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Adsorption of Malachite Green from Aqueous Solutions onto Rice Husks: Kinetic and Equilibrium Studies

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Abstract

A study was done to evaluate the removal of a cationic dye from simulated waste water onto rice husks (RH). Spectroscopic methods such as FTIR and SEM/EDX were used for adsorbent characterization. Experimental dependency on solution pH, initial dye concentration, agitation speed, adsorbent-particle size, temperature of the solution and contact time was evaluated. The adsorption data was tested using both Langmuir and Freundlich isotherms. The data fitted well into Langmuir isotherm model with a monolayer adsorption capacity of 6.5 mg/g. Further, the separation factor (R_L) value was less than unity indicating a favorable adsorption process. Adsorption kinetics was determined using pseudo-first-order, pseudo-second-order and intra-particle diffusion models. The results showed that the adsorption of malachite green onto rice husks followed pseudo-second-order model with a determination coefficient of 0.986. This work has revealed that rice husks have a great potential to sequester cationic dyes from aqueous solutions and therefore it can be utilized to clean contaminated effluents.

Keywords

Adsorption, Malachite Green, Rice Husks, Isotherm, Cationic Dye

1. Introduction

Over the years dyes have been used for coloring industrial products such as food, textile, paper, plastics, pharmaceuticals, cosmetics and tannery [1]. Currently, there are over 10,000 dyes which are used globally. They are synthetic and aromatic in nature which makes them more stable and difficult to biodegrade [2]. Consequently, these recalcitrant dyes have continued to cause deleterious effects

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on human health and aquatic ecosystems due to partial or poor treatment methods before disposal. Some dyes like malachite green (MG) and methylene blue (MB) are carcinogenic and therefore it is important to remove them from effluents with respect to both environmental and aesthetical reasons [3] [4].

Adsorption is an attractive and useful method for the removal of dyes from contaminated water samples. Scientists have adopted it as an alternative technology for eliminating noxious pollutants from water [5]. The process has advantages such as simple design with low investment in terms of initial cost [6]. Various adsorbents have been investigated for elimination of dyes from water samples. These adsorbents include activated carbon [7], straw [8] [9], lignocellulosic substrate from agricultural factories [10] and bagasse fly ash [11]. Some adsorbents used in water purification have some drawbacks. For instance, activated carbon (AC) requires regeneration for re-use and it is also cost-intensive [12].

Rice husks (RH) is a by-product of rice milling factory and accounts for about 20% of rice products [13]. Utilization of rice husks as a low-cost adsorbent to eradicate pollutants is attracting attention due to its large quantity in the environment. In this work, efficacy of RH for removal of cationic dye from water was investigated. Dependency of parameters like contact time, dye concentration, pH and adsorbent dosage on the removal of MG on rice husks was investigated. The objectives of the study were to evaluate the prospect to use RH to remove malachite green from contaminated water and to characterize the RH using spectroscopic methods such as FTIR, SEM/EDS and BET surface area.

2. Materials and Methods

2.1. Preparation of the Adsorbent

The rice husks (RH) was obtained from a milling industry in Kenya. It was cleaned with double distilled deionized water to get rid of any adhering dirt. It was air dried and finally dried in the oven (Memmert UM 400, Germany) at 105° C for three days to remove moisture. The RH was then sieved into three different particle sizes (<300, >300 <425, and >425 µm) and kept in airtight containers to be used in the subsequent experiments.

2.2. Preparation of Adsorbate

Malachite green ($C_{52}H_{54}N_4O_{12}$; C.I. 42,000; molecular weight 927.01 g/mol; λ_{620} nm) was purchased from Kobian Chemicals limited, Nairobi, Kenya and used as received. A solution of 1000 ppm was prepared using double distilled deionized water. Serial dilution of the stock solution was done and a calibration curve constructed. High purity grade reagents were used. The chemical structure of malachite green is given in Scheme 1.

2.3. Characterization of Rice Husks (RH)

FT-IR studies were carried out on a Perkin Elmer spectrophotometer with a scanning range of 4000 - 400 cm⁻¹. Pellet technique, which utilizes potassium bromide (KBr) for analysis was adopted. Functional groups in both raw and

Scheme 1. Chemical structure of malachite green.

reacted RH were identified from the spectra obtained. The surface morphology of the RH was done using Tescan Vega 3 XMU, (Czech Republic) scanning electron microscope which was coupled with energy dispersive X-ray spectrophotometer for elemental analysis. Surface area and porosity of the RH were determined using nitrogen adsorption method, (Analyzer (BET) micromeritics ASAP 2020, USA).

