

## Review Article

# A Systematic Review of Medicinal Plants of Kenya used in the Management of Bacterial Infections

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Kenya's vision 2030 partly aims at ensuring adequate health care for all, and the integration of traditional healthcare practices into the national healthcare system would present a more rapid alternative towards the realization of universal health coverage in Kenya. Currently, research on Kenyan medicinal plants with potential antibacterial activity remains vastly fragmented across numerous literature studies and databases; thus, it is imperative to collate and appraise these data for the ease of future research and possible clinical application. *Objective.* This review aims at exploring and compiling research evidence on medicinal plants used in the management of bacterial infections in Kenya, with a focus on their efficacy and safety. *Methodology.* A comprehensive web-based systematic review using the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines was executed to highlight the Kenyan medicinal plants used for the management of bacterial infections in Kenya. This review includes studies published until January 2021 from the PubMed, Science Direct, AJOL, and Google Scholar databases. *Results.* A total of 105 Kenyan medicinal plants belonging to 43 families have their *in vitro* activity against various human pathogenic bacteria evaluated. Plants from the Lamiaceae, Rutaceae, and Fabaceae families were the most commonly studied. *Aloe secundiflora*, *Toddalia asiatica*, *Senna didymobotrya*, *Warburgia ugandensis*, *Tithonia diversifolia*, *Fuerstia africana*, *Olea africana*, and *Harrisonia abyssinica* were the plants frequently evaluated within Kenya. The plants with the strongest antimicrobial activities were *Toddalia asiatica*, *Hagenia abyssinica*, *Ocimum gratissimum*, *Harrisonia abyssinica*, *Senna didymobotrya*, *Olea Africana*, *Camellia sinensis*, and *Tamarindus indica*. *Conclusion.* Based on a published work, it is evident that traditional medicine is seemingly an acceptable and efficient system among Kenyan communities in the management of bacterial infections. Kenya's rich biodiversity with diverse secondary metabolites presents a promising source of new therapeutic alternatives with possibly different mechanisms of action against bacteria.

## 1. Introduction

Despite the remarkable investment in health care witnessed over the past decade, microbial infections remain a major threat to human and animal health and are a cause of morbidity and mortality especially in low- and middle-income countries (LMICs). The rising cases of antibiotic resistance present a major health problem globally, and there is an immediate need for strategies to manage it as it relentlessly compromises the effectiveness of antimicrobial therapy and increases the threat of therapeutic failure [1–3]. Due

to an inefficient antimicrobial resistance (AMR) surveillance system, the exact liability of AMR in Kenya is indefinite although cases such as reduced susceptibility of community-acquired pneumococci, *Vibrio cholera* outbreaks, and methicillin-resistant *Staphylococcus aureus* (MRSA) from hospitalized patients have been reported [4].

Herbalism is the most preferred form of traditional medicine and is highly lucrative in the international market with annual sales ranging from US dollar 5 billion in Western Europe to US dollar 14 billion in China [5]. In Africa, herbal products are available in most markets in the

urban centers and rural areas [6]. Irrespective of the accessibility to modern medicines, various communities in Kenya (either deliberately or due to economic limitations) utilize medicinal plants for the management of microbial infections and other diseases; thus, various legislations are actively being formulated to regulate this practice [7]. Presently, there are over 400 plant species used for the management of common diseases in East Africa documented in several ethnobotanical [8–10].

As a developing nation with numerous healthcare challenges such as the high costs of medications, Kenya needs to grow its scientific base and create logical and effective solutions to manage them. Laboratory investigations and various clinical trials have often suggested the positive effects of phytomedicines both *in vivo* and *in vitro*; however, there has been little systematic appraisal of their benefits [11]. Due to their unrivaled chemical diversity, plants offer the infinite potential for innovative and effective antimicrobial agents, but there is the scantiness of information in regard to their efficacy and their safety levels [12]. Critical consideration to the prospect of producing pharmaceutical products using local raw materials is a worthy endeavor to ensure the affordability of drugs. In a bid to provide herbal practitioners and consumers with insight, this study primarily aimed at evaluating the bioactivity of Kenyan medicinal plants useful in the management of bacterial infections.

## 2. Materials and Methods

A comprehensive web-based systematic review using the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines on identification, screening, eligibility, and inclusion was executed to highlight the medicinal plants used for the management of bacterial infections in Kenya. This review covers published literature from 1994 up to January 2021 obtained from the PubMed, Science Direct, African Journals Online (AJOL), and Google Scholar databases. Grey literature [13] from the local university repositories and conference proceedings were also included in this review [14].

The literature search was performed using search terms identified from previous similar reviews. The Boolean search operators (AND and OR) were used to effectively combine the search terms [15]. The following search terms were used: Kenya AND antimicrobial plants OR Kenyan AND antimicrobial plants. Kenyan AND antibacterial plants OR Kenyan AND antibacterial plants. Kenya AND traditional medicine AND antimicrobial plants OR Kenyan AND traditional medicine AND antimicrobial plants, Kenya AND traditional medicine AND antibacterial plants OR Kenyan AND traditional medicine AND antibacterial plants, Kenya AND ethnopharmacological AND antimicrobial plants OR Kenyan AND ethnopharmacological AND antimicrobial plants and Kenya AND ethnopharmacological AND antibacterial plants OR Kenyan AND ethnopharmacological AND antibacterial plants [16].

Screening of search outputs was performed in two stages. First, the title and abstract of identified journal articles/theses were overviewed based on PICO (Participants

Intervention Comparison and Outcomes) and the studies classified as “yes” or “no” based on the information provided by the title and abstract. Thereafter, suitable articles/theses were downloaded and critically assessed for inclusion in the review [16].

The studies eligible for inclusion were limited to the English language. The assessment of eligibility of studies was performed by at least two people, independently, using the Critical Appraisal Skills Programme (CASP) appraisal checklist as a guide [16]. This study excluded research data from papers with poor methodology and retracted studies. The quality of the papers was assessed based on study design, description of the subject, method and assay, variables assessment, control groups, and data collection. To minimize bias, data extraction from selected study reports was independently performed by two reviewers and any disagreements resolved through discussion with the third reviewer [17].

## 3. Results

The study included research data from the pharmacological assays/ethno-medicinal studies reporting on Kenyan medicinal plants used for the treatment of bacterial infections. The initial database search identified a total of 105, 157 articles. After removing the duplicates ( $n = 15000$ ), 89857 studies were excluded based on the title and abstract. Three hundred (300) full-text articles were assessed for eligibility, from which 211 were excluded based on scope, methodological approach, and very little/no bioactivity reported. A total of seventy-nine (79) studies regarding the *in vitro* antibacterial activity of Kenyan medicinal plants were ultimately included in the review. No *in vivo* studies within Kenya on Kenyan medicinal plants with antibacterial activity were found.

Data collected included herbal plant name, plant family, part of plant used for extraction, extraction/preparation method, concentrations of extracts, bacteria species, data on reported activity, toxicity, exposure time, geographical information, the year of publication, and the first author (Table 1). A total of 105 medicinal plants from 43 families were studied for *in vitro* activity against various human pathogenic bacteria. Plants from Lamiaceae, Rutaceae, and Fabaceae families were the most common (Table 1).

*Aloe secundiflora* (5), *Toddalia asiatica* (5), *Senna didymobotrya* (5), *Warburgia ugandensis* (5), *Tithonia diversifolia* (4), *Fuerstia africana* (4), *Olea africana* (4), and *Harrisonia abyssinica* (4) were the plants frequently evaluated within Kenya. The plants with the strongest antimicrobial activities were *Toddalia asiatica*, *Hagenia abyssinica*, *Ocimum gratissimum*, *Harrisonia abyssinica*, *Conyza sumatrensis*, *Senna didymobotrya*, *Aloe secundiflora*, *Olea Africana*, *Vernonia glabra*, *Camellia sinensis*, *Tetradenia riparia*, and *Tamarindus indica* as they exhibited high mean inhibition zone values or low minimum inhibitory concentration (MIC) values. The zones of inhibition (ZOIs) were interpreted as low activity (1 mm–6 mm), moderate activity (7 mm–10 mm), high activity (11 mm–15 mm), and very high activity (>16 mm) (Zaidan et al., 2005). The frequently analyzed plant parts were leaves (37%), bark/stem bark (47%), fruits/seeds (5), pods (1%), and roots (24%).

TABLE 1: Systematic review of Kenyan antibacterial medicinal plants.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Candidiasis, diarrhea, sore throat, and wound healing</i>	The ethanol leaf extract exhibited a ZOI of $17.0 \pm 0.8$ mm compared to $8.4 \pm 0.7$ mm (erythromycin) and $8.0 \pm 0.8$ mm (gentamycin) against <i>Streptococcus pneumoniae</i>	Disk diffusion method	Eastern Kenya	None reported	[18]		
<i>Wounds, appetizer, and malaria</i>	The methanol leaf extract (100 mg/ml) exhibited a ZOI of $17 \pm 1$ mm against <i>Staphylococcus aureus</i> , $18 \pm 2$ mm, <i>Bacillus subtilis</i> , $17 \pm 2$ mm, <i>K. pneumoniae</i> , and $19 \pm 2$ mm against <i>E. coli</i> . The methanol leaf extract (1 g/ml) exhibited a ZOI ( $13.0 \pm 0.17$ mm) compared to ( $25.0 \pm 1.06$ ) ciprofloxacin against <i>S. aureus</i> , ZOI ( $17.0 \pm 1.38$ mm) compared to ( $20 \pm 2.47$ mm) against <i>E. coli</i> , and ZOI ( $18 \pm 0.35$ mm) compared to ( $22.0 \pm 1.06$ mm) ciprofloxacin against <i>E. faecalis</i>	Agar well assay	Department of Biological Sciences, Egerton University	Not reported	[19]		
<i>Aloe secundiflora Engl. (Asphodelaceae)</i>	Leaves	Disk diffusion method	Kenyatta University Arboretum	Not reported	[20]		
<i>Stomachache, polio, malaria, and chest problems</i>	The methanol leaf extract had an MIC of $9.375$ mg/mL against <i>P. aeruginosa</i> compared to amoxicillin $4.687$ mg/mL, MIC of $18.75$ mg/mL compared to amoxicillin $4.687$ mg/mL against <i>E. coli</i> (MIC and MBC of $37.5$ mg/mL compared to amoxicillin $4.687$ mg/mL against <i>S. aureus</i> and <i>S. typhi</i> )	Broth dilution method	Lake Victoria Region of Kenya	Not reported	[21]		
<i>Wound healing</i>	The methanol leaf extract had an MIC (mg/ml) of $9.1$ and an MBC (mg/ml) of $10.4$ and exhibited a ZOI of $16 \pm 1.27$ mm against <i>E. coli</i> compared to ciprofloxacin $17 \pm 1.38$ mm	Disc diffusion method/broth dilution method	Kenyatta University Arboretum	Not reported	[22]		

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Tithonia diversifolia</i> (Hemsl.) A. Gray (Asteraceae)	Constipation, stomach pains, liver pains, indigestion and sore throats and as an antiviral	Leaves	The ethyl acetate leaf extract exhibited a ZOI of $8.0 \pm 0.5$ mm against <i>Streptococcus pneumoniae</i> , compared to $2.4 \pm 0.6$ mm (gentamycin) $2.2 \pm 0.4$ mm (erythromycin)	Disk diffusion method	Eastern Kenya	A 70% ethanol extract of the aerial parts was toxic to the kidney and liver toxicity at the lowest dose tested (400 mg/kg). <i>T. diversifolia</i> should be used with caution as it may be toxic especially in prolonged use at higher doses	[18, 23]
			The methanol leaf extract (1 g/ml) exhibited ZOI of 21.6 mm, 19.3, and 18.0 against <i>S. aureus</i> , <i>P. aeruginosa</i> , and <i>K. pneumoniae</i> compared to amoxicillin 23.0 mm, 17.3 mm, and 17. 66 mm, respectively; MIC of 37.5 mg/ml against <i>S. aureus</i>	Agar disc diffusion method/broth dilution	Twiga Region in Central Province		[24]
			The ethyl acetate leaf extract exhibited a ZOI of 18.2 mm against <i>S. typhi</i> compared to chloramphenicol with a ZOI of 23.3 mm and ciprofloxacin with a ZOI of 26.0 mm	Disc diffusion method	Nyamira County		[25]
	Gastrointestinal disorders		The dichloromethane leaf extract (25 mg/ml) exhibited a ZOI of 18 mm against <i>S. aureus</i> and 14 mm against <i>P. aeruginosa</i>	Agar well diffusion method	University of Kabianga Botanical Garden, Kericho County		[26]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Skin diseases, diarrhea, dysentery, laxative, malaria</i>			The methanol root extracts exhibited a ZOI of 1.58 cm compared to streptomycin (1.30 cm) against <i>S. aureus</i>	Disk diffusion method	Kibuye, Kisumu County	The methanol and dichloromethane crude root extracts of had an LD <sub>50</sub> of 1927 mg/kg after a period of 14 days. the extracts at high concentration and at a high dose tend to be toxic	[27, 28]
<i>Malaria, skin conditions, livestock infections</i>			The methanol stem bark extracts (100 mg/ml) had a ZOI of 19.0 mm compared to 30 µg/ml gentamycin (19.0 mm) against <i>S. aureus</i> , ZOI (11.0 mm) compared to 30 µg/ml gentamycin (9.0 mm) against MRSA, ZOI (12.0 mm) compared to 30 µg/ml gentamycin (17.0 mm) against <i>K. pneumoniae</i>	Disk diffusion method	Bomet District		[29]
<i>Senna didymobotrys</i> (Fresen.) Irwin & Barneby (Caesalpiniaceae)	Roots, Stem barks, leaves		The methanol leaf extracts (1 g/ml) had ZOI (16.0 mm) compared to (60.0 mm) gentamycin (10 µg/ml) against <i>B. subtilis</i> . The methanol extracts (1 g/ml) had ZOI (16.0 mm) compared to (24.0 mm) gentamycin (10 µg/ml) against <i>S. aureus</i>	Disk diffusion method	Rarieda		[30]
<i>Diarrhea, fevers, abscesses of the skeletal muscles, and venereal diseases</i>			The methanol 2.5% root bark extract and 7.5% stem bark extracts here inhibited the growth of <i>S. aureus</i> , which was also observed in streptomycin (1 g/L)		The area under disease progress stairs (AUDPS)	Siaya, Nakuru, and Nandi counties	[31]
<i>Oral infections</i>			The ethanol leaf extract (1 mg/mL) exhibited a ZOI of 21.70 ± 0.88 mm, against <i>P. gingivalis</i> and (MIC 0.13 ± 0.00 mg/mL and MBC 0.50 ± 0.00 mg/mL). Amoxicillin had a ZOI of 40.3 mm	Agar well diffusion assay	Borabu Sub-county in Nyamira County		[32]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
			The essential oil ( $10\ \mu\text{L}$ ) exhibited a ZOI (mm) of $21.00 \pm 2.08$ against <i>E.coli</i> , $22.33 \pm 1.67$ against MRSA and $19.00 \pm 1.16$ <i>S. aureus</i> compared to tetracycline $26.00 \pm 0.58$ against <i>E.coli</i> , $9.00 \pm 0.58$ against MRSA and $11.67 \pm 0.88$ <i>S. aureus</i>	Disc diffusion method	Maseno area, Kisumu County	The root extract showed LD <sub>50</sub> >1000 mg/kg and CC <sub>50</sub> >100 $\mu\text{g}/\text{ml}$	[33, 34]
	Food poisoning, malaria, and sore throat		The stem bark methanol extract ( $1\ \text{g}/\text{ml}$ ) had a ZOI of $16.67 \pm 0.67$ mm against <i>S. aureus</i> compared to gentamycin ( $1.0\ \mu\text{g}/\text{disc}$ ) $25.33 \pm 0.67$	Disc diffusion method	Kakamega Forest		[35]
	Malaria and diuretic		The methanol root extract exhibited a ZOI (mm) of 7.0 against <i>E.coli</i> , 6.33 against <i>S. typhi</i> , and 8.66 against <i>S. aureus</i> compared to tetracycline 20.22 against <i>E.coli</i> , 16.00 against <i>S. typhi</i> , and 21.33 <i>S. aureus</i> . MIC and MBC of 9.375 mg/mL against <i>S. typhi</i> and <i>S. aureus</i>	Agar disc diffusion (DD) method broth microdilution technique	Bondo (Alego)		[36]
<i>Toddalia asiatica</i> L. (Rutaceae)	TB and measles	Fruits, stems, barks, roots, leaves	A formulated antiseptic herbal detergent exhibited ZOI of $24.30 \pm 0.67$ mm, $18.00 \pm 0.58$ mm, $16.00 \pm 0.58$ and $19.67 \pm 0.67$ mm against MRSA, <i>P. aeruginosa</i> , <i>E.coli</i> , and <i>S. typhi</i> , respectively, compared to the commercial hand wash $21.67 \pm 0.33$ mm, $19.67 \pm 0.67$ mm, $13.67 \pm 0.33$ mm and $18.33 \pm 0.33$	Disc diffusion method	Slopes of Kajulu Hills, Lake Victoria Basin		[37]
	Malaria and flu		The stem bark DMSO extract exhibited an ZOI of $10 \pm 0.3$ mm compared to flucloxacillin $10 \pm 0.1$ mm against MRSA	Agar well diffusion method	Narok		[38]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Stomachache, abdominal pains, fever, nausea, vomiting, plague, swollen testicles, dysentery, gonorrhoea, tuberculosis</i>	Stomachache, abdominal pains, fever, nausea, vomiting, plague, swollen testicles, dysentery, gonorrhoea, tuberculosis	The methanol whole plant extract had MIC (6.25 mg/ml) compared to (>1 mg/ml for antibiotic standards) against <i>S. aureus</i> and <i>P. aeruginosa</i> and MIC (250 mg/ml) against <i>E. coli</i>	The methanol whole plant extract had MIC (6.25 mg/ml) compared to (>1 mg/ml for antibiotic standards) against <i>S. aureus</i> and <i>P. aeruginosa</i> and MIC (250 mg/ml) against <i>E. coli</i>	Broth dilution method	Meru Central District	The methanol root bark extract had IC <sub>50</sub> ( $\mu$ g/ml) of 198.498 and was considered cytotoxic	[39, 40]
<i>Harrisonia abyssinica</i> Oliv. (Simaroubaceae)	Pneumonia, malaria, and eye ointment	Whole plant, leaves, barks, berries	respectively, compared to that of 0, 0, 0.25 , 0.25 mg/mL of streptomycin against <i>S. aureus</i> <i>B. cereus</i> , <i>P. aeruginosa</i> , and <i>E. coli</i> , respectively, and benzylpenicillin 0.6, 0.6 against <i>S. aureus</i> and <i>B. cereus</i>	Broth dilution method	Machakos and Kitui		[39]
<i>Fever, tuberculosis, and snake bite</i>	Fever, tuberculosis, and snake bite	The methanol-dichloromethane extract (100 mg/ml) had an ZOI of 20.1.6 mm compared to (18.1.2 mm) gentamycin against <i>S. aureus</i> methanol-dichloromethane extract (100 mg/ml) had an ZOI 30.1.7 mm compared to (15.1.3 mm) gentamycin against <i>E. coli</i>	Agar diffusion assay	Bondo District in Nyanza Province			[41]
<i>Infertility, menstrual problems, and stomach pain menstrual</i>	Infertility, menstrual problems, and stomach pain menstrual	The crude extracts showed a moderate activity against <i>S. aureus</i> (11 mm), <i>B. subtilis</i> (7.8 mm), <i>P. aeruginosa</i> (7.0 mm), <i>E. coli</i> (8.5 mm)	Disc diffusion method	Chuka, Meru-South District, Tharaka Nithi County			[42]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Fuerstia africana</i> T. C. E. Fr. (Lamiaceae)	Eye ailments, toothache	Leaves, aerial parts	The methanol leaf extracts (1g/ml) exhibited a ZOI of 17.0 mm compared to (26.0 mm) gentamycin (10 µg/ml) against <i>B. subtilis</i> . The methanol extracts (1g/ml) had a ZOI of 19.0 mm compared to (24.0 mm) gentamycin (10 µg/ml) against <i>S. aureus</i> , methanol extracts (1 g/ml) had a ZOI of 20.0 mm compared to (26.0 mm) gentamycin (10 µg/ml) against MRSA	Disk diffusion method	Kisii south	Extracts were found to be safe at 5000 mg/kg body weight per day. median lethal dose (LD <sub>50</sub> ) of methanol and DCM extracts is >5000 mg/kg	[30, 43]
Boils			The hexane leaf extract (100 mg/ml) exhibited a ZOI of 10.67 ± 0.33 mm compared to (17.33 ± 0.33) chloramphenicol (30 µg/ml) against <i>S. aureus</i> . ZOI (10.50 ± 0.29 mm) compared to (15.00 ± 0.00) chloramphenicol (30 µg/ml) against MRSA, and ZOI (9.67 ± 0.33 mm) compared to (16.50 ± 0.29) chloramphenicol (30 µg) against <i>P. aeruginosa</i>	Agar well diffusion method	Olgenguruone, Nakuru County, and Cheptenye, Kericho County		[43]
Oral infections			The methanol extract exhibited ZOI of (17.21 ± 0.22) compared to gentamycin (23.88 ± 0.01) against <i>K. pneumoniae</i> , ZOI of (14.24 ± 0.35) compared to <i>E. coli</i> and ZOI of (15.18 ± 0.42) compared to gentamycin (25.9 ± 0.01) against <i>S. aureus</i>	Agar well diffusion	Magadi, Kajiado District of Kenya		[44]
			The chloroform extract exhibited ZOI (15.88 ± 0.54) compared to chloramphenicol (21.7 ± 0.11) against <i>S. aureus</i>	Agar well diffusion	Vihiga County, Western Kenya		[45]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
	Sore throat and urinary tract infections		The ethanol stem bark extract (1 g/ml) exhibited ZOI (18.5 mm) compared to gentamycin (10 µg/ml) 19.5 mm and a MIC of (62.5 mg/ml) against <i>S. aureus</i> . The methanol extract had a ZOI of 8.3 mm) compared to gentamycin (19 mm) against <i>E. coli</i> and ZOI of 9.8 mm compared to gentamycin (21.0 mm) against <i>P. aeruginosa</i>	Agar well diffusion/ broth dilution	Bomet District	The methanol leaf extract had an LD <sub>50</sub> value of 3475 mg/kg was; thus, it is nontoxic	[46, 47]
<i>Olea africana</i> (Oleaceae)	Sap used for bone setting (fracture)	Stem bark, barks, twigs, leaves	The aqueous bark extract (1 g/ml) exhibited a ZOI of 10.2 ± 0.6 mm compared to (18.0 ± 0.1) streptomycin (25 µg/ml) against <i>S. aureus</i>	Disk diffusion method	Mbeere, and Embu-Eastern Province		[48]
	Chewing stick		The methanol extract exhibited a ZOI of 12.4 mm against <i>S. aureus</i> , MIC of 1.5 mg/ml against <i>E. coli</i> and 0.30 mg/ml against <i>S. aureus</i>	Broth dilution method	University of Kabianga Botanical Garden, Kericho County		[49]
	Chewing stick		The methanol leaf extract (25 mg/ml) exhibited ZOI 18 m against <i>Pseudomonas aeruginosa</i> , ZOI 19.20 mm against <i>S. aureus</i> and 17 mm against <i>E. coli</i>	Broth dilution method	University of Kabianga Botanical Garden, Kericho County		[26]
	Malaise, antiviral, and appetizer		The ethanol root extract exhibited a ZOI of 8.0 ± 0.9 mm against <i>S. pneumoniae</i> compared to 7.2 ± 0.1 mm (gentamycin) and 7.8 ± 0.3 mm (erythromycin)	Disk diffusion method	Eastern Kenya	The oral LD <sub>50</sub> of the extract was estimated to be >5000 mg/kg, generally safe at doses lower than 1000 mg/kg is in rats ()	[18, 50]
<i>Carissa edulis</i> Vahl. (Apocynaceae)	Kidney problems, pneumonia	Roots, stem, leaves	MICs and MBCs of 37.50 mg/ml against <i>S. typhi</i>	Broth dilution	Transmara West		[51]
	Gonorrhea, asthma		The methanol extract exhibited a ZOI of 9.00 mm against <i>S. typhi</i> compared to amoxicillin 16.0 mm and both MIC and MBC of 37.5 mg/mL against <i>S. typhi</i> and <i>S. aureus</i>	Agar disc diffusion method/broth microdilution technique	Lake Victoria Region		[36]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Rhus natalensis Bernh.</i> (Anacardiaceae)	Malaria	Roots, stems, barks and leaves	The methanol root extract had a MIC of 6.25 mg/against both <i>S. aureus</i> and <i>P. aeruginosa</i> and had a moderate activity with inhibition zone diameters of 11.6 mm against <i>S. aureus</i> and <i>P. aeruginosa</i> compared to gentamycin 25.3 mm and 18 mm	Broth dilution method	Kilifi district	The extracts were safe to the mammalian cells	[52]
38. <i>Prunus africana</i> (Hoolh f.) Kalkman (Rosaceae)	Diarrhea	Barks, stems	The isolated compound (1-epicatechin exhibited a ZOI of 15 ± 0.3 mm against <i>S. aureus</i> and (10 ± 0.2 mm) against <i>P. aeruginosa</i> compared to streptomycin 10 µg/disc 22 ± 0.2 mm and 20 ± 0.3 mm, respectively	Disc diffusion method	Kapkonga Iten, Eldoret town		[53]
	Microbial infections		The isolated compound 1 had a ZOI of exhibited ZOI of 21 mm against <i>S. aureus</i> compared to Chloramphenicol 20 mm	Agar diffusion method	Thika River in Gatanga division, Central Kenya		[54]
	Arrow poisoning and gonorrhea		The methanol bark extract showed a moderate activity against <i>S. aureus</i> (11.0 mm), <i>B. subtilis</i> (10.7 mm), <i>P. aeruginosa</i> (9.7 mm), and <i>E. coli</i> (8.0 mm)	Disc diffusion method	Chuka, Meru-South District, Tharaka Nithi County	The bark had an LD <sub>50</sub> of 2201 mg/kg. The stem bark extract was determined to be nontoxic at the therapeutic dose of 500 mg/kg body weight	[42, 55]
			The methanol stem bark extract exhibited a ZOI of 20 mm against <i>S. aureus</i> and MIC of 0.073 mg/ml. ZOI of 17 mm with MIC of 0.156 mg/ml against MRSA, ZOI of 15 mm and the MIC of 0.3125 mg/ml against <i>P. aeruginosa</i> . ZOI of 12 mm and the MIC of 2.50 mg/ml against <i>S. pneumoniae</i>	Disc diffusion assay	Rift Valley Province of Kenya		[56, 57]
	Chest pain and stomach problems		The hydro-methanolic bark extract exhibited a ZOI of 17.33 ± 0.882 mm against <i>S. typhi</i> and ZOI of 12.33 ± 0.333 mm against <i>Escherichia coli</i> compared to penicillin 27.67 ± 1.2 mm and 20.33 ± 0.333 mm	Agar well diffusion method	University of Eastern Africa, Baraton		[58]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Warburgia ugandensis</i> Sprague (Canellaceae)	Diarrhea, constipation and cough,	The methanol extract (100 mg/ml) exhibited a ZOI of 15.0 mm compared to (18.0 mm) chloramphenicol against <i>S. aureus</i> , ZOI (14.0 mm) compared to (24.0 mm) chloramphenicol against MRSA. The dichloromethane extract had a MIC of 3.125 mg/ml against <i>S. aureus</i> and MRSA	Disc diffusion test/ broth dilution	Ngong Forest	>5000 mg/kg body weight. Extract displayed no apparent deleterious toxicity	[55, 59]	
	STIs, diarrhea, and bronchitis	The methanol extract exhibited ZOI of $3.169 \pm 0.27$ mg/ml against <i>S. aureus</i>	Disk diffusion method	Rift Valley		[60]	
	Hepatitis, gonorrhea tuberculosis, bronchitis, and pneumonia	The methanol extract exhibited a ZOI of $19.33 \pm 0.333$ mm against <i>S. epidermidis</i> compared to penicillin $26.67 \pm 0.333$ , ZOI $17.00 \pm 0.882$ mm against <i>B. cereus</i> and ZOI $11.67 \pm 0.333$ mm against <i>E. coli</i> compared to penicillin $31.33 \pm 0.333$	Disc diffusion	Natural Forest around the University of Eastern Africa, Baraton		[61]	

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
	Dysentery, spice		Garlic juice exhibited a ZOI of 10.0 mm against <i>P. aeruginosa</i> , 11.7 mm against <i>E. coli</i> , 14.7 mm against <i>S. aureus</i> and 17.7 mm for <i>S. typhi</i> . The activity of ampicillin on <i>E. coli</i> and <i>S. typhi</i> was 11.7 mm and 18.7 mm ()	Disc diffusion test	Githurai Market, Nairobi	The LD <sub>50</sub> was found to be 3034 mg/kg, and maximum tolerated dose was 2200 mg/kg [63, 64]	[63]
<i>Allium sativum</i> L. (Liliaceae)	Infection, colds	Rhizome, bulbs	Garlic extract (GE) 200 µl/ml/ exhibited a ZOI of 14 mm compared to gentamycin 24 mm against <i>S. aureus</i> . The methanolic extract of garlic was effective against <i>E. coli</i> , <i>Staphylococcus aureus</i> , and <i>Pseudomonas aeruginosa</i> , with ZOI of 21 mm, 27 mm, and 28 mm, compared to tetracycline 19 mm, 22 mm, and 27 mm, respectively. The garlic methanolic (GM) extract had 0.14 µg/ml against <i>S. aureus</i> and <i>P. aeruginosa</i>	Disc diffusion method	Nakuru Municipal Council Market in Nakuru Town	Nakuru	[65]
	Reduce blood lipids and blood pressure		Green tea (0.1 mg/ml) exhibited a ZOI of 21.3 ± 0.33 mm against <i>E. coli</i> compared to gentamicin 22.3 ± 0.50 mm and ZOI of 23.7 ± 0.33 mm against <i>S. aureus</i> compared to gentamicin 23.2 ± 0.28 mm	Agar well diffusion method	Kenyatta University	There were no observed adverse effects at 2500 mg/kg body weight/day	[67, 68]
<i>Camellia sinensis</i> L. (Theaceae)	Beverage	Leaves	The aqueous crude green tea extracts 400 mg/ml had a ZOI of 20 ± 0.0 mm against <i>S. aureus</i> and MIC 100 mg/ml compared to streptomycin 20 ± 0.0 mm, ZOI of 18 ± 0.0 mm and MIC of 200 mg/ml against <i>E. coli</i> compared to streptomycin 10 ± 0.0 mm	Agar well diffusion method	Tea Research Foundation, Kangaita Substation in Kirinyaga	Ngere in Murang'a County	[69]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Azadirachta indica</i> A. Juss. (Meliaceae)	Udder infections	Leaves, barks, seeds	The neem extract (NE) 200 $\mu$ l/ml exhibited ZOI 11 mm against <i>S. aureus</i> compared to gentamycin 24 mm	Disc diffusion method	Kisauni in Mombasa County	Not reported	[65]
<i>Tagetes minuta</i> L. (Asteraceae)	Intestinal disorders and stomach problems	Leaves	Methanol bark extract exhibited ZOI of 25 mm, 24 mm, and 20 mm against <i>E. coli</i> , <i>P. aeruginosa</i> , and <i>S. aureus</i> , respectively, with MIC (mg/ml) of 15, 17, and 16 The methanolic leaf extract exhibited a ZOI of 17 $\pm$ 1.94 mm against <i>S. aureus</i> compared to vancomycin 25.0 mm and ciprofloxacin 22.0 mm (MIC 8.9 mg/ml; MBC 10.0 mg/ml)	Disc diffusion method	Chumani in Kilifi North Constituency		[70]
<i>Adansonia digitata</i> L. (Bombacaceae)	Oils may cause irritation to the skin		The methanol extract had a MIC (mg/ml) 8.7 and MBC (mg/ml) 10 and ZOI of 16 $\pm$ 1.27 mm against <i>E. coli</i> compared to ciprofloxacin 20 $\pm$ 3.11 mm	Disc diffusion method/broth dilution method	Kenyatta University Arboretum		[22]
<i>Ophalmia</i>			The synthesized AgNPs exhibited a ZOI of 17.1 $\pm$ 0.130 mm against <i>E. coli</i> and 12.9 $\pm$ 0.082 mm against <i>S. aureus</i> compared to ciprofloxacin with a ZOI of 33.4 $\pm$ 0.443 and 12.9 $\pm$ 0.082 mm against <i>E. coli</i> and <i>S. aureus</i>	Disc diffusion technique.	Makueni County	The stem extracts are non-toxic to brine shrimp larvae	[72, 73]
<i>Euclea divinorum</i> <i>Hern</i> (Ebenaceae)	Diarrhea, dysentery	Leaves, barks	The organic extract at 200 mg/ml and 100 mg/ml showed the highest inhibition zones of 14.33 mm and 12 mm, respectively, against MRSA compared to gentamycin 15.5 mm	Disc diffusion technique	Msambweni District		[74]
	Toothbrush, constipation and ulcers	Stems, barks, leaves	The DCM stem bark extract exhibited ZOI (mm) of 10.8 $\pm$ 0.26 against <i>P. aeruginosa</i> and 17.0 $\pm$ 0.42 against <i>S. aureus</i> compared to Augmentin 10.0 $\pm$ 0.02 against <i>P. aeruginosa</i> and 27.0 $\pm$ 0.02 against <i>S. aureus</i>	Disc diffusion method	Bunyala (Budalang <sup>“1</sup> ) district of Busia County	The root extracts have toxic effects and should be used with care; gargling of extracts is recommended instead of swallowing	[75, 76]
	Dental caries		The ethanolic root bark extract had MIC of 25, 50, 25 and 25 $\mu$ g/ml for <i>S. pyogenes</i> , <i>S. aureus</i> , <i>E. coli</i>	Broth dilution	Elgeyo Marakwet, Rift Valley		[77]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Salvadora persica</i> <i>L. var. persica</i> (Salivadoraceae)	Chest problems, stomachache, teeth problems	Roots, stems, barks	The organic root extract had 10 GU's (numerical growth units) at 0.5 mg/ml against <i>M. tuberculosis</i> compared to Isoniazid that had zero GU's at 0.5 mg/ml The methanol bark extract exhibited a ZOI of 21.66 mm against <i>S. aureus</i> compared to Amoxicillin 21.3 mm. ZOI of 20 mm against <i>P. aeruginosa</i> compared to amoxicillin 14.33 mm. ZOI of 15 mm against <i>E. coli</i> compared to amoxicillin 23.6 mm The DCM: MeOH crude leaf extract 200 mg/ml and 100 mg/ml exhibited ZOI (mm) of 1.3 and 10.3, respectively, against MRSA compared to 14 mm for amoxicillin [50 mg/ml]. The lowest MIC values were observed in DCM fraction (40 mg/ml) against MRSA	BACTEC mngIT™ 960 system Agar disk diffusion technique	Various conservancies in Samburu Nkaroni, Wamba Division, Samburu District	High concentrations >5 g/kg of the mammal's body weight can result in toxicity	[36, 78]
<i>Plectranthus barbatus Andrews</i> (Lamiaceae)	Oral thrush and diarrhea	Leaves, roots	The root extracts exhibited a ZOI of 18.67 mm, 20.00 mm, and 25.33 mm in <i>S. aureus</i> , MRSA, and <i>B. cereus</i> compared to streptomycin 39.67 ± 1.76 mm, 39.67 ± 1.76 mm, and 33.00 ± 1.15 mm ()	Agar well diffusion method	Msambweni Subcounty, Kwale County		[81]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Cordia purpurea</i> (Picc.) Aiton (Fabaceae)	Diarrhea	Roots, barks	The methanolic root extract exhibited a ZOI of 15 mm compared to (21.33 mm) amoxicillin against <i>S. aureus</i> and ZOI (23.66 mm) compared to (17.58 mm) amoxicillin against <i>P. aeruginosa</i> . The methanolic extract had a MIC of 18.75 mg/ml compared to 18.75 mg/ml cefpodoxime against <i>S. aureus</i> and 18.75 mg/ml compared to 9.372 mg/ml cefpodoxime against <i>P. aeruginosa</i>	Agar disc diffusion method/broth dilution	Samburu-Wamba conservancies	Not reported	[79]
			The methanolic extract exhibited a ZOI of 14.33 mm compared to (21.33 mm) amoxicillin against <i>S. aureus</i> and ZOI (19.66 mm) compared to (17.58 mm) amoxicillin against <i>P. aeruginosa</i> . The methanolic extract had MIC of 37.50 mg/ml compared to 18.75 mg/ml cefpodoxime against <i>S. aureus</i> and 37.50 mg/ml compared to 9.372 mg/ml cefpodoxime against <i>P. aeruginosa</i> . Organic bark extract exhibited zero GU's at 0.5 mg/ml against <i>M. tuberculosis</i> and <i>M. kansasii</i> compared to Isoniazid that had zero GU's at 0.5 mg/ml	Agar disc diffusion method/broth dilution	Samburu-Wamba conservancies	Various conservancies in Samburu	[36]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Croton macrostachyus</i> Hochst. Ex Delile: (Euphorbiaceae)	Diarrhea, stomach ache	Barks, roots	The ethyl acetate bark extract exhibited ZOI between $10.1 \pm 0.6$ mm and $16.0 \pm 1.2$ mm against <i>S. typhi</i> , <i>E. coli</i> and <i>K. pneumoniae</i> .  The methanolic extract exhibited a ZOI of 23.66 mm compared to (21.33 mm) amoxicillin against <i>S. aureus</i> and ZOI (18.0 mm) compared to (17.58 mm) amoxicillin against <i>P. aeruginosa</i> . The methanolic extract had an MIC of 37.50 mg/ml compared to 18.75 mg/ml cefpodoxime against <i>S. aureus</i> and 18.75 mg/ml cefpodoxime against <i>P. aeruginosa</i> . The essential oil from leaves exhibited ZOI ( $26.6 \pm 5.7$ mm) compared to $9.372$ mg/ml chloramphenicol against <i>E. coli</i>	Agar disc diffusion method	Baraton Community in Nandi District of Kenya	The aqueous stem extract does not provoke death until the dose of 16 g/kg. There is a wide margin of safety for the therapeutic use of the extract	[82, 83]
<i>Ocimum gratissimum L.</i> (Lamiaceae)	Ear infections, tooth gagle Sore eyes and rectal prolapse	Leaves	The essential oil from leaves extract had ZOI ( $21.7 \pm 2.1$ mm) compared to ( $28.0 \pm 0.7$ mm) chloramphenicol against <i>E. coli</i> . ZOI ( $26.6 \pm 5.7$ mm) compared to ( $23.5 \pm 2.1$ mm) chloramphenicol against <i>S. aureus</i>	Agar disc diffusion method	Meru Meru District of Eastern Kenya	The oil can cause an inflammatory response	[84, 85]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Ocimum suave</i> Wild (Lamiaceae)	Ear infections, cough and disinfectant	Leaves, roots	The methanolic leaf extract had a MIC of (6.25 mg/ml) compared to >1 mg/ml for antibiotic standards against <i>S. aureus</i> and methanolic extract had an MIC of 31.25 mg/ml compared to >1 mg/ml for antibiotic standards against <i>P. aeruginosa</i> and <i>E. coli</i> . The methanol root extract exhibited a mean ZOI of 14 mm against <i>S. aureus</i> compared to amoxicillin 21.3 mm, ZOI of 21 mm against <i>P. aeruginosa</i> compared to amoxicillin 17.5 mm and ZOI of 18 mm against <i>E. coli</i> compared to amoxicillin 23.6 mm.	Broth dilution method	Meru Central district	The aqueous leaf extract is non-toxic in acute and subchronic intake. No gross abnormalities, teratogenic, or histological changes observed	[40, 87]
<i>Premna retinosa</i> (Hochst.) Schauer (Compositae)	Respiratory-related illnesses	Roots	33.7 ± 0.3 mm (oxacillin 10 µg/disc and gentamycin 10 µg), methanolic extract had a ZOI of 8.7 mm compared to (22 mm) oxacillin 10 µg/disc and Gentamycin 10 µg against <i>S. aureus</i> and ZOI 11.7 mm) compared to (24 mm) oxacillin 10 µg/disc and gentamycin 10 µg against <i>E. coli</i>	Disc diffusion and microdilution techniques	Mbeere community, Kenya	The dichloromethane and ethyl acetate fractions were within the acceptable toxicity limit ( $CC_{50} < 90$ )	[88]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Hagenia abyssinica</i> (Bruce) JF Gmel (Rosaceae)	Diarrhea, stomachache, tongue infections, sores	Stem bark, leaves	The dichloromethane/methanol stem bark extract exhibited ZOI of 19.0 mm against <i>S. aureus</i> compared to Erythromycin (0.01 mg/ml) 22 mm, ZOI of 20.0 mm against <i>E. coli</i> compared to Erythromycin 0.01 mg/ml 20.0 mm, ZOI of 18 mm against <i>B. subtilis</i> compared to Erythromycin 0.01 mg/ml 20 mm The hexane leaf extract (100 mg/ml) compared to (17.33 ± 0.33) chloramphenicol (30 µg/ml) against <i>S. aureus</i> , ZOI (19.33 ± 1.33 mm) compared to (15.00 ± 0.00) chloramphenicol (30 µg/ml) against MRSA and ZOI (13.00 ± 1.00 mm) compared to (16.50 ± 0.29) chloramphenicol (30 µg) against <i>P. aeruginosa</i>	Agar well diffusion method	Aberdare ranges, Kiburu Forest Station	5000 mg/kg body weight per day. Median lethal dose (LD <sub>50</sub> ) of methanol and DCM extracts is >5000 mg/kg	[43, 89]
<i>Clerodendrum myricoides</i> (Hochst.) R. Br. ex Vatke; (Lamiaceae)	Respiratory diseases, tonsillitis, eye infections, gonorrhea	Whole plant	The aqueous extract (1 g/ml) exhibited ZOI (13.8 ± 0.2 mm) compared to (18.0 ± 0.1) streptomycin (25 µg/ml) against <i>S. aureus</i>	Disk diffusion method	Mbeere Community, Kenya	The methanol extracts within the acceptable toxicity limit with a CC <sub>50</sub> of >500 µg/ml the LD <sub>50</sub> value of 3475 mg/kg and thus is non-toxic	[90]
<i>Securidaca longipedunculata</i> Var. <i>parvifolia</i> (Polygalaceae)	Infusion reduces swellings	Roots, barks	The aqueous extract (1 g/ml) had a ZOI of 12.5 ± 2.2 mm compared to (18.0 ± 0.1) streptomycin (25 µg/ml) against <i>S. aureus</i>	Disk diffusion method	Mbeere, and Embu-Eastern Province	The extract has an LD <sub>50</sub> value of 771 mg/kg body weight and is nontoxic at relatively high concentrations	[48, 91]
	Sexually transmitted infections		The bark and root extract exhibited ZOI of 20.1 mm and 13.5 mm, respectively, against <i>N. gonorrhoeae</i> , compared to 12 mm and 19.1 mm ciprofloxacin and tetracycline	Disc diffusion method	Bungoma County		[92]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Tamarindus indica</i> L. (Fabaceae)	Meat preservative	Fruit paste, bark	The water extract exhibited a ZOI of 34.67 mm, and 24 mm against <i>E. coli</i> and <i>S. aureus</i> , respectively, compared to chloramphenicol	Disc diffusion test	Chepararia and Kongelai subcounties of West Pokot County	The pulp extract of <i>Tamarindus indica</i> at 3000 mg/kg and 5000 mg/kg body weight of resulted in no mortality and is practically nontoxic	[93, 94]
	Diarrhea, typhoid	Bark	The methanol bark extract (1 g/ml) had a ZOI of 14.5 mm compared to (24.0 mm) gentamycin (10 µg/ml) against <i>S. aureus</i> . The methanol extracts (1 g/ml) had a ZOI of 16.0 mm compared to (26.0 mm) gentamycin (10 µg/ml) against <i>B. subtilis</i> . The organic crude extract exhibited the good inhibition against <i>B. cereus</i> at 200 and 100 mg/ml concentrations with a ZOI of 13.87 mm and 12.167 mm, respectively, compared to gentamycin 15 mm	Disk diffusion method	Rarieida	The acute oral median dose ( $LD_{50}$ ) of the root bark extract was >6750 mg/kg body weight. Plant is of relatively low toxicity	[30]
<i>Zanthoxylum chalybeum Engl.</i> (Rutaceae)	Malaria, pneumonia, sore throat	Leaves, roots, barks	The organic extract exhibited mean inhibition zone values of $24.33 \pm 0.33$ mm against MRSA compared to streptomycin $39.67 \pm 1.76$ mm	Disc diffusion technique	Msambweni District	Msambweni Kwale County	[74]
<i>Lantana camara</i> L. (Verbenaceae)	Skin rashes, boils	Leaves	The methanol leaf extract (1 g/ml) had a ZOI of 17.0 mm compared to gentamycin (10 µg/ml) against <i>S. aureus</i>	Agar well diffusion method	Bomet District	For short-term use, the extract exhibited very low toxicity, while long-term exposure results in liver and kidneys. the root extract was the most toxic part	[46, 96]
		leaves	The organic leaf extracts MICs and MBCs of 37.5 mg/mL against both <i>S. aureus</i> and <i>P. aeruginosa</i>	Broth dilution technique	Around Lake Victoria Region		[36]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Mangifera indica L.</i> (Anacardiaceae)	Burns, scalds, sores, abscesses, food	Leaves, fruits	The methanol extracts (1 g/ml) had ZOI (18.5 mm) compared to gentamycin 10 $\mu\text{g}/\text{ml}$ (19 mm) against <i>S. aureus</i> , ZOI (13.0 mm) compared to gentamycin (20 mm) against <i>E. coli</i> , ZOI (17 mm) compared to 10 $\mu\text{g}/\text{ml}$ gentamycin (18.5 mm) against <i>P. aeruginosa</i> . The methanol extract exhibited a ZOI of $2.07 \pm 0.15$ cm against <i>S. aureus</i> compared to norfloxacin at 10 $\mu\text{g}$ 2.95 cm and ZOI $1.93 \pm 0.09$ compared to Norfloxacin at 10 $\mu\text{g}$ 2.95 cm against <i>E. coli</i> . The ethanol extract had ZOI (mm) of $9.2 \pm 0.3$ compared to $6.8 \pm 0.4$ (gentamycin) and $6.6 \pm 0.2$ (erythromycin) against <i>S. pneumoniae</i> .	Agar well diffusion Disc diffusion method	Bomet District Makueni and Embu	The oral or dermal administration of the extract showed no lethality at the limit doses of 2,000 mg/kg body weight, and no adverse effects were found	[46, 97]
<i>Terminalia brownii</i> Fresen (Combretaceae)	Diarrhea, ulcers, and sexually transmitted diseases	Bark, leaves, roots		Disk diffusion method	Eastern Kenya	The aqueous leaf extract (1 g/ml) had a ZOI of 18.0 $\pm$ 0.8 mm compared to (18.0 $\pm$ 0.1) streptomycin (25 $\mu\text{g}/\text{ml}$ ) against <i>S. aureus</i> , ZOI (11.7 $\pm$ 0.5 mm) compared to (16.0 $\pm$ 0.2) streptomycin (25 $\mu\text{g}/\text{ml}$ ) against <i>E. coli</i> , ZOI (12.8 $\pm$ 1.0 mm) compared to (15.0 $\pm$ 0.3 mm) streptomycin 25 $\mu\text{g}/\text{ml}$ against <i>B. subtilis</i> .	[48]
						The roots and stem bark extracts exhibited mild cytotoxic activity with LC <sub>50</sub> values ranging from 113.75 to 4356.76 and 36.12 to 1458.81 $\mu\text{g}/\text{ml}$	Mbeere, and Embu-Eastern Province

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Markhamia lutea</i> (Benth.) K. Schum. (Bignoniaceae)	Eye infection	Bark	The chloroform extracts had a ZOI of 22.82 mm against <i>E. coli</i> , 18.79 mm against <i>S. aureus</i> and 17.94 mm against <i>P. aeruginosa</i> compared to gentamycin 22.27 mm, 22.52 mm, and 20.17 mm, respectively	Agar well diffusion	Emuhaya Sub-county, Western Kenya	Not reported	[100]
<i>Asparagus setaceous</i> <i>Kunth Jessop</i> (Asparagaceae)	Syphilis, gonorrhea	Aerial parts, roots	The MIC values for ethanolic aerial part extract ranged from 3.2 mg/ml for <i>S. aureus</i> , 6.25 mg/ml for <i>E. coli</i> , and 25 mg/ml for <i>B. subtilis</i> , <i>P. aeruginosa</i> , and <i>S. faecalis</i> , while same plant, MIC ranged from 6.25 mg/ml for <i>S. aureus</i> and <i>B. subtilis</i> to 25 mg/ml for <i>E. coli</i> , <i>P. aeruginosa</i> , and <i>S. faecalis</i>	Broth dilution	Gatundu	Not reported	[101]
<i>Caesalpinia volkensii Harm</i> (Caesalpiniaceae)	Bronchitis , pneumonia	Leaves	The MIC values for the ethanolic leaf extract was 6.25 mg/ml for <i>S. aureus</i> , 12.5 mg/ml for <i>B. subtilis</i> and 25 mg/ml for <i>E. coli</i> and <i>P. aeruginosa</i>	Broth dilution	Gatundu	The organic extract had a median lethal dose of >2000 mg/kg body weight, hence is safe	[101], [102]
<i>Thylachium africanum Lour.</i> (Capparaceae)	Diarrhea	Bark	The methanol extract exhibited a ZOI of 18.66 mm against <i>S. aureus</i> compared to amoxicillin 21.3 mm, ZOI of 23.33 mm against <i>P. aeruginosa</i> compared to amoxicillin 17.5 mm, ZOI of 15 mm against <i>E. coli</i> compared to amoxicillin 23.6 mm	Agar disk diffusion technique	Namunyak, Wamba Division, Samburu District	Not reported	[79]
<i>Alectra sessiliflora</i> (Vahl) Kuntze (Scrophulariaceae)	Diarrhea, sexually transmitted infections, wounds	Whole plant	The methanol extract 50 mg/ml exhibited ZOI of 15.46 mm against <i>S. aureus</i> compared to chloramphenicol 19.23 mm, ZOI of 10.72 mm against <i>P. aeruginosa</i> compared to chloramphenicol 19.22 mm, ZOI of 9.76 mm against <i>E. coli</i> compared to chloramphenicol 19.10 mm	Disk diffusion method	Vihiga county	Not reported	[103]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Teclea nobilis</i> (Rutaceae)	Colds and chest problems	Leaves	The DCM extract exhibited a ZOI of 10 mm against <i>S. aureus</i> compared to gentamycin 13 mm and ampicillin 14 mm	Disk diffusion method	Siroch, Keiyo Sub-county, Elgeyo-Marakwet County	Not reported	[104]
<i>Ochna thomasiana</i> (Ochnaceae)	Microbial infection	Root, stem barks	The methanolic extract was found effective against <i>S. aureus</i> and <i>B. subtilis</i> , which gave ZOI of 15 mm and 20 mm, respectively, compared to tetracycline 20 mm and 18 mm	Disc diffusion method	Arabuko-Sokoke, forest in Malindi district, Kilifi County	Not reported	[105]
<i>Cinnamomum cassia</i> Presl. (Lauraceae)	Food poisoning, flavoring	Fruits	The ethanolic extract of cinnamon was effective against <i>E. coli</i> by with a ZOI of 27 mm and MIC of 0.12 µg/ml	Agar well diffusion method/broth dilution	Kenya University	Not reported	[66]
<i>Bidens pilosa</i> L. (Asteraceae)	Stomach upsets	Leaves	The stem bark DMSO extract exhibited ZOI of 12 ± 0.1 mm compared to Flucloxacillin 14 ± 0.7 mm against <i>E. coli</i> . The methanol extract (200 mg/ml) had a ZOI of 15.00 ± 0.00 mm against <i>B. cereus</i> compared to Gentamicin (40 µg/ml)	Agar well diffusion method	Narok	Extract showed no adverse effects in mice and chickens at a dose of 5% or less of food	[38, 106]
<i>Acacia lahai Stead. &amp; Hochsleit Benth.</i> (Fabaceae)	Skin eruptions	Barks	11.33 ± 0.29 mm against MRSA compared to Gentamicin (40 µg/ml) 15.67 ± 1.04 mm. The acetone extract (200 mg/ml) had a ZOI of 10.33 ± 0.58 mm against <i>P. aeruginosa</i> compared to Gentamicin (40 µg/ml)	Disk diffusion method	Mosonik hill, Sotik Sub-county, Bonet County	Not reported	[107]
<i>Bridelia micrantha</i> (Hochst.) Baill. (Euphorbiaceae)	Stomachache, diarrhea in children	Leaves	15.12 ± 0.63 mm The methanol extract 100 mg/mL exhibited ZOI of 19 mm and 13 mm against <i>S. aureus</i> and <i>S. typhi</i>	Disc diffusion method	Kilifi District	The extract has a wide margin of safety for oral use at doses below 2000 mg/kg	[49, 108]
<i>Grewia plagiophylla</i> K. Schum. (Malvaceae)	Dysentery, typhoid	Leaves	The methanol extract 100 mg/mL exhibited ZOI of 20 mm and 17 mm against <i>S. aureus</i> and <i>S. typhi</i>	Disc diffusion method	Kilifi District	Not reported	[49]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Vigna subterranea</i> (L.) (Fabaceae)	Traditional food	Nuts	The MIC values for organic extract ranged from <i>E. coli</i> —7.72 ± 0.35 µg/ml, <i>S. aureus</i> —12.5 ± 0.32 µg/ml, and <i>P. aeruginosa</i> —7.95 ± 0.10 µg/ml. At 100 µg/ml, <i>E. coli</i> , <i>S. aureus</i> , and <i>P. aeruginosa</i> showed a ZOI of 27 ± 0.74 mm, 25.3 ± 0.40 mm, and 25.1 ± 0.24 mm, respectively, compared to those of ceftriaxone, which were 37.0 ± 0.5, 41.3 ± 0.9, and 42.3 ± 0.9 mm.	Disc diffusion method	Bungoma county	Not reported	[109]
<i>Citrus limon</i> (L.) Osbeck (Rutaceae)	Sore throat, chest pain	Rhizomes	Lemon juice inhibited the growth of <i>S. typhi</i> with a ZOI of 11.0 mm and ZOI 11.0 mm against <i>P. aeruginosa</i> compared to chloramphenicol 20.0 ± 0.0 mm. The methanolic extract gave ZOI of 24 mm, and 20 mm against <i>E. coli</i> and <i>S. aureus</i> , respectively, compared to chloramphenicol 16 mm and 18.	Disc diffusion test	Githurai market, Nairobi	The juice is considered non-toxic and extremely safe for consumption even at above 80% concentration	[64, 110]
<i>Ziziphus abyssinica</i> Hochst (Rhamnaceae)	Meat preservative	Fruit paste	The ZOI in <i>S. aureus</i> varied from 16 ± 0.02 mm in replicate 2 to 18 ± 0.01 mm in replicate 1, <i>E. coli</i> (13 ± 0.02 mm in replicate 2 to 15 ± 0.02 mm in replicate 1), and in <i>K. pneumoniae</i> (20 ± 0.01 mm in replicate 3, to 20 ± 0.02 mm in replicates 1 and 2).	Disc diffusion test	Chepararia and Kongelai subcounties of West Pokot county	The acute toxicity ( $LD_{50}$ ) of the leaf extracts was found to be greater than 5000 mg/kg and is considered relatively safe for use	[94, 111]
<i>Mentha spicata</i> L. (Lamiaceae)	Common cold	Leaves	The organic extract showed a highest activity against <i>B. subtilis</i> (28.5 ± 0.3 mm), <i>S. aureus</i> (22.6 ± 1.0 mm), <i>B. cereus</i> (22.0 ± 0.3 mm), <i>E. coli</i> (21.7 ± 0.7 mm), <i>P. aeruginosa</i> (21.5 ± 0.9 mm), <i>S. typhimurium</i> (17.3 ± 0.3 mm), <i>K. pneumoniae</i> (15.3 ± 0.4 mm), and <i>P. mirabilis</i> (12.3 ± 0.5 mm)	Agar well diffusion method	Egerton University	The $LC_{50}$ value was 1701 g/ml in brine shrimp lethality assay, indicating that the plant extract is nontoxic	[112, 113]
<i>Indigofera lupatana Baker F.</i> (Leguminosae)	Cough, diarrhea, and gonorrhea	Roots	Mbeere District, in the Eastern Province of Kenya	Disc diffusion assay	The extract had an $LC_{50}$ value greater than 1000 µg/ml which is an indication that they are all nontoxic	[114]	

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Momordica charantia</i> L. (Cucurbitaceae)	Diabetes	Fruit	The extracts exhibited a ZOI of 10.66 mm against <i>S. aureus</i> compared to amoxicillin 21.03 mm and MIC and MBC of 37.5 mg/mL, ZOI of 9.33 mm MIC and MBC of 37.5 mg/mL against <i>P. aeruginosa</i> . The methanol and chloroform extracts together with the pure compound, friedelin, were active against <i>S. aureus</i> with zones of inhibition of 18.0, 22.0, and 10.0 mm, respectively. Gentamicin (10 µg/ml) had a ZOI of 26.0 mm against <i>S. aureus</i> .	Agar disc diffusion (DD) method/broth dilution technique	Lake Victoria Region	The LD <sub>50</sub> of the ethanolic extract is considered safe to be consumed below 2000 mg/kg	[36, 115]
<i>Blighia unijugata</i> Bak (Sapindaceae)	Tonic, antihelminthic	Roots, pods, and leaves	Disc diffusion assay	Kiangwachi, Kirinyaga District	The extract has the LD <sub>50</sub> of 5.629 ± 0.29 g/kg b. wt	[116, 117]	
<i>Moringa oleifera</i> Lam. (Moringaceae)	Antioxidant, spasms	Seeds, stem and bark	Broth microdilution technique	Moringa oleifera is genotoxic at supra-supplementation levels of 3000 mg/kg b.wt. However, intake is safe at levels ≤ 1000 mg /kg b.wt	[118, 119]		
<i>Maesa lanceolata</i> Forsk (Myrsinaceae)	Bacterial infections	Roots, leaves, and stem bark	Disc diffusion method	Elgeyo Marakwet county	DCM extracts of stem bark and leaves were lowly toxic. No mortality was observed within 24 hours	[120]	
<i>Satureja biflora</i> Buch-Ham (Lamiaceae)	Antimicrobial	Leaves	16.00 ± 1.0 mm	The essential oil exhibited a ZOI of (31 ± 0.5 mm), MIC 125 mg/mL against <i>S. typhi</i> and (24 ± 02 mm), 93.8 mg/mL against <i>S. aureus</i> compared to chloramphenicol 10 ± 1.0 mm, MIC 25 mg/mL against <i>S. typhi</i> and 24 ± 1.0 mm, MIC 31 mg/mL against <i>S. aureus</i> . An isolated compound epicatechin had zone diameter of growth inhibition of crude extract was (15.05 mm) against <i>S. aureus</i> and (14.02 mm) against <i>B. subtilis</i> compared to tetrycline 18.02 mm against <i>S. aureus</i> and <i>B. subtilis</i>	Not reported	[86]	
<i>Lannea schweinfurthii</i> (Engl.) Engl (Anacardiaceae)	Bacterial infections		Disc diffusion method	Bondo, Siaya County	Not reported	[121]	

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Annanus comosus</i> (Bromeliaceae)	Indigestion	Fruits	The MIC of nanoencapsulated bromelain against <i>Enterobacter</i> spp., <i>Citrobacter</i> spp., <i>Serratia</i> spp., and coagulase-negative <i>Staphylococci</i> was 25 µg/ml, while that of <i>E. coli</i> was 50 µg/ml. The MIC of nanoencapsulated bromelain against <i>Klebsiella</i> spp. and <i>S. aureus</i> was 200 µg/ml. Bromelain was effective against gram-positive and gram-negative bacteria. Streptomycin had a MIC of 22.2 µg/ml	Agar well diffusion method/broth microdilution method	Thika Town	Leaf extract is nontoxic	[122, 123]
<i>Helichrysum forskahlii</i> (Asteraceae)	Cough	Whole plant	<i>H. forskahlii</i> had the highest inhibition zone against MRSA of 19.5 and 18.5 mm in agar well and agar disk diffusion respectively. Chloramphenicol had ZOI 24 mm	Disc diffusion method/agar well diffusion method	Losho, Narok County	The brine shrimp lethality test found the plant to be highly toxic with a lethal concentration of 0.009 mg/ml	[124]
<i>Citrullus lanatus</i> (Cucurbitaceae)	Food	Fruit	The MIC value of the nanoparticles was $45.00 \pm 0.01$ mg/ml for <i>S. typhi</i> and $38.50 \pm 0.00$ mg/ml for <i>E. coli</i> , while the MBC value was $60.00 \pm 0.05$ mg/ml for <i>S. typhi</i> and $50.00 \pm 0.00$ mg/ml for <i>E. coli</i>	Disc diffusion method	Wakulima Market, Muthurwa Market, and Githurai Market within Nairobi county	$LD_{50}$ of EECLS was greater than 2000 mg/kg BW and the no observed adverse effect level (NOAEL) of EECLS was at a dose of 1000 mg/kg in rats	[125, 126]
<i>Hyptis spicigera</i> (Lamiaceae)	Stomach ache, pulmonary troubles	Leaves	The methanolic extract (1 g/ml) had a ZOI of 19.3 mm and 19.0 mm against <i>S. aureus</i> and <i>S. typhi</i> , respectively, compared to amoxicillin 23.0 mm and 21.3 mm. The methanolic extract had an MIC of 37.5 mg/ml against <i>S. aureus</i> and <i>S. typhi</i> compared to 18.75 mg/ml for amoxicillin	Agar disc diffusion method	Marera Region in Central Province	Not reported	[79]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Crotalaria quartiflora</i> (Fabaceae)	Diarrhoea	Leaves	The methanol extract (1 g/ml) had a ZOI of 21.0 mm, 19.3, 21.0, 20.7, 18.7, and 20.7 against <i>S. aureus</i> , <i>S. typhi</i> , <i>E. coli</i> , <i>P. aeruginosa</i> , and <i>K. pneumoniae</i> , respectively, compared to amoxicillin 23.0, 21.3, 20.2, 17.3, and 17.66 and an MIC of 37.5 mg/ml against <i>S. aureus</i> , <i>S. typhi</i> , <i>E. coli</i> , <i>P. aeruginosa</i> , and <i>K. pneumoniae</i> compared to 18.75 mg/ml for amoxicillin	Agar disc diffusion method/broth dilution	Tatu region in central province	Not reported	[79]
<i>Eurphobia hirta</i>	Diarrhea, asthma	Whole plant	The methanol extract (1 g/ml) had a ZOI of 21.0 mm, 18.66, 19.66, 16.33, 16.33, and 14.33 against <i>S. aureus</i> , <i>S. typhi</i> , <i>E. coli</i> , <i>P. aeruginosa</i> , and <i>K. pneumoniae</i> , respectively, compared to amoxicillin 23.0, 21.3, 20.2, 17.3, and 17.66 and MIC of 18.75 mg/ml against <i>S. aureus</i> , <i>S. typhi</i> , <i>E. coli</i> , <i>P. aeruginosa</i> , and <i>K. pneumoniae</i> compared to 18.75 mg/ml for amoxicillin	Agar disc diffusion method/broth dilution	Twiga Region in Central Province	The LD <sub>50</sub> of this plant is more than 5000 mg/kg	[79, 127]
<i>Lippia kituiensis</i> (Verbenaceae)	Diarrhea, chest problems	Leaves	The methanol extract (1 g/ml) had a ZOI of 23.3 mm and 17.6 mm, against <i>S. aureus</i> and <i>P. aeruginosa</i> , respectively, compared to amoxicillin 23.0, and 17.3. The methanol extract (1 g/ml) had an MIC of 37.5 mg/ml against <i>S. aureus</i>	Agar disc diffusion method/broth dilution	Marera Region in Central Province	Not reported	[79]
<i>Eurphobia scolopax</i> (Euphorbiaceae)	Stomach ache, common cold, TB	Stem	The extract exhibited zero GUs at 0.5 mg/ml against <i>M. kansasii</i> and <i>M. tuberculosis</i> compared to Isoniazid that had zero GUs at 0.5 mg/ml	BACTEC mgIT™ 960 system	Various conservancies in Samburu	Not reported	[36]
<i>Acacia horrida</i> (Fabaceae)	Diarrhoea, TB	Barks	<i>A. horrida</i> had appreciable inhibition (257 GUs) against <i>M. tuberculosis</i> (198 GUs) at the concentration of 0.5 mg/ml compared to Isoniazid that had zero GUs at 0.5 mg/m	BACTEC mgIT™ 960 system	Various conservancies in Samburu	Not reported	[36]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Phyllanthus urinaria</i> Linn (Phyllanthaceae)	Dysentery, diarrhea, stomach ache	Leaves, roots	MIC and MBC of 18.75 mg/ml and 37.50 mg/ml, respectively, against <i>E. coli</i>	Broth dilution	Transmara West	Not reported	[51]
<i>Rhamnus prinoides</i> L'He'r (Rhamnaceae)	Typhoid, stomach ache	Stem, roots	The extract inhibited <i>E. coli</i> with MIC and MBC of 9.37 mg/ml	Broth dilution	Transmara west	Rhamnus prinoides was nontoxic to brine shrimp	[51, 128]
<i>Tetradenia riparia</i> (Lamiaceae)	Respiratory problems, stomach ache, diarrhea, antiseptic	Roots, stem	The organic extract inhibited <i>S. epidermidis</i> with a ZOI of $27.67 \pm 0.333$ mm compared to penicillin $26.67 \pm 0.333$ and <i>E. coli</i> with a ZOI of $13.33 \pm 0.333$ mm compared to penicillin $31.33 \pm 0.333$	Disc diffusion	Natural forest around the University of Eastern Africa, Baraton	Toxic effect recorded for root and fruit extracts but not for leaf or stem extracts. In mice at dose 1.0 g/kg	[61, 129]
<i>Kigelia africana</i> Lam and Benth (Bignoniaceae)	Laxative, gonorrhea, tuberculosis, diarrhea	Fruits, barks	The methanolic extract had a ZOI of 11.3 mm compared to (19 mm) gentamycin against <i>S. aureus</i> and ZOI (10 mm) compared to (9 mm) chloramphenicol against MRSA. The MIC values of acetone extracts were 6.25 ng/ml against MRSA	Disc diffusion	Kaptumo Division, Nandi	Not reported	[130]
<i>Conyza sumatrensis</i> (Asteraeae)	Pimples	Leaves/ roots	The methanolic extract had a ZOI of 26.85 mm compared to (13.67 mm) chloramphenicol against <i>E. coli</i> and ZOI (27 mm) compared to (15.8 mm) chloramphenicol against <i>B. plumulus</i>	Agar diffusion assay method	Rarieda, Bondo district, of Nyanza province in Kenya	Experiments indicate the methanol extract to be safe even at high and repeated doses in pre-clinical studies	[131, 132]
<i>Pilosigma thonningii</i> (Fabaceae)	Cough, colds, chest pains, stomachache, wounds	Stem bark	The methanolic extract had MIC (3.125 mg/ml) compared to ( $>1$ mg/ml for antibiotic standards) against <i>S. aureus</i> , MIC (31.25 mg/ml) compared to ( $>1$ mg/ml for antibiotic standards) against and <i>E. coli</i> and MIC (15.625 mg/ml) against <i>P. aeruginosa</i>	Broth dilution method	Meru central district	Plant extracts had LD <sub>50</sub> values $>2000$ mg/kgbw and were hence deemed to be nontoxic	[40, 133]
<i>Erythrina abyssinica</i> (Fabaceae)	Anthrax, syphilis, gonorrhea, burns, body swellings	Root bark	The methanolic extract had an MIC of 3.125 mg/ml compared to $>1$ mg/ml for antibiotic standards against <i>S. aureus</i> , MIC (250 mg/ml) compared to ( $>1$ mg/ml for antibiotic standards) against and <i>E. coli</i> and MIC (125 mg/ml) against <i>P. aeruginosa</i>	Test tube method	Meru Central District	The extracts are not toxic to the human cell	[40, 134]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Rynchosia minima</i> DC. (Fabaceae)	Swelling	Roots	The methanolic extract had a ZOI of 11.5 mm compared to 15 mm gentamycin against <i>S. aureus</i>	Disk diffusion technique	Central Kenya	Not reported	[135]
<i>Entada abyssinica</i> (Fabaceae)	Gastrointestinal bacterial infections, bronchitis	Leaves	The methanol leaf extract (100 mg/ml) had ZOI (10, 33 mm) compared to (16.0 mm) zefazidime against <i>S. typhi</i>	Disk diffusion technique	Bondo (Sakwa) in western Kenya	Not reported	[36]
<i>Withania somnifera</i> (Solanaceae)	Microbial infections, cholesterol-lowering	Leaves, roots	The dichloromethane extract (100 mg/ml) had a ZOI of 16.0 mm compared to (18.0 mm) chloramphenicol against <i>S. aureus</i> , ZOI (14.0 mm) compared to (24.0 mm) chloramphenicol against MRSA, and MIC of 6.25 mg/ml against <i>S. aureus</i> and 12.5 mg/ml against MRSA	Disc diffusion test/ broth dilution	Ngong forest	The extract is relatively safe for use even in dose levels exceeding 200 µg/ml	[59]
<i>Thalictrum rhynchocarpum</i> (Ranunculaceae)	Stomach discomfort and bacterial infections	Roots, bark	The root extract had an MIC of 21.5 mg/ml against <i>B. subtilis</i> compared to ciprofloxacin 21.5 mg/ml	Broth dilution	Ngong Forest	Not reported	[136]
<i>Hugonia castaneifolia</i> (Linaceae)	Intestinal worms	Roots	The dichloromethane stem bark extract (100 mg/ml) was active against <i>S. aureus</i> (MIC 0.0008 mg/ml, respectively). Hexane stem bark extracts were active against <i>S. aureus</i> at 0.0031 mg/ml gentamicin and had an MIC of 0.5 mg/ml	Broth dilution	Coast Province of Kenya	Not reported	[29]
<i>Tabernanemontana staphiana Britten</i> (Apocynaceae)	STIs and respiratory-tract infections	Stem bark, root bark, fruits and leaves	The ethanolic root extract (100 mg/ml) had a ZOI of 18.0 mm compared to (22.0 mm) chloramphenicol against MRSA and multiple drug-resistant <i>S. aureus</i> (MDRS) and MIC of 3.9 µg/ml	Disc diffusion test/ broth dilution	Kaptagat Forest in Keiyo District	Not reported	[137]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Rhus vulgaris</i> (Anacardiaceae)	GIT disorders	Leaves, bark	The methanol extract of <i>Rhus vulgaris</i> showed significant antimicrobial activity against MRSA ( $12.00 \pm 0.00$ mm; MIC of $0.391$ mg/ml; minimum bactericidal concentration of $1.563$ mg/ml). Compared to ( $6.00 \pm 0.00$ mm) sulfamethoxazole/trimethoprim ( $23.7 \pm 1.25$ $\mu$ g). The methanol extract showed significant antimicrobial activity against <i>S. aureus</i> ( $19.50 \pm 0.71$ mm; MIC of $0.391$ mg/ml; compared to ( $24.19 \pm 3.60$ mm)) sulfamethoxazole/trimethoprim ( $23.7 \pm 1.25$ $\mu$ g)	Disc diffusion assay/minimum inhibitory concentration assay	Mwala Sub-County, Machakos County	There were no observable adverse effects from oral administration of the extracts (acute oral toxicity testing) at concentrations of $50$ mg/kg, $300$ mg/kg, and $2000$ mg/kg	[138]
<i>Zanthoxylum paracanthum</i> Kokwano (Rutaceae)	Diarrhoea	Root bark	The $\text{CH}_2\text{Cl}_2/\text{CH}_3\text{OH}$ (1 : 1) extract from the root bark had MIC values of $3.91$ , $1.95$ , $0.98$ , and $7.81$ $\mu$ g/ml, against MRSA, <i>E. coli</i> , <i>S. aureus</i> compared to $0.98$ , $0.49$ , and $0.98$ $\mu$ g/ml of omacillin	Minimum inhibitory concentration assay	Mrima Hills, Kwale County in Kenya	Not reported	[139]
<i>Centella asiatica</i> (Apiaceae)	Bacterial infections, diarrhea, skin lesions, psoriasis, keloids	Leaves	Organic crude extract of the leaf showed the highest activity ZOI of $16.33 \pm 0.33$ mm against <i>E. coli</i> compared to tetracycline $26.67 \pm 0.33$ mm	Disc diffusion method	Kisii County	The lethal dose and no observable adverse effect level were $2000$ mg/kg and $1000$ mg/kg	[32, 140]
<i>Aloe vera</i> (Asphodelaceae)	Blood purifier, malaria, skin disease, diabetes	Leaves	The organic extract exhibited a ZOI of $17 \pm 2 - 19 \pm 2$ mm against <i>S. aureus</i> , ( $18 \pm 2 - 20 \pm 1$ mm against <i>B. subtilis</i> ), ( $17 \pm 1 - 19 \pm 3$ mm) against <i>K. pneumoniae</i> , ( $16 \pm 1 - 20 \pm 3$ mm) against <i>E. coli</i>	Agar diffusion method	Department of biological sciences, Egerton university	Not reported	[141]
<i>Aloe volkensii</i> (Asphodelaceae)	Laxative, burns, wounds and sores	Leaves	The organic extract exhibited activity against <i>S. aureus</i> ( $19 \pm 1 - 20 \pm 2$ mm), <i>B. subtilis</i> ( $17 \pm 2 - 21 \pm 3$ mm), <i>K. pneumoniae</i> ( $18 \pm 2 - 19 \pm 1$ mm), <i>E. coli</i> ( $18 \pm 2 - 19 \pm 3$ mm)	Agar diffusion method	Department of Biological Sciences, Egerton University	Not reported	[141]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Senna spectabilis</i> (Fabaceae)	Laxatives	Leaves, pods	The organic leaf extract (100 mg/ml) had a ZOI of $9.6 \pm 0.6$ compared to chloramphenicol ( $11.7 \pm 2.3$ ) against <i>S. typhi</i>	Agar diffusion method	Mbeere North District, Embu County	Not reported	[142]
<i>Maytenus putterlickioides</i> (Celastraceae)	Malaria, emmenagogue, aphrodisiac	Roots	The methanol root extract (100 mg/ml) had a ZOI of $9.2 \pm 1.1$ compared to chloramphenicol ( $11.7 \pm 2.3$ ) against <i>S. typhi</i>	Agar diffusion method	Mbeere North District, Embu county	Not reported	[142]
<i>Olinia usambarensis</i> (Oliniaceae)	Malaria, abscess, cough, measles	Bark, roots, leaves	The methanol leaf extract (100 mg/ml) had ZOI ( $12.2 \pm 0.8$ ) compared to chloramphenicol ( $11.7 \pm 2.3$ ) against <i>S. typhi</i>	Agar diffusion method	Mbeere North District, Embu County	Not reported	[142]
<i>Crotalaria goodformis Vatke.</i> (Fabaceae)			The aqueous extract (1 g/ml) had a ZOI of $14.8 \pm 0.2$ mm compared to ( $18.0 \pm 0.1$ ) streptomycin (25 $\mu$ g/ml) against <i>S. aureus</i>	Disk diffusion method	Mbeere, and Embu-Eastern Province	Not reported	[48]
<i>Prosopis juliflora</i> (Sw.) DC (Fabaceae)	Open wounds and dermatological ailments.	Leaves	The ethanolic leaf extract (100 mg/ml) had a ZOI of $20.00 \pm 1.00$ mm compared to (19.0) erythromycin (1.5 $\mu$ g/ml) and (30.0 mm) chloramphenicol (30 $\mu$ g/ml) against <i>E. coli</i> , ZOI ( $15.33 \pm 0.58$ mm) compared to (11.0) erythromycin (15 $\mu$ g/ml) and (22.0 mm) chloramphenicol (30 $\mu$ g/ml) against <i>P. aeruginosa</i>	Disk diffusion method	Endao, Marigat District, in Baringo County	The pods are toxic, mainly for cattle and goats, and have piperidine alkaloids and can cause neurotoxicity	[143, 144]
<i>Oxypolis abyssinica</i> (Santalaceae)	Dysentery, typhoid	Roots	The aqueous root extract (1 g/ml) had ZOI ( $15.2 \pm 0.7$ mm) compared to ( $18.0 \pm 0.1$ ) streptomycin (25 $\mu$ g/ml) against <i>S. aureus</i> , ZOI ( $14.8 \pm 0.3$ mm) compared to ( $16.0 \pm 0.2$ ) streptomycin (25 $\mu$ g/ml) against <i>E. coli</i> , ZOI ( $15.5 \pm 0.5$ mm) compared to ( $15.0 \pm 0.3$ mm) streptomycin 25 $\mu$ g/ml against <i>B. subtilis</i>	Disk diffusion method	Mbeere, and Embu-eastern province	Not reported	[48]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Abrus precatorius</i> (Fabaceae)	Gonorrhoea, coughs in children	Leaves, roots	The aqueous leaf extract (1 g/ml) had a ZOI of 15.7 ± 0.5 mm compared to (18.0 ± 0.1) streptomycin (25 µg/ml) against <i>S. aureus</i> . The aqueous bark extract (1 g/ml) had a ZOI of 7.2 ± 0.8 mm compared to (16.0 ± 0.2) streptomycin (25 µg/ml) against <i>E. coli</i> , ZOI (15.5 ± 0.5 mm) compared to (10.7 ± 1.2 mm) streptomycin 25 µg/ml against <i>B. subtilis</i>	Disk diffusion method	Mbeere, and Embu-Eastern Province	It contains abrin, a toxalbumin that inhibits protein synthesis causing cell death, especially seeds	[48, 145]
<i>Ormocarpum trichocarpum</i> (Fabaceae)	Bone setting	Roots	The methanol extract (1 g/ml) had a ZOI of 15.5 mm compared to (26.0 mm) gentamycin (10 µg/ml) against <i>B. subtilis</i>	Disk diffusion method	Kisii South	Not reported	[30]
<i>Psidium guajava</i> (Myrtaceae)	Wounds, ulcers, cholera	Leaves	The methanol extracts (1 g/ml) had a ZOI of 19.7 mm compared to gentamycin (10 µg/ml) against <i>S. aureus</i> , ZOI (16.0 mm) compared to gentamycin (10 µg/ml) 19 mm against <i>E. coli</i> and, ZOI (16 mm) compared to 10 µg/ml gentamycin (17 mm) against <i>P. aeruginosa</i>	Agar well diffusion	Bomet District	The median lethal dose ( $LD_{50}$ ) of bark extract is greater than 5000 mg/kg body weight	[46, 145]
<i>Cyathula polycarpa</i> (Amaranthaceae)	Diabetes, skin infections, pneumonia	Stem barks	The methanol extract 100 mg/ml had a ZOI of 14.2 mm compared to 30 µg/ml gentamycin (19.0 mm) against <i>S. aureus</i> , ZOI (16.0 mm) compared to 30 µg/ml gentamycin (9.0 mm) against MRSA, ZOI (10.0 mm) compared to 30 µg/ml gentamycin (21.0 mm) against <i>P. aeruginosa</i>	Disk diffusion method	Bomet District	The methanol extract was very safe with a CC <sub>50</sub> of 100%, while water extract were toxic with CC <sub>50</sub> of 23.75% and 31.56% as compared to the positive control Chloroquine with CC <sub>50</sub> of 25.28 and 51.94% at concentration 1000 and 100 mg mL <sup>-1</sup>	[29]
<i>Blumea axillaris</i> (Lam.) DC. (Asteraceae)	Skin disease	Aerial parts	Methanol extracts had a ZOI 16.41 ± 0.31 compared to chloramphenicol (21.7 ± 0.11) against <i>S. aureus</i>	Agar well diffusion	Vihiga County, Western Kenya	None reported	[45]
<i>Chamaecrista mimosoides</i> (L.) Greene (Fabaceae)	Respiratory system disorders, dysentery	Aerial parts, roots	The aqueous extracts had a ZOI 30.00 ± 1.46 compared to chloramphenicol (21.7 ± 0.11) against <i>S. aureus</i>	Agar well diffusion	Vihiga County, Western Kenya	Not reported	[45]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Lantana trifolia L.</i> (Verbenaceae)	Cough and common colds	Aerial parts	The methanol extracts had a ZOI of 20.59 ± 0.92 compared to chloramphenicol (21.7 ± 0.11) against <i>S. aureus</i>	Agar well diffusion	Vihiga County, Western Kenya	The ethanol extracts of <i>Lantana trifolia</i> ( $LC_{50}$ 32.3 $\mu$ g/ml exhibited mild toxicity and are safe for short-term use (Moshi et al., 2010)	[45]
<i>Terminalia kilimandscharica Engl.</i> (Combretaceae)	Cough, sexually transmitted diseases	Barks	The methanolic bark extract had a MIC of 25, 15.6, 37.5, and 150 mg/ml against <i>S. aureus</i> , <i>B. cereus</i> , <i>P. aeruginosa</i> , and <i>E. coli</i> , respectively, compared to that of 0, 0, 0.25, 0.25 mg/ml of streptomycin against <i>S. aureus</i> , <i>B. cereus</i> , <i>P. aeruginosa</i> , and <i>E. coli</i> , respectively, and benzylpenicillin 0.6 and 0.6 against <i>S. aureus</i> and <i>B. cereus</i> , respectively.	Broth dilution	Machakos and Kitui	The methanolic bark extracts had an $LC_{50}$ of <1000 $\mu$ g/ml, which is considered relatively nontoxic	[39]
<i>Pentas lanceolata</i> (Rubiaceae)	Genital and oral thrush	Roots	The ethyl acetate extract (100 mg/ml) had a ZOI of (10.96 ± 0.08 mm) compared to gentamycin (23.88 ± 0.01 mm) against <i>K. pneumoniae</i> , ZOI of (12.08 ± 0.26 mm) compared to gentamycin (23.88 ± 0.01 mm) against <i>Escherichia coli</i> and ZOI of (11.39 ± 0.6 mm) compared to gentamycin (25.9 ± 0.01 mm) against <i>S. aureus</i> .	Agar well diffusion	Magadi, Kajiado District of Kenya	Not reported	[44]
<i>Senecocompositus hildebrandtii</i> Schinz (Amaranthaceae)	Purgative	Roots	The ethyl acetate root extract (100 mg/ml) had a ZOI of 11.28 ± 0.09 mm compared to gentamycin (0.1 $\mu$ g/ml), (23.88 ± 0.01 mm) against <i>K. pneumoniae</i> , ZOI of (10.33 ± 0.06 mm) compared to gentamycin (0.1 $\mu$ g/ml) (23.88 ± 0.01) against <i>E. coli</i> , and ZOI of (10.66 ± 0.18) compared to gentamycin (0.1 $\mu$ g/ml) (25.9 ± 0.01) against <i>S. aureus</i>	Agar well diffusion	Magadi, Kajiado District of Kenya	Not reported	[44]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Combretum molle R.Br. ex G.Don</i> (Combretaceae)	Tooth brush, stomach ache, and dysentery	Stem bark	The ethanolic stem bark extract (0.5 mg) exhibited ZOI (mm) of 7.6 ± 0.24 against <i>P. aeruginosa</i> , 15.4 ± 0.3 against <i>E. coli</i> , and 2.2 ± 0.4 against <i>S. aureus</i> compared to Augmentin 8.0 ± 0.02 against <i>P. aeruginosa</i> , 19.0 ± 0.03 against <i>E. coli</i> and 17.0 ± 0.02 against <i>S. aureus</i>	Disc diffusion method	Mwingi District in Kitui County	For the acute toxicity test, no death and signs of poisoning were observed in the treated groups. In the subacute study, LD <sub>50</sub> in the rats after intraperitoneal administration was 700 mg/kg	[76, 146]
<i>Combretum illairii</i> (Combretaceae)	Skin infections, wounds dressings and ointments	Roots, stems, leaves	The methanol leaf extract (100 mg/ml) had ZOI of 15.60 mm and 17.00 mm against <i>S. aureus</i> and <i>P. aeruginosa</i> , respectively, against gentamycin (30 µg/ml) 25.3 mm and 18 mm	Disc diffusion assay	Kilifi District	The stem bark extracts had neither cytotoxicity nor brine shrimp lethality. plant extracts [29]	[29]
<i>Combretum tanaense</i> (Combretaceae)		Roots	The methanol root extract (100 mg/ml) had a ZOI of 11.50 ± 0.5 mm compared to ciprofloxacin (0.32 µg/ml) 14.75 ± 0.25 mm against <i>S. aureus</i> and ZOI of 12.25 ± 0.25 mm compared to ciprofloxacin (0.32 µg/ml) 16.00 ± 0.00 mm against <i>K. pneumoniae</i>	Agar well diffusion assay	Mount Kenya University Botanical Garden, Thika	Not reported	[104]
<i>Vernonia brachyalyx</i> (Asteraceae)	Antimalarial, emetic	Stem, leaves	The ethanol extract had a ZOI of 9.5 ± 1.2 mm against <i>S. pneumoniae</i> compared to 8.2 ± 0.6 (erythromycin) 7.2 ± 0.5 (gentamycin 15 µg)	Disk diffusion method	Eastern Kenya	An isolated compound (16,17-dihydrobrachyalyxolide) displayed high toxicity against human lymphocytes	[18]
<i>Vernonia amygdalina</i> (Asteraceae)	Stomach discomfort and bacterial infections	Leaves	The methanol leaf extract (1 g/ml) exhibited a ZOI 17.0 mm compared to gentamycin (10 µg/ml) 19 mm against <i>S. aureus</i>	Agar well diffusion	Bomet District	The extract had an LD <sub>50</sub> of 288.5 mg/kg body weight. It has relative toxicity [147]	[46]

TABLE 1: Continued.

Plant	Ethnopharmacological use	Part used	Bioactivity	Assay method used	Collection site in Kenya	Side effects/contraindications/ toxicity	References
<i>Vernonia glabra</i> (Steetz) Oliv. & Hiern (Asteraceae)	Gastrointestinal problems , snake bites	Leaves, roots	The dichloromethane/methanol leaf extract (1 g/ml) exhibited a ZOI of 1.85 cm against <i>S. aureus</i> compared to streptomycin with a ZOI of 1.30 cm. dichloromethane/methanol extract of flower showed significant activity only against <i>S. aureus</i> , with the lowest MIC of 1.5625 mg/100 µl, compared to streptomycin with a MIC of 6.25 mg/100 µl	Disc diffusion	Machakos	Not reported	[27]
<i>Vernonia adonis</i> (Asteraceae)	Oral health	Stem bark	The methanol stem bark extract (100 mg/ml) exhibited a ZOI of 13.00 ± 0.577 mm against <i>E. aerogenes</i> , 11.00 ± 0.577 mm against <i>S. pyogenes</i> , 9.67 ± 0.333 mm against <i>S. epidermidis</i> , and 9.00 ± 0.577 mm against <i>E. faecalis</i> . The acetone stem bark extract (100 mg/ml) exhibited a ZOI of 16.00 ± 0.577 mm against <i>E. aerogenes</i> , 11.33 ± 0.882 mm against <i>S. epidermidis</i> , and 10.00 ± 0.577 mm against <i>S. faecalis</i> . Penicillin (100 mg/ml) exhibited a ZOI of 38.00 ± 0.577 mm against <i>E. faecalis</i> , 43.33 ± 0.882 against <i>S. pyogenes</i> , 19.33 ± 0.333 against <i>S. epidermidis</i> , and 36.33 ± 0.882 against <i>E. aerogenes</i>	Disc diffusion method	University of Eastern Africa, Baraton, Nandi County	Not reported	[148]
<i>Vernonia hymenolepis</i> (Asteraceae)	Infections, toothache	Leaves	The MBC and MIC values of aqueous leaf extract was 400 mg/ml against <i>S. aureus</i> , while the DCM/methanol leaf extract had MIC and MBC of 400 mg/ml against <i>P. aeruginosa</i> and <i>E. coli</i> , and MIC of 100 mg/ml against <i>S. aureus</i> . Amoxicillin had MIC and MBC of 3.125 mg/ml and 6.25 mg/ml against <i>E. coli</i> , respectively	Broth dilution	Trans Nzoia County	Not reported	[149]

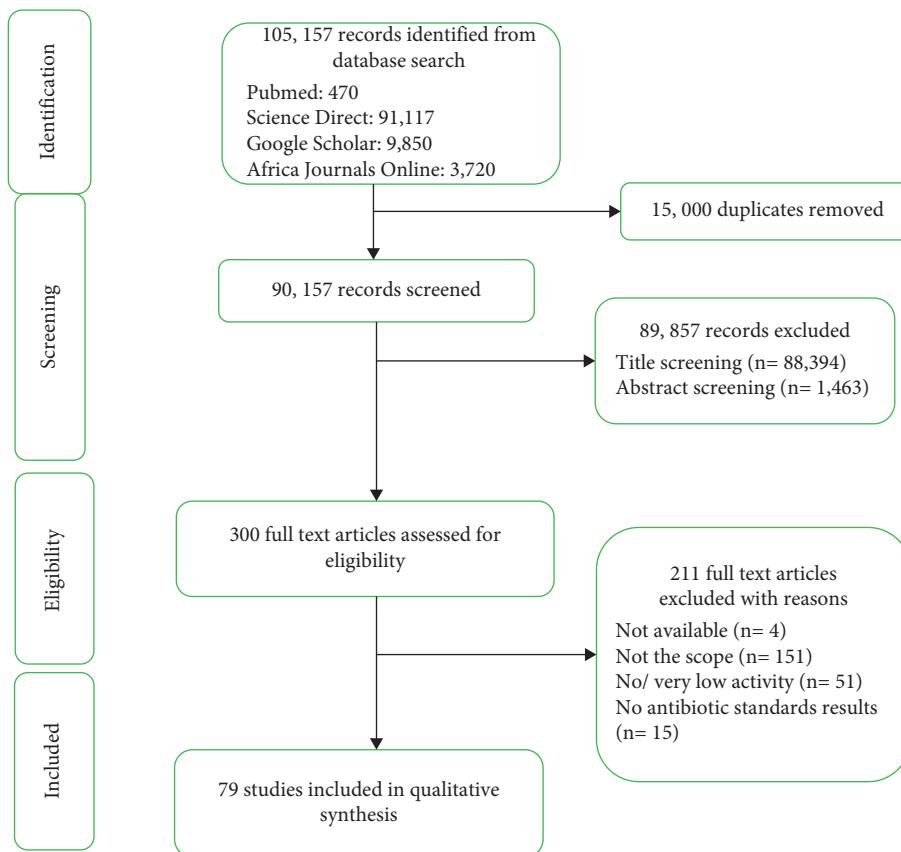


FIGURE 1: Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement of search results.

Water and methanol were the most used solvents for plant extract preparation, whereas ethanol and dichloromethane were the least utilized solvents used (Table 1).

The reported medicinal plants were commonly used in the treatment of STIs, respiratory diseases, diarrhea, and oral infections (Table 1). (Figure 1).

#### 4. Discussion

Plants generally accumulate diverse bioactive compounds in varying concentrations in the different parts of a plant, and this eventually affects the efficacy of medicinal plants. The leaves (37%), bark/stem bark (47%), and roots (24%) were the most utilized plant parts against bacterial infections. The variances in their antimicrobial activities could be due to the synergistic or antagonistic actions of various secondary metabolites present [60].

**4.1. Nutraceuticals.** From the review, several common foods/spices are reported to have potential antibacterial benefits. For example, green tea (*Camellia sinensis*) that is often famed for its antioxidant activity, exhibited good antibacterial activity zone of inhibition (ZOI) of  $21.3 \pm 0.33$  mm against *E. coli* and a ZOI of  $22.3 \pm 0.50$  mm against *S. aureus* compared to gentamicin  $22.3 \pm 0.50$  mm (against *E. coli*) and ZOI  $23.7 \pm 0.33$  mm (against *S. aureus*) at a concentration of 0.1 mg/ml [67]. The aqueous crude green tea extracts at a concentration of 400 mg/ml exhibited ZOI of

$20 \pm 0.0$  mm which was similar to that of streptomycin against *S. aureus*. The extract also displayed a ZOI of  $18 \pm 0.0$  mm against *E. coli* compared to ZOI of  $10 \pm 0.0$  mm of streptomycin against *E. coli*  $20 \pm 0.0$  mm, ZOI of  $18 \pm 0.0$  mm, and MIC 200 mg/ml against *E. coli* compared to streptomycin  $10 \pm 0.0$  mm [69]. The Bambara nut (*Vigna subterranea*) had an MIC value of  $7.72 \pm 0.35 \mu\text{g}/\text{ml}$  for *E. coli*,  $12.5 \pm 0.32 \mu\text{g}/\text{ml}$  for *S. aureus* and  $7.95 \pm 0.10 \mu\text{g}/\text{ml}$  for *P. aeruginosa* at 100  $\mu\text{g}/\text{ml}$ , and showed zone of inhibition of  $27 \pm 0.74$  mm,  $25.3 \pm 0.40$  mm, and  $25.1 \pm 0.24$  mm *E. coli*, *S. aureus*, and *P. aeruginosa*, respectively [109].

**4.2. Complementary Medicine.** As has been shown in previous ethnomedical surveys by Omwenga et al., traditional medicine is widely practiced in Kenya and is culturally acceptable. It is estimated that about 75% population in Kenya seeks health care among traditional healers [8–10, 150]. In certain instances, people utilize both traditional and modern medicine simultaneously. Njoroge and Kibunga noted that herbal products were used as complementary therapy in the management of diarrhea by residents in Thika, Kenya [151]. The lack of enquiry about Traditional Complementary and Alternative Medicine (TCAM) use and the conventional healthcare providers' negative attitude towards TCAM were cited as some of the reasons why patients fail to reveal their TCAM use [152].

The regulatory framework for the practice of traditional medicine in Kenya is still underway [153], but several crude drugs or formulated herbal products with reported antibacterial activity are already available in the Kenyan market, for example, the Lifebuoy germ protection antibacterial herbal hand and body soap and the Dettol herbal bar soap. Skin care products (soaps and lotions) formulated from plant extracts (*Thevetia peruviana*, *Tithonia diversifolia*, *Azadirachta indica*, *Aloe secundiflora*) had antimicrobial properties. Soap made from *Tithonia diversifolia* plant extract was the most effective against *E. coli*, while *Azadirachta indica* soap was the most effective against *C. albicans*. *T. diversifolia* soap exhibited the highest activity against *E. coli* [154].

The ethanolic extract of *E. divinorum* root bark had a MIC of 25, 50, and 25 µg/ml for *Streptococcus pyogenes*, *Staphylococcus aureus*, and *Escherichia coli*, respectively (Table 1). A herbal toothpaste formulated with the ethanol extract of *E. divinorum* root bark had a higher antimicrobial activity against the tested microorganisms compared to Colgate herbal toothpaste formulated with fluoride [77]. Also, the formulation containing the aqueous extracts of *T. asiatica* (50 mg/ml) stem bark exhibited pronounced antimicrobial activity as indicated by zone of inhibition diameters of 24 mm (MRSA) and 22 mm (*M. gypseum*) compared to 22 mm and 14 mm, respectively, by the commercial hand wash (50 mg/ml). In the model hand washes efficacy experiment, the formulated herbal detergent attained a 78.8% reduction of pathogenic load as compared to 67.9% reduction with the commercial hand wash [37].

Unfortunately, despite the surge in the consumption of herbal products and the limited number of standardized herbal products in the Kenyan market, the pharmacovigilance for herbal medicines is nonexistent in Kenya [155]. The increased demand for herbal products has resulted in the market being flooded with adulterated products and false herbal claims on the products' labels for marketing purposes. For instance, 50% of the investigated products by Ngari et al. [75] lacked antimicrobial activity against test bacteria (*Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Proteus vulgaris*, *Bacillus subtilis*, *Candida albicans*, *Escherichia coli*, *Streptococcus mutans*, *Enterococcus faecalis*, and *Lactobacillus acidophilus*). The evaluation of herbal suspensions (used in the management of oral health in Nairobi County, Kenya) by Ngari et al. [75] reported a lack of detectable phytochemicals in the suspensions and noted that this occurrence could be due to very low concentrations of phytochemicals that could not be detected by standard laboratory methods or mineral adulterants might have been used. Studies have demonstrated that antimicrobial properties of natural products can be enhanced by the addition of metal ions [14, 156].

**4.3. Polyherbalism.** The use of herbs as combinations is common practice with many herbal practitioners and aimed at giving better results as compared to single herbs and also treating more than one ailment [157]. Ngari et al. [75] evaluated herbal pastes and suspensions used in the

management of oral health in Nairobi County, Kenya, and various products showed significant antimicrobial activity that is comparable to positive control. For example, product HS4 composed of *W. ugandensis*, *M. piperita*, and *S. aromaticum* had ZOI of  $33.1 \pm 0.85$  mm,  $20.3 \pm 1.71$  mm, and  $19.3 \pm 1.65$  mm against *E. coli*, *S. aureus*, and *P. aeruginosa*, respectively, compared to that of co-trimoxazole of  $27.5 \pm 0.7$  mm,  $8.5 \pm 0.5$  mm, and  $10 \pm 0.0$  mm, respectively. Product HS5 composed of *Aloe vera gel*, *W. ugandensis*, and *W. sominifera* had a ZOI of  $25.3 \pm 0.25$  mm,  $20.25 \pm 1.26$  mm, and  $21.5 \pm 1.71$  mm against *E. coli*, *S. aureus*, and *P. aeruginosa*, respectively, compared to co-trimoxazole  $27.5 \pm 0.7$  mm,  $8.5 \pm 0.5$  mm, and  $10 \pm 0.0$  mm, respectively. This biological activity is attributed to the presence of various secondary metabolites in plants [14].

Mbuthia et al. evaluated the synergistic properties of water-soluble green and black tea extracts with penicillin G. The antimicrobial results showed a marked increase in the inhibition zone diameters on the combination of green tea extracts with penicillin G. The catechins, theaflavins, and thearubigins are the antimicrobial agents present in tea [67]. Synergistic inhibition by green tea extracts and penicillin G is due to the presence of dual binding sites on the bacterial surface for antibiotic and tea extract [158].

**4.4. Bioactivity/Assay Methods.** Disc diffusion method was the preferred method to assay for antibacterial activity. A clearing zone of 9 mm or greater for Gram-positive and Gram-negative bacteria was used as the criterion for designating significant antibacterial activity [159]. The in vitro MIC results were classified as described in the study by Pessini et al., 2003: the antimicrobial activity of the extracts that displayed MIC lower than 100 µg/ml was considered very high; 100–500 µg/ml, high; 500–1000 µg/ml, moderate; 1000–4000 µg/ml, low; and anything above this, inactive. The plants with the strongest antimicrobial activities were *Toddalia asiatica*, *Hagenia abyssinica*, *Ocimum gratissimum*, *Harrisonia abyssinica*, *Conyza sumatrensis*, *Senna didymobotrya*, *Aloe secundiflora*, *Olea Africana*, *Vernonia glabra*, *Camellia sinensis*, *Tetradenia riparia*, and *Tamarindus indica* as they exhibited high mean inhibition zone values or low minimum inhibitory concentration (MIC) values.

Several plants exhibited a high activity superior or comparable to the standard antibiotic drugs (Table 1); the methanol-dichloromethane extract (100 mg/ml) of *Harrisonia abyssinica* had a ZOI of  $20 \pm 1.6$  mm compared to gentamycin (ZOI of  $18 \pm 1.2$  mm) against *S. aureus* and a ZOI of  $30 \pm 1.7$  mm against *E. coli* compared to gentamycin (ZOI of  $15.1 \pm 3$  mm) against *E. coli* [41]. The methanolic extract of *Croton macrostachyus* exhibited ZOI (23.66 mm) compared to (21.33 mm) amoxicillin against *S. aureus* and ZOI (18.0 mm) compared to (17.58 mm) amoxicillin against *P. aeruginosa*. The methanolic extract had an MIC of 37.50 mg/ml compared to 18.75 mg/ml cefpodoxime against *S. aureus* and 18.75 mg/ml compared to 9.372 mg/ml cefpodoxime against *P. aeruginosa* [79].

Plants such as *Toddalia asiatica*, *Hagenia abyssinica*, *Senna didymobotrya*, *Aloe secundiflora*, and *Camellia sinensis* displayed good activities; thus, they may be considered for the assessment of *in vivo* activity and possibly formulated into different consumable forms. Korir et al. recommended that for plants with very low or no activity, bioactivity on all parts of the plants, for example, root, stem bark, and leaves combined ought to be done against a wide variety of pathogenic bacteria in order to conclusively report that a certain plant is inactive [29].

**4.5. Toxicity.** Despite herbal remedies being affordable, their lack of efficacy and safety evaluation is a great impediment to their acceptance into mainstream medicine. The safety assessment of herbal remedies remains a challenge as most of the studies of herbal medicines are directed at the toxicological properties of single plant formulations, yet most herbal preparations, especially those used in traditional medicine, contain multiple herbs [160].

From this review, 45% of the plants were relatively safe, 44% of the plants have not been assessed for their safety, and 11% of the plants were reported to have high toxicity (Table 1). The plants with very high toxicity can be further explored for the antitumor activity or as insecticides.

**4.6. Plant Conservation Status.** Other than *W. ugandensis* and *Prunus africana*, most of the plants identified in this review are largely available and are not under any serious threat to become extinct. Since most of them are obtained from wild habitats, sustainable use of the reported medicinal plants against bacterial infections is advised as a conservation measure. The cultivation of wild medicinal plants is an important approach to safeguard the herbal industry. Biotechnological techniques such as plant cell or tissue culture, biochemical conversions, and clonal propagation of indigenous medicinal plants are another potential strategy in improving herbal medicine [161].

## 5. Conclusion

This review demonstrates the potential of medicinal plants to treat bacterial infections alongside justifying the use of these plant traditional medicine. It may serve as a starting point of research geared towards the clinical application of these plants. There is a need for standardization to improve the acceptance of herbalism by mainstream health practitioners.

## 6. Recommendation

Further research into the *in vivo* activity of plants displayed remarkable *in vitro* activity. Plants exhibiting strong antibacterial activity can be evaluated for their interactions with conventional antibiotics, and those displaying synergistic activity may provide useful leads in antibiotic therapy.

## Data Availability

No data were used in the study.

## Conflicts of Interest

The authors declare that they have no conflicts of interest.

## References

- [1] M. Eryilmaz, M. E. Bozkurt, M. M. Yildiz, and A. Akin, “Antimicrobial resistance of urinary *Escherichia coli* isolates,” *Tropical Journal of Pharmaceutical Research*, vol. 9, pp. 205–209, 2010.
- [2] J. Emacar, P. Okemo, G. Gatheri, and S. Kariuki, “Antibiotic resistance patterns of *Escherichia coli* isolated from HIV-sero positive adults at Mbagathi district hospital, Nairobi, Kenya,” *Journal of Applied Biosciences*, vol. 27, pp. 1705–1714, 2010.
- [3] UNICEF Report, *Maternal, Newborn and Child Health Working Paper on Access to Healthcare through Community Health Workers in East and Southern Africa United Nations Children’s Fund (UNICEF)*, UNICEF, New York, NY, USA, 2014.
- [4] Government of Kenya, *National Policy for the Prevention and Containment of Antimicrobial Resistance*, Government of Kenya, Nairobi, Kenya, 2017.
- [5] World Health Organization, *WHO Global Report on Traditional and Complementary Medicine*, World Health Organization, Geneva, Switzerland, 2019.
- [6] E. Rukangira, “The African herbal Industry: constraints and challenges,” in *Proceedings of the Natural Products and Cosmeceuticals Conference*, pp. 1–23, Erboristeria, Domani, 2001.
- [7] W. Wanzala and M. K. Walingo, “Ethnomedicines and health management in Kenya: which way forward?” *Journal of Complementary Medicine and Alternative Healthcare*, vol. 10, no. 3, 2019.
- [8] E. O. Omwenga, A. Hensel, A. Shitandi, and F. M. Goycoolea, “Ethnobotanical survey of traditionally used medicinal plants for infections of skin, gastrointestinal tract, urinary tract and the oral cavity in Borabu sub-county, Nyamira county, Kenya,” *Journal of Ethnopharmacology*, vol. 176, pp. 508–514, 2015.
- [9] J. Kimondo, J. Miaron, P. Mutai, and P. Njogu, “Ethnobotanical survey of food and medicinal plants of the Ilkisonko Maasai community in Kenya,” *Journal of Ethnopharmacology*, vol. 175, pp. 463–469, 2015.
- [10] F. M. Mutie, L. L. Gao, V. Kathambi et al., “An ethnobotanical survey of a dryland botanical garden and its environs in Kenya: the Mutomo hill plant sanctuary,” *Evidence-Based Complementary and Alternative Medicine*, vol. 2020, Article ID 1543831, 22 pages, 2020.
- [11] O. Oniyangi and D. H. Cohall, “Phytomedicines (medicines derived from plants) for sickle cell disease,” *Cochrane Database of Systematic Reviews*, vol. 2, Article ID CD004448, 2018.
- [12] S. N. Njeru, J. Matasyoh, C. G. Mwaniki, and K. Kobia, “A review of some phytochemicals commonly found in medicinal plants,” *International Journal of Medicinal Plants Photon*, vol. 105, pp. 135–140, 2013.
- [13] K. Godin, J. Stapleton, S. I. Kirkpatrick, R. M. Hanning, and S. T. Leatherdale, “Applying systematic review search methods to the grey literature: a case study examining guidelines for school-based breakfast programs in Canada,” *Systematic Reviews*, vol. 4, no. 1, p. 138, 2015.
- [14] R. W. Scherer and I. J. Saldanha, “How should systematic reviewers handle conference abstracts? a view from the trenches,” *Systematic Reviews*, vol. 8, no. 1, p. 264, 2019.

- [15] W. M. Bramer, G. B. de Jonge, M. L. Rethlefsen, F. Mast, and J. Kleijnen, "A systematic approach to searching: an efficient and complete method to develop literature searches," *Journal of the Medical Library Association*, vol. 106, no. 4, pp. 531–541, 2018.
- [16] G. Alebie, B. Urga, and A. Worku, "Systematic review on traditional medicinal plants used for the treatment of malaria in Ethiopia: trends and perspectives," *Malaria Journal*, vol. 16, no. 1, pp. 307–316, 2017.
- [17] N. T. Huy, T. Van Giang, D. H. D. Thuy et al., "Factors associated with dengue shock syndrome: a systematic review and meta-analysis," *PLoS Neglected Tropical Diseases*, vol. 7, no. 9, Article ID e2412, 2013.
- [18] E. K. Amadi, P. G. Kareru, J. M. Keriko, and J. Kiptoo, "Antimicrobial activity of selected medicinal plants' extracts against *Streptococcus pneumonia*," *International Journal of Botany Studies*, vol. 1, no. 1, pp. 20–22, 2016.
- [19] P. N. Waithaka, E. M. Gathuru, B. M. Githaiga, and R. Z. Kazungu, "Antimicrobial properties of *Aloe vera*, *Aloe volkensii* and *Aloe secundiflora* from Egerton university," *Acta Scientific Microbiology*, vol. 1, pp. 06–10, 2018.
- [20] H. O. Rachuonyo, P. E. Ogola, W. M. Arika, and J. R. Wambani, "Efficacy of crude leaf extracts of *Aloe secundiflora* on selected enteric bacterial pathogens and *Candida albicans*," *Journal of Antimicrobiology*, vol. 2, p. 112, 2016.
- [21] R. M. Mariita, J. A. Orodho, P. O. Okemo, C. Kirimuhuza, J. N. Otieno, and J. J. Magadula, "Methanolic extracts of *Aloe secundiflora* Engl. inhibits in vitro growth of tuberculosis and diarrhea-causing bacteria," *Pharmacognosy Research*, vol. 3, pp. 95–99, 2011.
- [22] H. R. Opinde, G. W. Gatheri, and A. K. Nyamache, "Antimicrobial evaluation of crude methanolic leaf extracts from selected medicinal against *Escherichia coli*," *Journal of Bacteriology and Parasitology*, vol. 7, p. 272, 2016.
- [23] T. O. Elufioye, O. I. Alatise, F. A. Fakoya, J. M. Agbedahunsi, and P. J. Houghton, "Toxicity studies of *Tithonia diversifolia* A. Gray (Asteraceae) in rats," *Journal of Ethnopharmacology*, vol. 122, no. 2, pp. 410–415, 2009.
- [24] O. E. Omwenga, R. M. Mariita, L. Alaro, and P. O. Okemo, "Evaluation of methanolic extracts of six medicinal plants used by herbal practitioners in central province-Kenya," *International Journal of Pharmaceutical Sciences and Research*, vol. 2, no. 4, pp. 867–874, 2011.
- [25] P. M. Ogoti, *Bioprospecting for Effective Antibiotics from Selected Kenyan Medicinal Plants against Four Clinical Salmonella Isolates* Jomo Kenyatta University of Agriculture and Technology, Juja, Kenya, 2017, <https://ir.jkuat.ac.ke/handle/123456789/3432>.
- [26] K. Douglas and J. Jeruto, "Phytochemistry and antimicrobial activity of extracts from medicinal plants *Tithonia diversifolia* and *Olea africana*," *British Journal of Pharmaceutical Research*, vol. 12, no. 3, pp. 1–7, 2016.
- [27] C. K. Kitonde, D. S. Fidahusein, C. W. Lukhoba, and M. M. Jumba, "Antimicrobial activity and phytochemical screening of *Senna didymobotrya* used to treat bacterial and fungal infections in Kenya," *International Journal of Educational Research*, vol. 2, no. 1, 2014.
- [28] L. B. Nyamwamu, M. Moses, M. M. Ngeiywa et al., "Cytotoxicity and in vitro antiamoebic activity of senna didymobotrya crude root extracts in comparison with metronidazole against entamoeba histolytica," 2015, <https://www.semanticscholar.org/paper/CYTOTOXICITY-AND-IN-VITRO-ANTIAMOEBIC-ACTIVITY-OF-Nyamwamu-Ngeiywa/65a5ea31bca67ec6a9b555dbbf689addfb3e791>.
- [29] R. K. Korir, C. Mutai, C. Kiiyukia, and C. Bii, "Antimicrobial activity and safety of two medicinal plants traditionally used in Bomet district of Kenya," *Research Journal of Medicinal Plant*, vol. 6, no. 5, pp. 370–382, 2012.
- [30] J. India, "Efficacy of some medicinal plants used in various parts of Kenya in treating selected bacterial and fungal pathogens," 2015, <https://irlibrary.ku.ac.ke/handle/123456789/14965>.
- [31] P. Jeruto, P. F. Arama, B. Anyango, and G. Maroa, "Phytochemical screening and antibacterial investigations of crude methanol extracts of *Senna didymobotrya* (Fresen.) H. S. Irwin & Barneby," *Journal of Applied Biosciences*, vol. 114, pp. 11357–11367, 2017.
- [32] E. O. Omwenga, F. M. Goycoolea, A. Hensel, and A. Shitandi, "Antimicrobial, cytotoxicity and preliminary phytochemical determination of commonly used medicinal plants to treat oral cavity, urinary tract and gut infections by inhabitants of Borabu sub-county, Nyamira County, Kenya," *Malaysian Journal of Microbiology*, vol. 16, no. 4, pp. 312–322, 2020.
- [33] J. A. Orwa, L. Ngeny, N. M. Mwikwabe, J. Ondicho, and I. J. O. Jondiko, "Antimalarial and safety evaluation of extracts from *Toddalia asiatica* (L) Lam. (Rutaceae)," *Journal of Ethnopharmacology*, vol. 145, no. 2, pp. 587–590, 2013.
- [34] M. M. Gakuubi, K. N. Micheni, and W. Wanzala, "In vitro antibacterial activity of essential oil from the fruits of *Toddalia asiatica* (L) Lam. (Rutaceae)," *Journal of Biologically Active Products from Nature*, vol. 7, no. 1, pp. 52–61, 2017.
- [35] I. Onjero, "Antimicrobial activity and interaction of *Toddalia asiatica* coumarins with two known-drugs," 2020, <https://irlibrary.mmust.ac.ke/xmlui/bitstream/handle/123456789/1436/Isaiah%20Onjero.pdf?sequence=1&isAllowed=y>.
- [36] R. M. Mariita, C. K. P. Ogol, N. O. Oguge, and P. O. Okemo, "Antitubercular and phytochemical investigation of methanol extracts of medicinal plants used by the Samburu community in Kenya," *Tropical Journal of Pharmaceutical Research*, vol. 9, no. 4, pp. 379–385, 2010.
- [37] W. L. L. Munyendo and A. K. Kiprop, "Design, preparation and evaluation of germicidal *Toddalia asiatica* herbal anti-septic detergent," *Journal of Applied Pharmaceutical Science*, vol. 6, no. 11, 2016.
- [38] A. O. Maima and W. L. L. Munyendo, "Antimicrobial assay of aqueous extracts of selected ethno-pharmacologic alternatives used by the Maasai community of Narok, Kenya," *European Journal of Medicinal Plants*, vol. 26, no. 3, pp. 1–11, 2019.
- [39] C. G. Wagate, W. D. Gakuya, M. O. Nanyangi, F. K. Njonge, and J. M. Mbaria, "Antibacterial and cytotoxic activity of Kenyan medicinal plants," *Memorias Do Instituto Oswaldo Cruz*, vol. 103, no. 7, pp. 650–652, 2008.
- [40] J. K. Musau, J. M. Mbaria, and D. W. Gakuya, "The antibacterial activity of some medicinal plants used in Meru central district, Kenya," *The Kenya Veterinarian*, vol. 35, no. 1, pp. 18–24, 2011.
- [41] R. K. Mayaka, M. K. Langat, J. O. Omolo, and P. K Cheplogoi, "Antimicrobial prenylated acetophenones from berries of *Harrisonia abyssinica*," *Planta Medica*, vol. 78, pp. 383–386, 2012.
- [42] E. Madivoli, E. Maina, P. Kairigo et al., "In vitro antioxidant and antimicrobial activity of *Prunus africana* (Hook. f.) Kalkman (bark extracts) and *Harrisonia abyssinica* Oliv. extracts (bark extracts): a comparative study," *Journal of Medicinal Plants for Economic Development*, vol. 2, no. 1, 2018.

- [43] L. C. Ngeny, E. Magiri, C. Mutai, N. Mwikwabe, and C. Bii, "Antimicrobial properties and toxicity of *Hagenia abyssinica* (B.) J. F. Gmel, *Fuerstia Africana* T. C. E. Fries, *Asparagus racemosus* (willd.) and *Ekebergia capensis* sparrm," *African Journal of Pharmacology and Therapeutics*, vol. 2, no. 3, pp. 76–82, 2013.
- [44] E. N. Matu, P. G. Kirira, E. V. M. Kigondu, E. Moindi, and B. Amugune, "Antimicrobial activity of organic total extracts of three Kenyan medicinal plants," *African Journal of Pharmacology and Therapeutics*, vol. 1, no. 1, pp. 14–18, 2012.
- [45] B. K. Amugune, J. W. Mwangi, G. N. Thoithi, and I. O. Kibwage, "In vitro screening of ten selected traditionally used medicinal plants in Vihiga county, Kenya for anti-bacterial and antifungal activity," *International Journal of Medicinal Plants and Natural Products*, vol. 3, no. 2, pp. 37–44, 2017.
- [46] K. R. Cheruiyot, D. Olila, and J. Katereggia, "In-vitro anti-bacterial activity of selected medicinal plants from Longisa region of Bomet district, Kenya," *African Health Sciences*, vol. 9, no. Suppl 1, pp. S42–S46, 2009.
- [47] G. J. Amabeoku and K. Bamuamba, "Evaluation of the effects of *Olea europaea* L. subsp. *africana* (Mill.) P.S. Green (Oleaceae) leaf methanol extract against castor oil-induced diarrhoea in mice," *Journal of Pharmacy and Pharmacology*, vol. 62, no. 3, pp. 368–373, 2010.
- [48] P. G. Kareru, A. N. Gachanja, J. M. Keriko, and G. M. Kenji, "Antimicrobial activity of some medicinal plants used by herbalists in eastern province, Kenya," *African Journal of Traditional, Complementary, and Alternative Medicines*, vol. 5, no. 1, pp. 51–55, 2007.
- [49] D. Kemboi and A. Gitonga, "Antimicrobial activity of *Bridelia micrantha* and *Grewia plagiophylla* leaf extracts," *Journal of Pharmaceutical Research International*, vol. 12, no. 3, pp. 1–7, 2016.
- [50] J. Ya'u, B. A. Chindo, A. H. Yaro, S. E. Okhale, J. A. Anuka, and I. M. Hussaini, "Safety assessment of the standardized extract of *Carissa edulis* root bark in rats," *Journal of Ethnopharmacology*, vol. 147, no. 3, pp. 653–661, 2013.
- [51] H. O. Nyang'au, J. Maingi, and A. Kebira, "The efficacy of some medicinal plants used locally within Transmara west, Narok County, Kenya against selected enterobacteria and candida," *IOSR Journal of Pharmacy and Biological Sciences*, vol. 12, no. 1, pp. 115–122, 2017.
- [52] R. Korir, F. Kimani, J. Gathirwa, M. Wambura, and C. Bii, "In-vitro antimicrobial properties of methanol extracts of three medicinal plants from Kilifi district—Kenya," *African Journal of Health Sciences*, vol. 20, no. 1, pp. 5–10, 2007.
- [53] P. W. Njoroge and S. A. Opiyo, "Some antibacterial and antifungal compounds from root bark of *Rhus natalensis*," *American Journal of Chemistry*, vol. 9, no. 5, pp. 150–158, 2019.
- [54] H. M. Mwangi, W. T. Mabusela, B. M. Abegaz, and O. O. Martin, "Antimicrobial activities of a novel biflavonoid and other constituents from *Rhus natalensis*," *Journal of Medicinal Plants Research*, vol. 7, no. 10, pp. 619–623, 2013.
- [55] L. W. Karani, F. M. Tolo, S. M. Karanja, and C. Khayeka-Wandabwa, "Safety of *Prunus africana* and *Warburgia ugandensis* in asthma treatment," *South African Journal of Botany*, vol. 88, pp. 183–190, 2013.
- [56] F. M. Tolo, G. M. Rukunga, F. W. Muli et al., "The anti-viral effect of *Acacia mellifera*, *Melia azedarach* and *Prunus africana*, extracts against herpes simplex virus type 1 infection in mice," *Journal of Tropical Microbiology and Biotechnology*, vol. 2, no. 1, pp. 3–9, 2006.
- [57] C. Bii, K. Korir, J. Rugutt, and C. Mutai, "The potential use of *Prunus africana* for the control, treatment and management of common fungal and bacterial infections," *Journal of Medicinal Plants Research*, vol. 4, no. 11, pp. 995–998, 2010.
- [58] M. C. Ngule, M. H. Ndiku, and F. Ramesh, "Chemical constituents screening and in vitro antibacterial assessment of *Prunus africana* bark hydromethanolic extract," *Journal of Natural Sciences Research*, vol. 4, no. 16, pp. 85–90, 2014.
- [59] P. G. Mwitari, P. A. Ayeka, J. Ondicho, E. N. Matu, and C. Bii, "Antimicrobial activity and probable mechanisms of action of medicinal plants of Kenya: *Withania somnifera*, *Warburgia ugandensis*, *Prunus africana* and *Plectranthus barbatus*," *PLoS One*, vol. 8, no. 6, Article ID e65619, 2013.
- [60] J. O. Abuto and D. A. Morono, "Interaction effects of site, samples, plant parts and solvent types on antimicrobial activity of the Kenyan populations of *Warburgia ugandensis* (Sprague)," *Journal of Pharmaceutical, Chemical and Biological Sciences*, vol. 6, pp. 1–12, 2018.
- [61] M. Ngule and M. Ndiku, "Antibacterial potency of ethnobotanical plants as alternative remedies to curtail nosocomial infections: a case study of five native plants in Kenya," *British Journal of Pharmaceutical Research*, vol. 6, no. 4, pp. 284–292, 2015.
- [62] M. M. Njire, N. L. M. Budambula, and J. Kiuru, "Antimicrobial activity of *Warburgia ugandensis* against gram negative multi-drug resistant bacteria," *Journal of Agriculture, Science and Technology*, vol. 16, no. 2, 2014.
- [63] H. G. Mikail, "Phytochemical screening, elemental analysis and acute toxicity of aqueous extract of *Allium sativum* L. bulbs in experimental rabbits," *Journal of Medicinal Plants Research*, vol. 4, no. 4, pp. 322–326, 2010.
- [64] O. D. Nyaitondi, R. Wanjau, H. N. Nyambaka, and A. Hassanali, "Anti-bacterial properties and GC-MS analysis of extracts and essential oils of selected plant product," *Biofarmasi Journal of Natural Product Biochemistry*, vol. 16, pp. 44–58, 2018.
- [65] E. E. Makhulu, N. S. Nyaga, S. Wambugu, and G. Areba, "Synergistic antimicrobial activity of crude ethanolic extracts of garlic and neem leaves against bovine mastitis pathogens: an in vitro assay," *International Journal of Basic and Clinical Pharmacology*, vol. 6, no. 9, 2017.
- [66] G. O. Mauti, E. M. Mauti, G. Ouno, and B. Mabeya, "Antibacterial activity of garlic, tulsi, bitter guard and cinnamon extracts against wound pathogens," *Journal of Scientific and Innovative Research*, vol. 4, no. 44, pp. 178–181, 2015.
- [67] S. K. Mbuthia, F. N. Wachira, and R. K. Koech, "In-vitro antimicrobial and synergistic properties of water-soluble green and black tea extracts," *African Journal of Microbiology Research*, vol. 8, no. 14, pp. 1527–1534, 2014.
- [68] Y.-W. Hsu, C.-F. Tsai, W.-K. Chen, C.-F. Huang, and C.-C. Yen, "A subacute toxicity evaluation of green tea (*Camellia sinensis*) extract in mice," *Food and Chemical Toxicology*, vol. 49, no. 10, pp. 2624–2630, 2011.
- [69] J. O. Obwoge, J. K. Kinyua, D. W. Kariuki, and G. N. Magoma, "Phytochemical screening and antimicrobial studies of green, orthodox and black Kenyan tea," *Journal of Agriculture, Science and Technology*, vol. 16, no. 3, 2014.
- [70] P. N. Waithaka, E. M. Gathuru, B. M. Githaiga, and J. K. Tembo, "Synergistic antimicrobial effect of Egerton university cow's urine and neem tree (*Azadirachta indica*) crude extracts on selected infectious human and plant pathogenic microbes," *International Academy of Engineering and Medical Research*, vol. 1, no. 3, 2016.

- [71] J. A. Soule, “*Tagetes minuta*: a potential new herb from South America,” in *New Crops*, J. Janick and J. E. Simon, Eds., pp. 649–654, Wiley, New York, NY, USA, 1993.
- [72] M. F. Musila, S. F. Dossaji, J. M. Nguta, C. W. Lukhoba, and J. M. Munyao, “In vivo antimalarial activity, toxicity and phytochemical screening of selected antimalarial plants,” *Journal of Ethnopharmacology*, vol. 146, no. 2, pp. 557–561, 2013.
- [73] W. Njue, J. K. Kithokoi, J. Mburu, H. Mwangi, and S. Swaleh, “Green sonochemical synthesis of silver nanoparticles using *Adansonia digitata* leaves extract and evaluation of their antibacterial potential,” *European Journal of Advanced Chemistry Research*, vol. 1, no. 2, 2020.
- [74] M. M. Kaigongi, “Antimicrobial activity, toxicity and phytochemical analysis of four medicinal plants traditionally used in Msambweni District, Kenya,” 2014, <https://repository.uonbi.ac.ke/handle/11295/74294?show=full>.
- [75] F. W. Ngari, R. N. Wanjau, E. M. Njagi, and N. K. Gikonyo, “Antimicrobial and phytochemical investigation of herbal suspensions used in management of oral health in Nairobi County, Kenya,” *Journal of Biology, Agriculture and Healthcare*, vol. 4, no. 14, 2014.
- [76] A. O. Abiba, “Antibacterial efficacy and safety of selected Kenyan medicinal plants,” 2013, <https://www.semanticscholar.org/paper/Antibacterial-Efficacy-and-Safety-of-Selected-Abiba/9c596064195df743f118378c37e4de8e0b97d219>.
- [77] I. Mbabazi, P. Wangila, and I. O. K’Owino, “Antimicrobial activity of *Euclea divinorum* hern (ebenaceae) leaves, tender stems, root bark and an herbal toothpaste formulated from its ethanolic root bark extract,” *International Journal of Research and Reports in Dentistry*, vol. 3, pp. 8–16, 2020.
- [78] S. Kumar, C. Rani, and M. Mangal, “A critical review on *Salvadora persica*: an important medicinal plant of arid zone,” *International Journal of Phytomedicine*, vol. 4, pp. 292–303, 2012.
- [79] E. O. Omwenga, P. Okemo, and P. Mbugua, “In vitro antimicrobial and preliminary phytochemical screening of some Samburu anti-diarrheal medicinal plants-Kenya,” *South Asian Journal of Experimental Biology*, vol. 89, 2012.
- [80] F. M. Musila, J. M. Nguta, C. W. Lukhoba, and S. F. Dossaji, “Antibacterial and antifungal activities of 10 Kenyan *Plectranthus* species in the *Coleus* clade,” *Journal of Pharmacy Research*, vol. 11, no. 8, pp. 1003–1015, 2017.
- [81] J. M. Nguta and M. N. Kiraithe, “In vitro antimicrobial activity of aqueous extracts of *Ocimum suave* Willd, *Plectranthus barbatus* andrews and *Zanthoxylum chalybeum* Engl. against selected pathogenic bacteria,” *Biomedical and Biotechnology Research Journal (BBRJ)*, vol. 3, no. 1, pp. 30–34, 2019.
- [82] J. K. Obey, A. von Wright, J. Orjala, J. Kauhanen, and C. Tikkanen-Kaukanen, “Antimicrobial activity of *Croton macrostachyus* stem bark extracts against several human pathogenic bacteria,” *Journal of Pathogens*, vol. 2016, Article ID 1453428, 5 pages, 2016.
- [83] A. Maroyi, “Ethnopharmacological uses, phytochemistry, and pharmacological properties of *Croton macrostachyus* hochst. Ex delile: a comprehensive review,” *Evidence-Based Complementary and Alternative Medicine*, vol. 2017, Article ID 1694671, 17 pages, 2017.
- [84] L. O. Orafidiya, E. O. Agbani, E. O. Iwalewa, K. A. Adelusola, and O. O. Oyedapo, “Studies on the acute and sub-chronic toxicity of the essential oil of *Ocimum gratissimum* L. leaf,” *Phytomedicine*, vol. 11, no. 1, pp. 71–76, 2004.
- [85] L. G. Matasyoh, J. C. Matasyoh, F. N. Wachira, M. G. Kinyua, A. W. Muigai, and T. K. Mukama, “Antimicrobial activity of essential oils of *Ocimum gratissimum* L. From different populations of Kenya,” *African Journal of Traditional, Complementary, and Alternative Medicines*, vol. 5, no. 2, pp. 187–193, 2008.
- [86] J. C. Matasyoh, J. J. Kiplimoa, N. M. Karubiu, and T. P. Hailstorksc, “Chemical composition and antimicrobial activity of the essential oil of *Satureja biflora* (lamiaceae),” *Bulletin of the Chemical Society of Ethiopia*, vol. 21, no. 2, pp. 249–254, 2007.
- [87] P. V. Tan, C. Mezui, G. Enow-Orock, N. Njikam, T. Dimo, and P. Bitolog, “Teratogenic effects, acute and sub chronic toxicity of the leaf aqueous extract of *Ocimum suave* Wild (Lamiaceae) in rats,” *Journal of Ethnopharmacology*, vol. 115, no. 2, pp. 232–237, 2008.
- [88] S. N. Njeru, M. A. Obonyo, S. O. Nyambati, and S. M. Ngari, “Antimicrobial and cytotoxicity properties of the crude extracts and fractions of *Premna resinosa* (Hochst.) Schauer (Compositae): Kenyan traditional medicinal plant,” *BMC Complementary and Alternative Medicine*, vol. 15, no. 1, p. 295, 2015.
- [89] E. Karumi, “Anthelmintic and antimicrobial activity of *Hagenia abyssinica* (bruce) J. F. Gmel (rosaceae),” *East and Central African Journal of Pharmaceutical Sciences*, vol. 16, no. 4, pp. 77–82, 2013.
- [90] S. Njeru, M. Obonyo, S. Nyambati, S. Ngari, R. Mwakubambanya, and H. Mavura, “Antimicrobial and cytotoxicity properties of the organic solvent fractions of *Clerodendrum myricoides* (Hochst.) R. Br. ex Vatke: Kenyan traditional medicinal plant,” *Journal of Intercultural Ethnopharmacology*, vol. 5, no. 3, pp. 226–232, 2016.
- [91] S. M. Auwal, M. K. Atiku, M. W. Alhassan, and S. S. Mohammed, “Phytochemical composition and acute toxicity evaluation of aqueous root bark extract of *Securidaca longepedunculata* (Linn),” *Bayero Journal of Pure and Applied Sciences*, vol. 5, no. 2, pp. 67–72, 2013.
- [92] E. N. Wekesa, M. Ojunga, and Z. N. Otieno-Ayayo, “(Polygalaceae) against two standard isolates of *Neisseria gonorrhoeae*,” *International Journal of Biochemistry Research and Review*, vol. 29, no. 6, pp. 61–68, 2020.
- [93] M. G. Abukakar, A. N. Ukwuani, and R. A. Shehu, “Phytochemical screening and antibacterial activity of *Tamarindus indica* pulp extract,” *Asian Journal of Biochemistry*, vol. 3, no. 2, pp. 134–138, 2008.
- [94] MO. Nyaberi, “Development and phytochemical characterization of a herbal preservative from *Tamarindus indica* and *Ziziphus abyssinica* herbs,” 2018, <https://ir.jkuat.ac.ke/handle/123456789/4792>.
- [95] J. K. Bunddotich, “Study of acute toxicity of *Zanthoxylum chalybeum* using mice and brine shrimp experimental models,” 2012, <https://repository.uonbi.ac.ke/handle/11295/6806>.
- [96] B. M. Pour and S. Sasidharan, “In vivo toxicity study of *Lantana camara*,” *Asian Pacific Journal of Tropical Biomedicine*, vol. 1, no. 3, pp. 230–232, 2011.
- [97] R. A. Reddeman, R. