemical study. *J Histochem Cytochem.* 39: 927-936.

40. De Winter JAR, Trapman J, Brinkmann AO, Boersma WJA, Mulder E, Schroeder FH, Claassen E, Van Der Kwast TH. 2005. Androgen receptor heterogeneity in human prostatic carcinomas visualized by immunohistochemistry. *J Pathol.* 160: 329-332.
41. Death AK, McGrath KC, Sader MA, Nakhla S, Jessup W, Handelsman DJ, Celermajer DS. 2004. Dihydrotestosterone promotes vascular cell adhesion molecule-1 expression in male human endothelial cells via a nuclear factor-kappa B-dependent pathway. *Endocrinology.* 145:1889-97.
42. Dingemans KP, Teeling P, Lagendijk JH, Becker AE. 2000. Extracellular matrix of the human aortic media: an ultrastructural histochemical and immunohistochemical study of the adult aortic media. *Anat Rec* 258: 1–14.
43. Drury RAB and Wallington EA. 1967. *Carleton's Histological Technique.* Oxford University Press, New York. 114-143.
44. Duvall WL, Vorchheimer DA .2004. Multi-bed vascular disease and atherothrombosis: scope of the problem. *J Thromb Thrombolysis.* 17:51–61.
45. Ebrahim S, Papacosta O, Whincup P, Wannamethee G, Walker M, Nicolaides AN, Dhanjil S, Griffin M, Belcaro G, Rumley A, Lowe DO. 1999. Carotid plaque, intima media thickness, cardiovascular risk factors, and prevalent cardiovascular disease in men and women: the British Regional Heart Study. *Stroke.* 30: 841–850.
46. Eng J. 2003. Sample Size Estimation: How Many Individuals Should Be Studied? *Radiology.* 227:309–313.
47. English KM, Steeds R, Jones TH, Channer KS. 1997. Testosterone and coronary heart disease: is there a link? *Q J Med.* 90:787-791.

48. Espeland MA, Applegate W, Furberg CD, Lefkowitz D, Rice L, Hunninghake D. 1995. Estrogen Replacement Therapy and Progression of Intimal-Medial Thickness in the Carotid Arteries of Postmenopausal Women. *Am. J. Epidemiol.* 142: 1011-1019.
49. Faury G. 2001. Function–structure relationship of elastic arteries in evolution: from microfibrils to elastin and elastic fibres. *Pathologie Biologie.* 49: 310-325.
50. Fernie J M, Lamb D. 1985. New method for measuring intimal component of pulmonary arteries. *J Clin Pathol.* 38:1374-1379.
51. Fisher AB, Chien S, Barakat AI, Nerem RM. 2001. Endothelial cellular response to altered shear stress. *Am J Physiol Lung Cell Mol Physiol.* 281:L529-L533.
52. Fisher AB, Al-Mehdi AB, Manevich Y. 2002. Shear stress and endothelial cell activation. *Crit Care Med.* 30:S192-7.
53. Fodor M, van Leeuwen FW, Swaab DF. 2002. Differences in Postmortem Stability of Sex Steroid Receptor Immunoreactivity in Rat Brain. *J Histochem Cytochem.* 50: 641-650.
54. Folsom AR, Kronmal RA, Detrano RC, O’leary DH, Bild DE, Bluemke DA, Budoff MJ, Liu K, Shea S, Szklo M, Tracy SP, Watson KE, Burke GL. 2008. Coronary Artery Calcification Compared With Carotid Intima-Media Thickness in the Prediction of Cardiovascular Disease Incidence. *Ann internal med.* 168: 1333-1339.
55. Fujimoto R, Morimoto I, Morita E, Sugimoto H, Ito Y, Eto S. 1994. Androgen receptors, 5 $\alpha$  reductase activity and androgen-dependent proliferation of vascular smooth muscle cells. *J Steroid Biochem Mol Biol.* 50:169–174.

56. Futterman LG and Lemberg L. 2003. The Effects of Aging on Arteries. *Am J Critical Care.* 12: 472-475.
57. Frauchiger B, Schmid HP, Roedel C, Moosmann P, Staub D. 2001. Comparison of carotid arterial resistive indices with intima-media thickness as sonographic markers of atherosclerosis. *Stroke.* 32: 836–841.
58. Gabella G. 1995. Complex structure of the common carotid artery of sheep. *Anat Rec.*243; 376 – 383.
59. Gallardo F, Lloreta J, Garcia F, Moll X, Baro T, Gonzalez La, Morote J, Reventos J, Mogas T. 2009. Immunolocalization of Androgen Receptors, Estrogen Receptors, and Estrogen b Receptors in Experimentally Induced Canine Prostatic Hyperplasia. *J Androl.* 30:240–247.
60. Glagov S. 1994. Intimal hyperplasia, vascular modeling, and the restenosis problem. *Circulation.* 89:2888-2891.
61. Glagov S, Weisenberg E., zarins CK, Stankunavicius R, Kolettis GJ. 1987. Compensatory enlargement of human atherosclerotic coronary arteries. *N Eng J Med.* 316:1371–1375.
62. Glagov S, Zarins C, Giddens DP, Ku DN. 1988. Hemodynamics and atherosclerosis. Insights and perspectives gained from studies of human arteries. *Arch Pathol Lab Med.* 112:1018-31.
63. Gossel M, Rosol M, Malyar NM, Fitzpatrick LA, Beighley PE, Zamir M, Ritman EL. 2003. Functional anatomy and hemodynamic characteristics of vasa vasorum in the walls of porcine coronary arteries. *Anat Rec A Discov Mol Cell Evol Biol.* 272:526-37.

64. Goubergrits L, Affeld K, Fernandez-Britto J, Falcon L. 2002. Geometry of the human common carotid artery. A vessel cast study of 86 specimens. *Pathol Res Pract.* 198:543-51.
65. Greenland P, Abrams J, Aurigemma GP. 2000. "Prevention conference V: beyond secondary prevention identifying high risk patients for primary prevention. Non-invasive test for atherosclerotic burden," Writing Group III. *Circulation.* 101: 16–22.
66. Grobbee DE, Bots ML.1994. Carotid intima-media thickness as indicator of generalized atherosclerosis. *J Intern Med.* 236: 567–573.
67. Gross L, Epstein EZ, Kugel MA. 1934. Histology of coronary arteries and their branches in the human heart. *Am J Pathol* 10: 253–274.
68. Gutterman DD. 1999. Adventitia-dependent influences on vascular function. *American Journal of Physiol heart Circ physiol.* 277: H1265-H1272.
69. Hanke H, Lenz C, Hess B, Spindler KD, Weidemann W. 2001. Effect of testosterone on plaque development and androgen receptor expression in the arterial vessel wall. *Circulation.*103:1382–1385.
70. Hansen F, Mangell P, Sonesson B, Lanne T.1995. Diameter and compliance in the human common carotid artery — variations with age and sex. *Ultrasound in Med and Biol.* 21:1-9.
71. Hansson GK. 2005. Inflammation, Atherosclerosis, and Coronary Artery Disease. *N Engl J Med* 2005; 352:1685-169.
72. Harrison DG, White CW, Hiratzka LF, Doty DB, Barnes DH, Eastham CL, Marcus ML.1984 The value of lesion cross-sectional area determined by quantitative coronary



- angiography in assessing the physiologic significance of proximal left anterior descending coronary arterial stenoses. *Circulation*. 69:1111-1119.
73. Hart RG, Benavente O. 1999. Stroke: Part I. A clinical update on prevention. *Am Fam Physician*. 59:2475-82.
74. Hartman JD. 1977. Structural Changes within the Media of Coronary Arteries Related to Intimal Thickening. *Am J Pathol*. 89:13-34.
75. Hashimura K, Sudhir K, Nigro J, Ling S, Williams MRI, Komesaroff PA, Little PJ. 2005. Androgens Stimulate Human Vascular Smooth Muscle Cell Proteoglycan Biosynthesis and Increase Lipoprotein Binding. *Endocrinology*. 146: 2085-2090.
76. Hodis HN, Mack WJ, Labree L, Selzer RH, Liu CR, Liu CH, Azen SP. 1998. The Role of Carotid Arterial Intima-Media Thickness in Predicting Clinical Coronary Events. *Ann intern med*. 128:262-269.
77. Hort W, Lichti H, Kalbfleisch H, Kohler F, Frenzel H, Milzner-Schwarz U. 1982. The size of human coronary arteries depending on physiological and pathological growth of the heart, the age of the supplying areas and the degree of coronary sclerosis: a postmortem study. *Virchows Arch A Pathol Anat Histol*; 397:37-59.
78. Howard G, Sharrett AR, Heiss G, Evans GW, Chambless Le, Riley Wa Burke Gl. 1993. Carotid artery intimal-medial thickness distribution in general populations as evaluated by B-mode ultrasound. ARIC Investigators. *Stroke*. 24:1297-1304
79. Jadhav UM, Kadam NN. 2001. Carotid intima-media thickness as an independent predictor of coronary artery disease. *Indian Heart J*. 53:458-62.

80. Jarauta E, Mateo-Gallego R, Bea A, Burillo E, Calmarza P, Civeira F. 2010. Carotid intima-media thickness in subjects with no cardiovascular risk factors. *Rev Esp Cardiol.* 63:97-102.
81. Janssen PJ, Brinkmann AO, Boersma WJ, Van Der Kwast TH. 1994. Immunohistochemical detection of the androgen receptor with monoclonal antibody F39.4 in routinely processed, paraffin-embedded human tissues after microwave pre-treatment. *J. Histochem. Cytochem.* 42: 1169 - 1175.
82. Juonala M, Jarvisalo MJ, Torkko NM, Kahonene M, Viikari JSA, Raitakari OT. 2005. Risk Factors Identified in Childhood and Decreased Carotid Artery Elasticity in Adulthood; The Cardiovascular Risk in Young Finns Study. *Circulation.* 112: 1486-1493.
83. Juul A, Skakkebaek NE. 2002. Androgens and the ageing male. *Hum. Reprod.* 8: 423-433.
84. Kalin MF, Zumoff B. 1990. Sex hormones and coronary disease: a review of the clinical studies. *Steroids.* 55:330-52.
85. Kamiya A, Togawa T. 1980. Adaptive regulation of wall shear stress to flow change in the canine carotid artery. *AJP* 239: H14-H21.
86. Kannel WB, Hjortland MC, McNamara PM, Gordon T. 1976. Menopause and the risk of cardiovascular disease. The Framingham Study. *Ann Intern Med.* 85:447-52.
87. Kawasaki T, Sasayama S, Yagi S, Asakawa T, Hirai T. 1987. Non-invasive assessment of the age related changes in stiffness of major branches of the human arteries. *Cardiovascular Research.* 21:678-687.

88. Kimani JK. 1981. Subendothelial fibrillar laminae in the carotid arteries of the giraffe (*Giraffa camelopardalis*). *Cell Tiss Res.* 219: 441-443.
89. Kimani JK. 1983. The structural organization of the tunica intima in the carotid arteries of the giraffe (*Giraffa camelopardalis*). *Afr J Ecol.* 21: 309–315.
90. Kimani JK. 1983. The structural organization of the carotid arterial system of the giraffe (*Giraffa camelopardalis*). *Afr J Ecol.* 21: 317–324.
91. Kimura N, Mizokami A, Oonuma T, Sasano H, Nagara H. 1993 Immunocytochemical localization of androgen receptor with polyclonal antibody in paraffin-embedded human tissues. *J Histochem Cytochem.* 41:671–678
92. Kobayashi N, Sakai T. 1997. Emergence and distribution of intimal muscle cells in the postnatal rat aorta. *Cell. Tiss. Res.*, 289: 487-97.
93. Kotsis V, Stabouli S, Karafillis I, Nilsson P. 2011. Early vascular aging and the role of central blood pressure. *J Hypertens.* 29:1847-53.
94. Krsti RV. 2009. Human Microscopic anatomy. Springer Verlag, Berlin. 49pp.
95. Labropoulos N, Zarge J, Mansour MA, Kang SS, Baker WH. 1998. Compensatory arterial enlargement is a common pathobiologic response in early atherosclerosis. *Am J Surg.* 176:140–143.
96. Lakatta EG, Levy D. 2003. Arterial and Cardiac Aging: Major Shareholders in Cardiovascular Disease Enterprises. Part I: Aging Arteries: A “Set Up” for Vascular Disease. *Circulation.* 107: 139-146.
97. Laurent S, Boutourie P, Lacolley P. 2005. Structural and Genetic Bases of Arterial Stiffness. *Hypertension.* 45: 1050-1055.

98. Lawlor D, Ebrahim S, Whincup P, Sterne J, Papacosta O, Wannamethee G, Dhanjil S, Griffin M, Nicolaides A, Davey S. 2004. Sex differences in body fat distribution and carotid intima media thickness: cross sectional survey using data from the British regional heart study. *J Epidemiol Community Health*. 58: 700–704.
99. Lee HY, Oh BH. 2010. Aging and arterial stiffness. *Circ J*. 74:2257-62.
100. Lerner DJ, Kannel WB. 1986. Patterns of coronary heart disease morbidity and mortality in the sexes: a 26-year follow-up of the Framingham population. *Am J Cardiol*. 111:383–390.
101. Li SJ, Li XY, Li Y. 2008. Regulation of atherosclerotic plaque growth and stability by testosterone and its receptor via influence of inflammatory reaction. *Vasc Pharmacol*. 49: 14-18.
102. Lipman NS, Jackson LR, Trudel LJ, Weis-Garcia F. 2005. Monoclonal Versus Polyclonal Antibodies: Distinguishing Characteristics, Applications, and Information Resources. *ILAR Journal*. 46: 258-268.
103. Liu Y, Ding J, Bush TL, Lonenecker JC, Nieto FJ, Golden SH, Szklo M. 2001. Relative Androgen Excess and Increased Cardiovascular Risk after Menopause: A Hypothesized Relation. *Am. J. Epidemiol*. 154: 489-494.
104. Liu PY, Death AK, Handelsman DJ. 2003. Androgens and cardiovascular disease. *Endocr Rev*. 24: 313–340.
105. Liu PY, Christian RC, Ruan M, Miller VM, Fitzpatrick LA. 2005. Correlating Androgen and Estrogen Steroid Receptor Expression with Coronary Calcification and Atherosclerosis in Men without Known Coronary Artery Disease. *J Clin Endocrinol Metabol*. 90: 1041-1046.

106. Ludwig M, von Petzinger-Kruthoff A, von Buquoy M, Stumpe KO. 2003. Intima media thickness of the carotid arteries: early pointer to arteriosclerosis and therapeutic endpoint. *Ultrasound Med.* 24:162-74.
107. Luo X, Yang Y, Cao T, Li Z. 2011. Differences in left and right carotid intima-media thickness and the associated risk factors. *Clin Radiol.* [Epub ahead of print]
108. Ma R, Wu S, Lin Q. 2005. Homologous Up-Regulation of Androgen Receptor Expression by Androgen in Vascular Smooth Muscle Cells. *Horm Res.* 63: 6 -14.
109. Ma SM, Wei CK, Liang CC, Chou JM, Lee SY. 2011. The Age Correlation of the Carotid Intima-Media Thickness According to Sex and Side in Asymptomatic Subjects. *Acta Neurol Taiwan.* 20:29-34
110. Mainwaring WIP, Mangan FR. 1973. A Study Of The Androgen Receptors In A Variety Of Androgen-Sensitive Tissues. *J Endocrinol.* 59: 121-139
111. Masawa N, Glagov S, Zarins CK. 1994. Quantitative morphologic study of intimal thickening at the human carotid bifurcation, II: the compensatory enlargement response and the role of the intima in tensile support. *Atherosclerosis.* 107:147–155.
112. McCrohon JA, Death AK, Nakhla S, Jessup W, Handelsman DJ, Stanley KK, Celermajer DS. 2000. Androgen receptor expression is greater in macrophages from male than from female donors. A sex difference with implications for atherogenesis. *Circulation.* 101:224–226.
113. McGill HC, Sheridan PJ. 1981. Nuclear uptake of sex steroid hormones in the cardiovascular system of the baboon. *Circ Res.* 48:238-244.

114. McGrath KCY, McRobb LS, Heather AK. 2008. Androgen therapy and atherosclerotic cardiovascular disease. *Vasc Health Risk Manag.* 4: 11–21.
115. McRobb L, Handelsman DJ, Heather AK. 2009. Androgen-induced progression of arterial calcification in apolipoprotein E-null mice is uncoupled from plaque growth and lipid levels. *Endocrinology.* 150:841-8.
116. Milner JS, Moore JA, Rutt BK, Steinman DA. 1998. Hemodynamic of human carotid artery bifurcations: computational studies with models reconstructed from magnetic resonance imaging of normal subjects. *J Vasc Surg.* 28:143-56.
117. Movat HZ, More RH, Haust D. 1958. The Diffuse Intimal Thickening of the Human Aorta with Aging. *Am J Pathol.* 34: 1023–1031.
118. Nakagami H, Morishita R. 2008. Aging of Blood Vessels. *Anti-Aging Med.* 5: 73-77.
119. Nakashima Y, Chen YX, Kinukawa N, Sueishi K. 2002. Distributions of diffuse intimal thickening in Human arteries; preferential expression in atherosclerosis prone arteries from early age. *Virchows Arch.* 441: 279-288.
120. Nakashima Y, Fujii H, Sumiyoshi S, Wight TN, Sueishi K. 2007. Early Human Atherosclerosis: Accumulation of Lipid and Proteoglycans in Intimal Thickenings Followed by Macrophage Infiltration. *Arterioscler Thromb Vasc Biol.* 27:1159-1165
121. Ng MK, Quinn CM, McCrohon JA, Nakhla S, Jessup W, Handelsman DJ, Celermajer DS, Death AK. 2003. Androgens up-regulate atherosclerosis-related genes in macrophages from males but not females: molecular insights into gender differences in atherosclerosis. *J Am Coll Cardiol.* 42:1306-13.

122. Nilsson PM, Lurbe E, Laurent S. 2008. The early life origins of vascular ageing and cardiovascular risk: the EVA syndrome. *J Hypertens.* 26:1049-57.
123. Okada K, Maeda N, Tatsukawa M, Shimizu C, Sawayama Y, Hayashi J. 2004. The influence of lifestyle modification on carotid artery intima-media thickness in a suburban Japanese population. *Atherosclerosis.* 173:329-37.
124. Okeahialam BN, Alonge BA, Pam SD, Puepet FH. 2011. "Carotid Intima Media Thickness as a Measure of Cardiovascular Disease Burden in Nigerian Africans with Hypertension and Diabetes Mellitus,". *Int J Vasc Med.*
125. O'Leary DH, Polak JF, Kronmal RA, Manolio TA, Burke GL, Wolfson SK. 1999. Carotid-Artery Intima and Media Thickness as a Risk Factor for Myocardial Infarction and Stroke in Older Adults. *N Engl J Med.* 340:14-22.
126. Orsi MA, Domeniconi FR, Artoni BSM, Filho GJ. 2006. Carotid arteries in the dog: Structure and histophysiology. *Int. J. Morphol.*, 24:239-244.
127. Osika W, Dangardt F, Montgomery SM, Volkmann R, Gan LM, Friberg P. 2009. Sex differences in peripheral artery intima, media and intima media thickness in children and adolescents. *Atherosclerosis.* 203: 172-177.
128. Özdemir H, Artaş H, Serhatlioğlu S, Oğur E. 2006. Effects of overweight on luminal diameter, flow velocity and intima-media thickness of carotid arteries. *Diagn Interv Radiol;* 12:142-146.
129. Parchami A, Dehkordi RAF, Derakhshan A. 2009. Comparative histomorphometric study of the common carotid artery and its terminal branches in sheep and goats. *Bul J Veç Med.* 12: 165–170.

130. Parchami A and Dehkordi RAF. 2011. Sexual Dimorphism of Sheep Carotid Artery. *Global Veterinaria*. 6: 152-155.
131. Parke WW, Whalen JL, Bungler PC, Settles HE. 1995. Intimal musculature of the lower anterior spinal artery. *Spine*. 20:2073-9.
132. Penn MS, Saidel GM, Chisolm GM. 1994. Relative significance of endothelium and internal elastic lamellae in regulating the entry of macromolecules into arteries in vivo. *Circ. Res*. 74:74-82.
133. Persson J, Formgren J, Israelsson B, Berglund G. 1994. Ultrasound determined intima-media thickness and atherosclerosis: direct and indirect validation. *Arterioscler Thromb*. 14:261-264.
134. Perusquia M, Stallone JN. 2010. Do androgens play a beneficial role in the regulation of vascular tone? Nongenomic vascular effects of testosterone metabolites. *Am J Physiol Heart Circ Physiol*. 298:H1301-7.
135. Pignoli P, Tremoli E, Poli A, Oreste P, Paoletti R. 1986. Intimal plus medial thickness of the arterial wall: a direct measurement with ultrasound imaging. *Circulation*. 74: 1399-1406.
136. Pinto YM, Pinto SJ, Paul M, Merker HJ. 1998. The electron microscopic morphology of the common carotid artery in rats. *Ann Anat*. 180:223-35.
137. Polak JF, Person SD, Wei GS, Godreau A, Jacobs DR, Harrington A, Sidney S, O'Leary DH. 2010. Segment-Specific Associations of Carotid Intima-Media Thickness With cardiovascular Risk Factors The Coronary Artery Risk Development in Young Adults (CARDIA) Study. *Stroke*. 41:9-15



138. Ramirez ML, Azcona CM, Blasco FA, Morreale HFE. 2007. Androgen excess is associated with the increased carotid intima-media thickness observed in young women with polycystic ovary syndrome. *Human Reproduction*. 22:3197-3203.
139. Raitakari OT, Juonala M, Kahonen M, Taittonen L, Laitinen T, Torkko NM, Jarvisalo MJ, Uhari M, Jokinen E, Ronnema T, Akerblom HK, Viikari JSA. 2003. Cardiovascular Risk Factors in Childhood and Carotid Artery Intima-Media Thickness in Adulthood; The Cardiovascular Risk in Young Finns Study. *JAMA*. 290:2277-2283.
140. Reckelhoff JF, Zhang H, Srivastava K, Granger JP. 1999. Gender Differences in Hypertension in Spontaneously Hypertensive Rats, Role of Androgens and Androgen Receptor. *Hypertension*. 34:920-923.
141. Rexrode KM, Ridker PM, Hegener HH, Buring JE, Manson JE, Zee RYL. 2008. Genetic Variation of the Androgen Receptor and Risk of Myocardial Infarction and Ischemic Stroke in Women. *Stroke*. 39:1590-1592.
142. Riley WA, Freedman DS, Higgs NA, Barnes RW, Zinkgraf SA, Berenson GS. 1986. Decreased arterial elasticity associated with cardiovascular disease risk factors in the young Bogalusa Heart Study. *Arteriosclerosis*. 6:253-261.
143. Roach MR. 1970. The static elastic properties of carotid arteries from fetal sheep. *Canadian J Physiol Pharmacol*. 48:695-708.
144. Rocha ME, Wickham LA, Da Silvera LA, Krenzer KL, Yu FS, Toda I, Sullivan BD, Sullivan DA. 2000. Identification of androgen receptor protein and 5 $\alpha$ -reductase mRNA in human ocular tissues. *Br J Ophthalmol*. 84:76-84.

145. Rosfors S, Hallerstam S, Urstad KJ, Zetterling M, Carlström C. 1998. Relationship Between Intima-Media Thickness in the Common Carotid Artery and Atherosclerosis in the Carotid Bifurcation. *Stroke*. 29:1378-1382.
146. Ruan L, Chen W, Srinivasan SR, Sun M, Wang H, Toprak A, Berensin GS. 2009. Correlates of Common Carotid Artery Lumen Diameter in Black and White Younger Adults. *Stroke*. 40: 702-707
147. Rubio I, Yanez R, Gallo G, Almaguer G, Garcia A, Morato T, Chamorro G, Geballos G. 1998. Rapid and possibly nongenomic effects of testosterone on isolated and perfused rat heart. *Proc West Pharmacol Soc*. 41:131–132.
148. Sack MN, Rader DJ, Cannon RO.1994. Oestrogen and inhibition of oxidation of low-density lipoproteins in postmenopausal women. *The lancet*. 343: 269-270.
149. Sader MA, Griffiths KA, McCredie RJ, Handelsman DJ, Celermajer DS. 2001. Androgenic anabolic steroids and arterial structure and function in male bodybuilders. *J Am Col Cardiol*. 37: 224-230.
150. Sader MA, Celermajer DS.2002. Endothelial function, vascular reactivity and gender differences in the cardiovascular system. *Cardiovascular Research*. 53: 597–604.
151. Safar ME, Levy BI, Struijker-Boudier H. 2003. Current perspectives on arterial stiffness and pulse pressure in hypertension and cardiovascular diseases. *Circulation*. 107: 2864–2869.

152. Sajjad Y, Quenby S, Nickson P, Lewis-Jones DI, Vince G. 2004. Immunohistochemical localization of androgen receptors in the urogenital tracts of human embryos. *Reproduction*. 128 331-339
153. Salonen JT, Salonen R. 1991. Ultrasonographically assessed carotid morphology and the risk of coronary heart disease. *Arterioscler Thromb*.11:1245–1249.
154. Samijob SK, Willigersa JM, Barkhuysena R, Kitslaarb PJEHM, Renemanc RS, Brands PJ, Hoeksa APG. 1998. Wall shear stress in the human common carotid artery as function of age and gender. *Cardiovasc Res*. 39: 515-522.
155. Schievink WI, Bjornsson J, Piepgras DG. 1994. Coexistence of fibromuscular dysplasia and cystic medial necrosis in a patient with Marfan's syndrome and bilateral carotid artery dissections. *Stroke*. 25:2492-2496.
156. Schwartz SM, Campbell GR, Campbell JH.1986. Replication of smooth muscle cells in vascular disease. *Circ Res*. 58: 427- 444.
157. Shadwick RE. 1999. Mechanical design in arteries. *J Exp Biol*. 202: 3305–3313.
158. Sharma P, Lohani B, Chataut SP. 2009. Ultrasonographic evaluation of carotid intima-media thickness in hypertensive and normotensive individuals. *Nepal Med Coll J*. 11: 133-135.
159. Sieveking DP, Lim P, Chow RWY, Dunn LI, Bao S, Mcgrarth KCY, Heather AK, Handelsman DJ, Celermajer DS, Ng MKC. 2010. A sex-specific role for androgens in angiogenesis. *JEM*. 207: 2345-352.

160. Simons PCG, Algra A, Bots ML, Grobbee DE, van der Graaf Y. 1999. Common Carotid Intima-Media Thickness and Arterial Stiffness; Indicators of Cardiovascular Risk in High-Risk Patients The SMART Study (Second Manifestations of ARterial disease). *Circulation*. 100:951-957.
161. Sinnathamby CS. 2006. *Last's anatomy regional and applied*. 11<sup>th</sup> edition, Churchill Livingstone, Edinburgh, 354.
162. Sinning C, Wild PS, Echevarria FM, Wilde S, Schnabel R, Lubos E, Herkenhoff S, Bickel C, Klimpe S, Gori T, Münzel TF, Blankenberg S, Espinola-Klein C; Gutenberg-Heart Study. 2011. Sex differences in early carotid atherosclerosis (from the community-based Gutenberg-Heart Study). *Am J Cardiol*. 107:1841-7.
163. Somjen D, Kohen F, Jaffe A, Amir-Zaltsman Y, Knoll E, Stern N. Effects of gonadal steroids and their antagonists on DNA synthesis in human vascular cells. *Hypertension* 1998; 32: 39-45.
164. Standring S, Ellis H, Healy JC, Johnson D, Williams A (Eds). 2004. Vascular supply of the head and neck region, In: *Gray's Anatomy* 39<sup>th</sup> edition. Elsevier Churchill Livingstone, London. 543-551.
165. Stergiopoulos N, Vulliamoz S, Rachev A, Meister JJ, Greenwald SE. 2001. Assessing the homogeneity of the elastic properties and composition of the pig aortic media. *J Vasc Res* 38: 237-246.
166. Tell GS, Howard G, McKinney WM. 1989. Risk factors for site specific extra cranial carotid plaque distribution as measured by B-Mode ultrasound. *J clinic epidemiol*. 42: 551-559.

167. Thomas M, Dadgar N, Aphale A, Harrell JM, Kunkel R, Pratt WB, Lieberman AP. 2004. Androgen receptor acetylation site mutations cause trafficking defects, misfolding, and aggregation similar to expanded glutamine tracts. *J Biol Chem.* 27; 279:8389-95.
168. Timmins LH, Wu Q, Yeh AT, Moore JE, Jr., 2010. Greenwald SE. Structural inhomogeneity and fiber orientation in the inner arterial media. *Am J Physiol Heart Circ Physiol* 298: H1537–H1545.
169. Todd ME, Friedman SM. 1972. The ultrastructure of peripheral arteries during the development of DOCA hypertension in the rat. *Z Zellforsch Mikrosk Anat* 128: 538–554.
170. Tracy RE. 1966. Sex difference in coronary disease: two opposing views. *J Chronic Dis.* 19:1245-51.
171. Urbina EM, Srinivasan SR, Tang R, Bond MG, Kieltyka L, Berenson GS; Bogalusa Heart Study. 2002. Impact of multiple coronary risk factors on the intima-media thickness of different segments of carotid artery in healthy young adults (The Bogalusa Heart Study). *Am J Cardiol.* 90: 953–958.
172. Virmani R, Kolodgie FD, Burke AP, Farb A, Schwartz SM. 2000. Lessons from sudden coronary death. A comprehensive morphological classification scheme for atherosclerotic lesions. *Arterioscler Thromb Vasc Biol.* 20: 1262–1275.
173. Wagenseil JE, Nerurkar NL, Knutsen RH, Okamoto RJ, Li DY, Mecham RP. 2005. Effects of elastin haploinsufficiency on the mechanical behavior of mouse arteries. *Am J Physiol Heart Circ Physiol.* 289: H1209–H1217.

174. Wang HD, Ratsep MT, Chapman A, Boyd R. 2010. Adventitial fibroblasts in vascular structure and function: the role of oxidative stress and beyond. *Canadian J Physiol Pharmacol.* 88: 177-186.
175. Watson WL, Silverstone SM. 1939. Ligature of the common carotid artery in cancer of the head and neck. *Ann Surg.* 109: 1-27.
176. White CW, Wright CB, Doty DB, Hiratza LF, Eastham CL, Harrison DG, Marcus ML. 1984. Does visual interpretation of the coronary arteriogram predict the physiologic importance of a coronary stenosis? *N Engl J Med.* 310:819-824.
177. Wilens SL. 1951. The Nature of Diffuse Intimal Thickening of Arteries. *Am J Pathol.* 27: 825-839.
178. Williams JK, Heistad DD. 1996. The vasa vasorum of the arteries. *J Mal Vasc.* 21:266-9.
179. Williams RI, Ling S, Dawood T, Hashimura K, Dai A, Li H, Liu JP, Funder JW, Shudir K, Komesaroff PA. 2002. Dehydroepiandrosterone inhibits vascular smooth muscle cell proliferation independent of ARs and ERs. *J Clinical End Met.* 87: 176-181.
180. Willekes C, Brands PJ, Willigers JM, Hoeks APG, Reneman RS. 1999. Assessment of local differences in intima-media thickness in the human common carotid artery. *J. Vasc. Res.,* 36: 222-8.
181. Wolinsky H, Glagov S. 1967. A lamellar unit of aortic medial structure and function in mammals. *Circ Res* 20: 99-111.

182. Wu FCW, von Eckardstein A. 2003. Androgens and Coronary Artery Disease. *End Revs.* 24: 183-217.
183. Yamazoe N, Hashimoto N, Kikuchi H, Kang Y, Nakatani H, Hazama F. 1990. Study of the elastic skeleton of intracranial arteries in animal and human vessels by scanning electron microscopy. *Stroke* 21: 765-770.
184. Zarins CK, Giddens DP, Bharadvaj BK, Sottiurai VS, Mabon RF, Glagov S. 1983. Carotid bifurcation atherosclerosis. Quantitative correlation of plaque localization with flow velocity profiles and wall shear stress. *Circ Res.* 53:502-14.
185. Zarins CK, Weisenberg E, Kolettis G, Stankunavicius R, Glagov S. 1987. Differential enlargement of artery segments in response to enlarging atherosclerotic plaques. *J Vasc Surg.* 1988; 7: 386-394.
186. Ziembicka AK, Przewlocki T, Tracz W, Pieniazek P, Musialek P, Sokolowski A. 2005. Gender Differences in Carotid Intima-Media Thickness in Patients with Suspected Coronary Artery Disease. *Am J Cardiol.* 96:1217-1222.

APPENDIX 1

CONSENT FORM.

Study number: \_\_\_\_\_

This form will be used to give a brief summary of the present study to the relatives of the deceased. After the information, relatives who consent for the sections of the carotid artery of the deceased to be used in this study will sign the present form as an approval for the use of the deceased material in the present study.

**Background:** We are carrying out a survey on the carotid artery intimomedial thickness and androgen receptors. We shall use cadaveric material for this study.

**Aim of the study:** This study will be assessing the carotid artery intimomedial thickness and androgen receptors in man.

**Benefits:** Information obtained from this study may help to underscore the anatomy of the carotid arteries among Kenyans. This will guide clinicians when approaching this area which is delicate in surgery among Kenyans.

**Confidentiality:** The name of the deceased shall not be disclosed or used in the study.

**Extent of harvesting:** Only three 5 mm section of the vessel wall shall be harvested for the present study.

**Disposal:** Material that may be useful in the subsequent related study shall be retained. All other material harvested shall be buried at Langata Cemetary after the present study.

I, the undersigned, having been explained to and understood the importance of this study, do allow the cadaver to be enrolled in the study.

---

Name and title

Signature

We, the investigators, having explained in detail the purpose of the study, hereby submit that privacy shall be maintained and only details related to this study will be published.

---

Signature

Date

Contact of the principal investigator: Tel 0721486182

Contact of KNH/UON-ERC Chairman: 0202726300 ext 44102



## CHETI CHA RUHUSA

Nambari ya uchunguzi: \_\_\_\_\_

Fomu hii itatumiwa kuwajulisha jamii ya aliyefariki kuhusu uchunguzi ambao tunanua kufanya. Baada ya maelezo, ikiwa jamii watakubali, watatia sahihi kenye fomu hii kutoa idhini kwa wachunguzi kutumia sehemu ya mishipa ya alieyafriki kwa uchunguzi huu.

**Kitambulisho:** Sisi tunanua kufanya uchunguzi wa mshipa ya damu ya shingo uitwao carotid kwa watu waliofariki.

**Lengo la uchunguzi:** Uchunguzi huu utatuwezesha kutambua maumbile ya mshipa huu haswa katika wakenya kwani umeonyesha mabadiliko mbali mbali.

**Manufaa:** Uchunguzi huu utasaidia madaktari wa upasuaji na madaktari wengine kufanya upasuaji salama haswa katika wakenya wenye maumbile tunayo ya chambua.

**Siri:** Aliyafariki hatatambuliwa kwa jina lake katika uchunguzi huu.

**Kiwango cha uchunguzi:** Sehemu ndogo ya mshipa wake zenye urefu wa milimita tano pekee itachikuliwa katika uchunguzi huu.

**Mazishi:** Sehemu zote zilizo chukuliwa zitazikwa Lang'ata cemetery baada ya uchunguzi huu.

Mimi, nadhibitisha kwamba nimeelezwa na nikafahamu lengo na manufaa ya uchunguzi huu na kwa hiari yangu mwenyewe kuwapa wachunguzi idhini ya kuhusisha mwili wa marehemu katika uchunguzi huu.

---

Jina na Hadhi

Sahihi

Sisi watafiti tunadhibitisha kwamba tumemuelezea mlezi kuhusu uchunguzi huu ipasavyo na kwamba habari inayohusiana na uchunguzi huu tu ndio utakao chapishwa.

---

Sahihi

Tarehe

Nambari ya simu ya: mtafiti mkuu: tel 0721486182

Mwenyekiti wa kamati chunguzi: 020 2726300 ext 44102

0