2.4. Experimental Design for Batch Adsorption Studies

2.4.1. Effect of Contact Time and Initial Dve Concentration

The above experiments were investigated using 40 mL of the MG dye in 250 mL Erlenmeyer flasks. A dosage of 0.1 g of the rice husks (RH) with particle size of > 300 < 425 μm was used. The flasks were shaken on a Thermolyne orbital shaker at a speed of 200 rpm. Varying concentrations of malachite green dye (2.3 - 9.3 mg/L) were prepared for the investigation of influence of initial dye concentration. Analysis of the supernatant solution of MG was measured using a Shimad-zu UV-Visible spectrophotometer at a wavelength of 620 nm.

2.4.2. Effect of Solution pH

Malachite green (MG) dye solutions with a pH ranging from 3 to 9 were prepared. The pH was adjusted to the desired value using either 0.1 M NaOH or 0.1 M HCl. 0.1 g of the adsorbent in 40 mL of each adjusted MG solution was agitated at 200 rpm at varying time intervals and then the absorbance was measured.

2.4.3. Effect of Agitation Speed

Effect of agitation speed was investigated by using a range of shaking speeds (50 - 250 rpm) for different experiments. 0.1 g of rice husks in 40 mL of the dye solution was agitated on an orbital shaker at a pH of 7. The initial dye concentration was 9.3 mg/L whereas the particle size was $> 300 < 425 \mu m$.

2.4.4. Effect of Adsorbent Particle Size

To evaluate the influence of particle size on the adsorption of malachite green, rice husks (RH) of various meshes (<300, >300 <425, and >425 μ m) was utilized. 0.1 g of RH in 40 mL of dye solution was agitated in 250 mL conical flasks at 200 rpm and pH of 7 at different time intervals. The dye concentration was kept constant at 9.3 mg/L for each experiment. A graph of percentage removal (%) against time in minutes was plotted.

2.4.5. Effect of Temperature

Adsorption of malachite green onto rice husks was investigated at 296.15, 303.15, 313.15 and 323.15 K with initial dye concentration of 9.3 mg/L and adsorbent dosage of 0.1 g. The solution was shaken at 200 rpm at different time intervals. All experiments were done in triplicates and absorbance measured at a wavelength of 620 nm.

The percentage dye removal (%) and amount of dye adsorbed on to Rice husks, q_e (mg/g) was computed using Equations (1) and (2), respectively:

% adsorption =
$$100(C_o - C_e)/C_o$$
 (1)

$$Q_e = (C_o - C_e)V/M \tag{2}$$

where C_o and C_e are initial and equilibrium concentrations (mg/L) of the dye, q_e is amount of dye adsorbed onto the RH at equilibrium (mg/g), V is the volume of the dye used (L) and M is the mass of the adsorbent (g).

3. Results and Discussion

3.1. Characterization of Rice Husks

3.1.1. FTIR Analysis

FT-IR studies for rice husks were carried out using potassium bromide (KBr) pellet technique (**Figure 1**) with scanning range of 4000 - 400 cm⁻¹. The broad peak at 3317.41 - 3900.02 cm⁻¹ stretch indicates presence of -OH and -NH groups. The peaks observed at 2913.68 and 1364.37 cm⁻¹ were due to stretching and bending vibration of C-H bond in methyl groups respectively (Kushwaha *et al.* 2014). The peak at 2520.51 cm⁻¹ is characteristic of -OH from carboxylic acids whereas the stretching around 2014.69 - 2162.07 cm⁻¹ is due to -C = C from alkynes. The peaks between 1629.45 - 1732.02 cm⁻¹ are characteristic of carbonyl group.

The presence of –OH group alongside carbonyl group indicates presence of carboxylic groups in the rice husks. The peak at 1459.11 - 1508.07 cm⁻¹ is characteristic of aromatic groups whereas the strong peak at 1031.69 - 1046.79 cm⁻¹ is due to C-O bending. The –OH, –NH, carbonyl and carboxyl groups are vital sorption sites [14]. Some peaks shifted after adsorption (3317.41 to 3341.91, 3730.52 to 3788.79, 3900.02 to 3911.20, 2520.51 to 2580.85 and 1629.45 to 1635.73) whereas other peaks disappeared after adsorption (3548.75 and 3511.30). This was a clear indication that adsorption of MG onto rice husks (RH) had taken place and new bonds were formed between MG and RH.

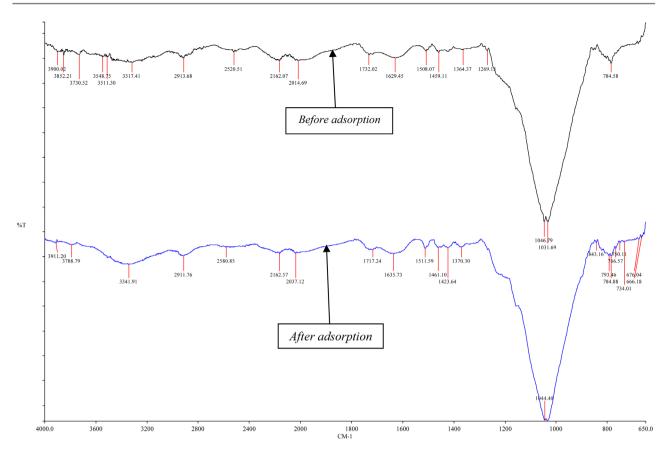


Figure 1. FTIR spectrum before and after adsorption of malachite green.

3.1.2. SEM/EDX Analysis

The surface morphology of rice husks was done using scanning electron microscope (SEM) at 150 × magnification (Figure 2 & Figure 3). Figure 2 is a micrograph of rice husks (RH) before adsorption process. It was found that the adsorbent has an irregular porous surface which may be responsible for malachite green adsorption. Figure 3 represents the reacted rice husks with malachite green after adsorption. It is evident that after removal of the dye, the surface of the RH was smooth implying that the unfilled adsorption sites of the adsorbent were covered by the dye and therefore adsorption had taken place. Elemental analysis of the rice husks (Table 1) indicated substantial amounts of oxygen and silicon (42.8% and 51.82% respectively). These elements are essential in formation of bonds during adsorption process.

3.1.3. BET Surface Area and Porosity

The BET (Brunauer, Emmett and Teller) single point (P/P_o) analysis for the specific surface area was 9.8 m²/g whereas the pore volume was 0.068671 cm³/g. Pore size distribution was 280.3962 Å. These parameters affect the extent in which adsorption process takes place.

3.2. Batch Experiments

3.2.1. Effect of Contact Time

The effect of contact time was evaluated with dye concentration of 9.3 mg/L,

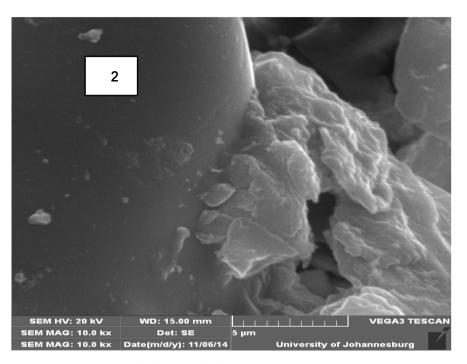


Figure 2. SEM micrograph of raw rice husks.

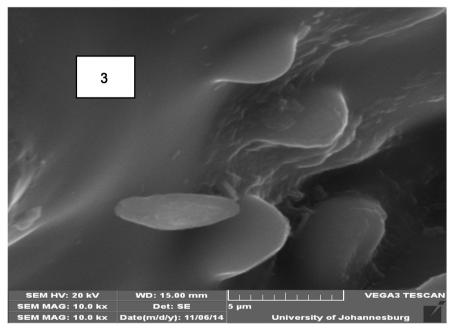


Figure 3. SEM micrograph of reacted rice husks.

adsorbent dosage of 0.1 g, a solution volume of 40 mL and agitation speed of 200 rpm. The results revealed that the uptake of the dye was rapid at the initial stages of adsorption reaction (30 minutes) and thereafter the adsorption was slow as it approached equilibrium (Figure 4). The above trend of adsorption is ascribed to substantial amount of vacant surface sites accessible for adsorption at the initial stage. However, as the reaction approached equilibrium all the sites were already occupied by the dye hence the adsorption was slow.

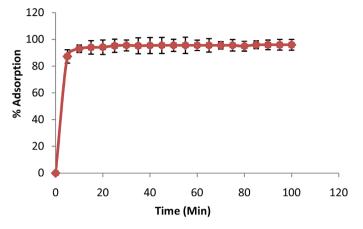


Figure 4. Effect of contact time on malachite green removal by RH.

Table 1. Elemental analysis of rice husks.

Element	Weight%	Atomic%
O	42.80	57.42
Mg	0.22	0.19
Al	0.59	0.47
Si	51.82	39.61
P	0.44	0.31
S	0.31	0.20
Ca	2.08	1.12
Mn	0.45	0.18
Fe	1.28	0.49
Totals	100.00	

The curve in **Figure 4** indicates probable monolayer coverage of dye on the surface of rice husks. Rajesh *et al.* [15] recently reported similar results for removal of malachite green on *Hydrilla Verticilla* biomass. The same trend was also reported by Ahmad and Kumar [16] for the adsorption of malachite green onto treated ginger waste.

3.2.2. Effect of Initial Dye Concentration

The influence of initial concentration of malachite green (MG) onto rice husks was investigated with dye concentrations ranging from 2.3 to 9.3 mg/L and adsorbent dosage of 0.1 g. The percentage (%) removal decreased with increase of initial dye concentration of MG (**Figure 5**). The % decrease in adsorption is ascribed to saturation of the active binding sites of the rice husks at higher concentrations of malachite green.

3.2.3. Effect of pH

The pH of the solution affects the surface charge of the adsorbent and degree of ionization of the dye [17]. In the present study, the pH values were varied from 3 - 9 (**Figure 6**) while the other parameters were kept constant. 0.1 M HCl and 0.1 M NaOH solutions were used to correct the pH to the desired value. The percentage (%) adsorption of the dye increased with increase in pH up to an

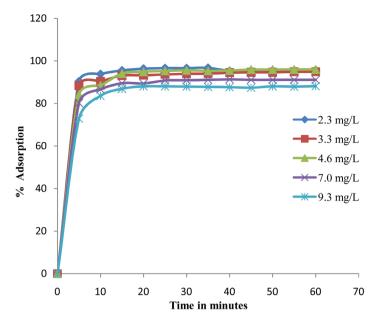


Figure 5. Effect of initial dye concentration on adsorption of MG onto rice husks.

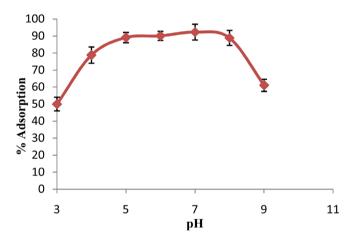


Figure 6. Effect of pH on adsorption of malachite green onto rice husks.

optimal value of pH 7.

For pH values greater than 7, the % adsorption of the dye decreased hence the optimal pH of 7 was selected for subsequent experiments. The low adsorption of malachite green (MG) onto rice husks at pH 3 - 6 could be attributed to H⁺ ions in excess that compete with the dye cations for the adsorption sites [18].

In the pH range of 8 - 9 the adsorption decreases probably due to formation of soluble hydroxyl complexes.

3.2.4. Effect of Agitation Speed

The study was conducted by varying the speed from 50 to 250 rpm on a Thermolyne orbital shaker. Adsorbent dosage was kept constant at 0.1 g, particle size used was >300 <425 μ m, pH of 7 and the volume of the dye solution was 40 mL. The maximum removal (93.4%) of malachite green onto rice husks was achieved

at 200 rpm and then the adsorption decreased when the agitation speed was increased to 250 rpm (**Figure 7**). Increasing the agitation speed decreases the boundary of the transfer of dye molecules from the bulk solution to adsorbent surface.

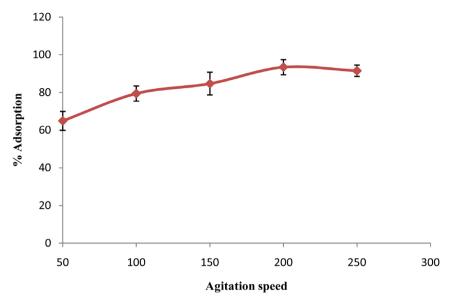


Figure 7. Effect of agitation speed on adsorption of MG onto rice husks.

3.2.5. Effect of Adsorbent Particle Size on Dye Removal

The influence of adsorbent particle size on adsorption of malachite green (MG) was tested using three different meshes (<300, >300 <425, and >425 μm). The adsorbent dosage was 0.1 g; volume of the dye solution was 40 mL, pH of 7 and shaking speed of 200 rpm. The percentage adsorption of MG decreased from 98 to 95% on increasing the particle size from >300 to >425 μm (Figure 8). The higher adsorption of MG onto smaller particle size of the adsorbent was attributed to increased accessibility of binding sites due to increased surface area for bulk adsorption of the dye.

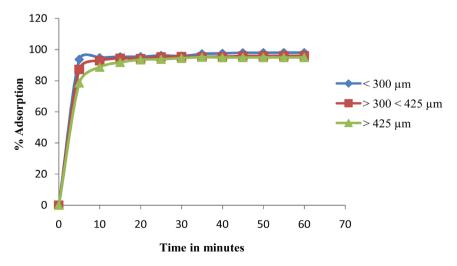


Figure 8. Effect of particle size of RH on malachite green removal.

3.2.6. Effect of Temperature on Dye Adsorption

Experiments were carried out at a temperature range of 296.15 to 323.15 K. The percentage adsorption of malachite green (MG) decreased (97.3% to 79.0%) with increasing temperature implying that the process was exothermic, **Figure 9**. This could be ascribed to weakening of adsorption forces between the active sites of the adsorbent and adsorbate. Similar results were reported recently in the removal of malachite green using sugarcane baggasse [19].

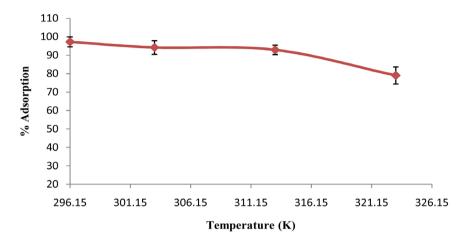


Figure 9. Effect of temperature on adsorption of malachite green dye.

4. Equilibrium Isotherms

4.1. Langmuir Adsorption Isotherm

The Langmuir equation is commonly expressed as follows [20]:

$$C_{\alpha}/Q_{\alpha} = 1/K_{I}Q_{m} + C_{\alpha}/Q_{m} \tag{3}$$

where Q_m is monolayer adsorption capacity (mg/g), K_L is Langmuir isotherm constant. The values of Q_m and K_L can be calculated by plotting C_e/Q_e against C_e (**Figure 10**). The essential characteristics of a Langmuir isotherm model can be expressed in terms of dimensionless constant separation factor, R_L [17] which is defined by

$$R_L = 1/(1 + K_L C_o) \tag{4}$$

The value of R_L indicate the type of bio sorption isotherm to be either unfavorable ($R_L > 1$), linear ($R_L = 1$), favorable ($0 < R_L < 1$) or irreversible ($R_L = 0$).

The Langmuir isotherm model gave a good fit to the equilibrium adsorption data, with r^2 of 0.972 compared to Freundlich isotherm which gave determination coefficient of 0.93. It gave adsorption capacity of 6.5 mg/g which was slightly higher than 1.484 mg/g value which was reported recently by Chanzu and coworkers although their r^2 was 0.704; a value which is lower than what is reported in this study, [17]. This suggested that the removal of malachite (MG) dye followed monolayer coverage onto homogeneous rice husks surface and therefore interaction between dye molecules was neglible [21]. In the present work, the R_L value was found to be 0.61 indicating favorable adsorption (**Table** 2) whereas the K_L value was 0.28 L·mg⁻¹.

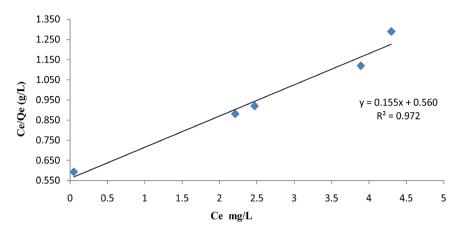


Figure 10. Langmuir adsorption isotherm for adsorption of MG onto rice husks.

4.2. Freundlich Adsorption Isotherm

The Freundlich equation was applied for the analysis of the initial dye concentration data obtained from equilibrium studies. The equation assumes a heterogeneous adsorption surface and active sites with different energy. The equation is expressed as follows [4]:

$$q_e = K_f C_e^{1/n} \tag{5}$$

The linear form of Equation (5) is given in the following equation

$$\ln q_e = \ln K_f + 1/n \ln C_e \tag{6}$$

where K_f and n are adsorption capacity and intensity respectively and their values can be obtained from intercept and the slope of the graph of $\ln q_c$ against $\ln C_c$ (**Figure 11**). Their corresponding values were 0.43 and 27 respectively (**Table 2**). The results from Freundlich isotherm model gave a lower adsorption capacity (0.43 mg·g⁻¹) of malachite green onto rice husks (RH) compared to that of Langmuir model of 6.5 mg/g. Further, the results demonstrated a better fitting of Langmuir isotherm model than Freundlich model.

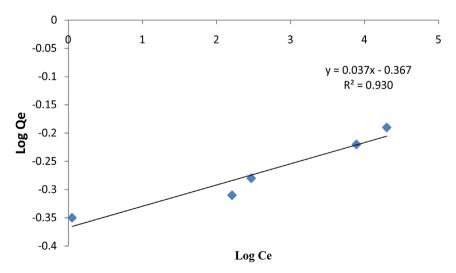


Figure 11. Freundlich adsorption isotherm for removal of MG onto rice husks.

Table 2. Langmuir and Freundlich isotherm parameters for adsorption of MG onto rice husks.

Langmuir isotherm model constants			Freundlich isotherm model constants			
$Q_m (\mathrm{mg} \cdot \mathrm{g}^{-1})$	$K_L (L \cdot mg^{-1})$	R_L	R^2	$K_F(\mathrm{mg}{\cdot}\mathrm{g}^{-1})$	п	R^2
6.5	0.28	0.61	0.972	0.43	27	0.93

5. Kinetic Studies

5.1. Pseudo-First-Order Kinetics on Malachite Green Adsorption

The kinetic data was fitted into Lagergren pseudo-first-order rate equation which is expressed as [22] [23]:

$$Log(q_e - q_t) = log q_e - k_1 t / 2.303$$
 (7)

where q_e and q_t are the adsorption capacities at equilibrium and time t whereas k_1 (min⁻¹) is the rate constant for the adsorption process. The values of q_e and k_1 are given in **Table 3**. Adsorption of malachite green (MG) onto rice husks biomass did not follow pseudo-first-order kinetics model since there was no agreement between the experimental (Exp q_e) and calculated (q_e , cal) adsorption capacities (**Table 3**). Furthermore, the coefficient of determination (r^2) value was relatively lower (0.78) compared to pseudo-second-order value of 0.986 (**Figure 12**).

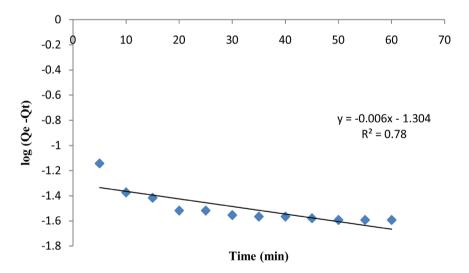


Figure 12. Pseudo-first-order adsorption kinetics of MG onto rice husks.

5.2. Pseudo-Second-Order Kinetics on Malachite Green Adsorption

The Lagergren pseudo-second order kinetics equation is expressed in a linear form as shown below [16] [24] [25]:

$$t/q_t = 1/k_2 q_e^2 + 1/q_e t (8)$$

where the equilibrium adsorption capacity (q_e) and the second order constant k_2 (g·mg⁻¹·min⁻¹) can be determined from the slope and intercept of plot of t/q_t versus t (Figure 13). The experimental data from this work fitted well into

pseudo-second-order model with a determination coefficient (R^2) of 0.986). There was good relationship between experimental and calculated value of q_e (**Table 3**) indicating that the adsorption experiment followed pseudo-second-order than the pseudo-first-order kinetics model. Similar results were reported for removal of methylene blue from aqueous solutions using *Eichhornia Crassipes* [24]. The second order constant, k_2 was 4.3 g·mg⁻¹·min⁻¹.

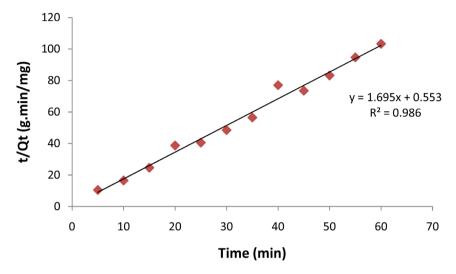


Figure 13. Pseudo-second-order adsorption kinetics of MG onto rice husks.

Table 3. Kinetic parameters for adsorption of MG onto rice husks.

	Pseudo-first order			Pseudo-second-order		
$\operatorname{Exp} q_{\mathfrak{S}} (\operatorname{mg} \cdot \operatorname{g}^{-1})$	q _e , cal. (mg·g ⁻¹)	K_1 (min ⁻¹)	R^2	$q_{ m e}$, cal. (mg·g $^{-1}$)	$K_2 (g \cdot mg^{-1} \cdot min^{-1})$	R^2
0.622	0.05	0.014	0.78	0.625	4.3	0.986

5.3. Mechanism of Adsorption of Malachite Green onto Rice Husks Weber-Morris Intraparticle Diffusion Model

The above model was applied to illustrate existence of competitive adsorption processes in the removal of malachite green onto rice husks. The following expression is used to describe the model [4] [26]:

$$q_t = k_{id} t^{0.5} + c (9)$$

where k_{id} is the intra particle diffusion constant (mg·g⁻¹·min^{0.5}) and c represents the boundary layer thickness. The values of k_{id} and c were calculated from the slope and intercept of the graph (**Figure 14**) of Q_t against \sqrt{t} in minutes respectively. From the graph, K_{id} was found to be 0.057 mg·g⁻¹·min^{0.5} and c was 0.303 with a determination coefficient (r^2) of 0.966. Since the line of the graph (**Figure 14**) did not pass through the origin, it indicates that intra particle diffusion was not the only process controlling the adsorption of malachite green dye onto the rice husks.

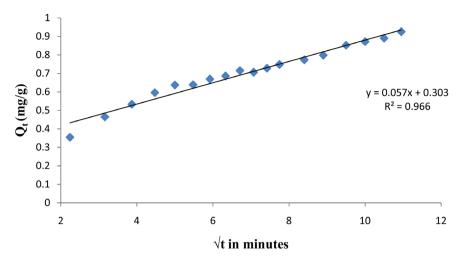


Figure 14. Intraparticle diffusion graph for adsorption malachite green.

6. Conclusion

The results obtained from this work indicate that rice husks (RH) can be used as a low-cost adsorbent for sequestering malachite green from aqueous solutions such as waste waters. The characterization studies with scanning electron microscope (SEM) and Fourier transform infra red spectroscopy (FTIR) indicated presence of sufficient pores and functional groups on RH, which can be used in the adsorption process. Further, this study revealed that batch adsorption is influenced by factors such as initial dye concentration, pH of solution, agitation speed, contact time and temperature. Maximum dye removal of 95.7% was achieved using initial dye concentration of 2.3 g/L and pH of 7. The Langmuir adsorption isotherm gave the best fit to the experimental data, suggesting monolayer adsorption on a homogeneous surface. The adsorption kinetics followed the pseudo-second-order model with a determination coefficient (r^2) of 0.986. The percentage adsorption of malachite green (MG) decreased from 97.3% to 79.0% with increasing temperature (296.15 - 323.15 K), indicating an exothermic process. The line of intraparticle diffusion graph did not pass through the origin and therefore there were other mechanisms controlling the adsorption of MG onto rice husks.

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References

[1] Zhang, H., Tang, Y., Liu, X., Ke, Z., Su, X., Cai, D., Wang, X., Liu, Y., Huang, Q. and Yu, Z. (2011) Improved Adsorptive Capacity of Pine Wood Decayed by Fungi *Poria cocos* for Removal of Malachite Green from Aqueous Solutions. *Desalination*, **274**,

- 97-104. https://doi.org/10.1016/j.desal.2011.01.077
- [2] Farajzadeh, M.A. and Fallahi, M.R. (2005) Study of Phenolic Compounds Removal from Aqueous Solution by Polymeric Sorbent. *Journal of the Chinese Society*, **52**, 295-301. https://doi.org/10.1002/jccs.200500045
- [3] Kushwala, A.K., Gupta, N. and Chattopadhyaya, M.C. (2010) Adsorption of Malachite Green Dye on Chemically Modified Silica Gel. *Journal of Chemical and Pharmaceutical Research*, **2**, 34-45.
- [4] Kushwaha, A.K., Gupta, N. and Chattopadhyaya, M.C. (2014) Removal of Cationic Methylene Blue and Malachite Green from Aqueous Solution by Waste Materials of *Daucuscarota. Journal of Saudi Chemical Society*, 18, 200-207. https://doi.org/10.1016/j.iscs.2011.06.011
- [5] Gupta, V.K., Jain, C.K., Chandra, S. and Agarwal, S. (2002)) Removal of Lindane and Malathion from Waste Water Using Bagasse fly Ash, a Sugar Industry Waste. *Water Research*, **36**, 2483-2490. https://doi.org/10.1016/S0043-1354(01)00474-2
- [6] Srivastava, B., Jhelum, V., Basu, D. and Patanjali, P.K. (2009) Adsorbents for Pesticide Uptake from Contaminated Water. A Review. *Journal of Scientific and Industrial Research*, 68, 839-850.
- [7] Sun, D., Zhang, Z., Wang, M. and Wu, Y. (2013) Adsorption of Reactive Dyes on Activated Carbon Developed from *Enteromorphaproriera*. *American Journal of Analytical Chemistry*, **4**, 17-26. https://doi.org/10.4236/ajac.2013.47A003
- [8] Chen, X., Chen, X., Wan, X., Weng, B. and Huang, Q. (2010) Water Hyacinth (*Eichhrniacrassipes*) Waste as an Adsorbent for Phosphorus Removal from Swine Waste Water. *Bioresource Technology*, 101, 9025-9030. https://doi.org/10.1016/j.biortech.2010.07.013
- [9] Valipour, A., Raman, V.K. and Ghole, V.S. (2011) Phytoremediation of Domestic Waste Water Using *Eichhorniacrassipes. Journal of Environmental Science and En*gineering, 53, 183-190.
- [10] Boudesocque, S., Guillon, E., Aplincourt, M., Martel, F. and Noël, S. (2008) Use of Low-Cost Biosorbent to Remove Pesticides from Waste Water. *Journal of Environ*mental Quality, 37, 631-638. https://doi.org/10.2134/jeq2007.0332
- [11] Gupta, V. and Ali, I. (2001) Removal of DDD and DDE from Waste Water Using Bagasse Fly Ash, a Sugar Industry Waste. *Water Research*, **35**, 33-40. https://doi.org/10.1016/S0043-1354(00)00232-3
- [12] Chiung, F.C., Ching, Y.C., Kuo, E.H., Shu, C.L. and Wolfgang, H. (2008) Adsorptive Removal of the Pesticide Methomylusing Hyper Cross Linked Polymers. *Journal of Hazardous Material*, **155**, 295-304. https://doi.org/10.1016/j.jhazmat.2007.11.057
- [13] Mansaray, K.G. and Ghaly, A.E. (1998) Thermo Gravimetric Analysis of Rice Huskss in an Air Atmosphere. *Energy Source*, 20, 653-663. https://doi.org/10.1080/00908319808970084
- [14] Poojari, A.C., Maind, S.D. and Bhalerao, S.A. (2015) Effective Removal of Cr (VI) from Aqueous Solutions Using Rind of Orange (*Citrus Sinensis*) (L.) Osbeck. *International Journal of Current Microbiology and Applied Sciences*, **4**, 653-671.
- [15] Rajesh, K.R., Rajasimman, M., Rajamohan, N. and Sivaprakash, B. (2010) Equilibrium and Kinetic Studies on Sorption of Malachite Green Using *Hydrilla verticillata* biomass. *International Journal of Environmental Research*, **4**, 817-824.
- [16] Ahmed, R. and Kumar, R. (2010) Adsorption Studies of Hazardous Malachite Green onto Treated Ginger Waste. *Journal of Environmental Management*, 91, 1032-1038. https://doi.org/10.1016/j.jenvman.2009.12.016
- [17] Chanzu, H.A., Onyari, J.M. and Shiundu, P.M. (2012) Biosorption of Malachite

- Green from Aqueous Solutions onto Polylactide/Spent Brewery Grains Films: Kinetic and Equilibrium Studies. *Journal of Polymers and the Environment*, **20**, 665-672. https://doi.org/10.1007/s10924-012-0479-5
- [18] Hameed, B.H. and El-Khaiary, M.I. (2008) Malachite Green Adsorption by Rattan Sawdust: Isotherm, Kinetic and Mechanism Modeling. *Journal of Hazardous Materials*, **159**, 574-579. https://doi.org/10.1016/j.jhazmat.2008.02.054
- [19] Sharma, N. and Nandi, B.K. (2013) Utilization of Sugarcane Baggase, an Agricultural Waste to Remove Malachite Green Dye from Aqueous Solutions. *Journal of Materials and Environmental Science*, **4**, 1052-1065.
- [20] Langmuir, I. (1916) The Constitution and Fundamental Properties of Solids and Liquids. *Journal of the American Chemical Society*, 38, 2221-2295. https://doi.org/10.1021/ja02268a002
- [21] Ofomaja, A.E. and Ho, Y.S. (2008) Effect of Temperatures and pH on Methyl Violet Biosoption by Mansonia Wood Sawdust. *Bioresource Technology*, 99, 5411-5417. https://doi.org/10.1016/j.biortech.2007.11.018
- [22] Labidi, N.S. and Kacemi, N.E. (2016) Adsorption Mechanism of Malachite Green onto Activated Phosphate Rock: A Kinetics and Theoretical Study. *Bulletin of Environmental Studies*, 1, 69-74.
- [23] Khope, R.U. and Gawande, N.J. (2015) Kinetic Models for the Adsorption of Cobalt from Aqueous Phase Using Granular Activated Carbon. *Journal of Chemical and Pharmaceutical Research*, **7**, 551-556.
- [24] Wanyonyi, W.C., Onyari, J.M. and Shiundu, P.M. (2013) Adsorption of Methylene Blue Dye from Aqueous Solutions Using *Eichhrnia crassipes. Bulletin of Environ*mental Contamination and Toxicology, 91, 362-366. https://doi.org/10.1007/s00128-013-1053-0
- [25] Ho, Y.S. and McKay, G. (1999) Pseudo-Second Order Model for Sorption Processes. *Process Biochemistry*, **34**, 451-465. https://doi.org/10.1016/S0032-9592(98)00112-5
- [26] Weber, W.J. and Morris, J.C. (1963) Kinetics of Adsorption on Carbon from Solutions. *Journal of the Sanitary Engineering Division*, **89**, 31-39.



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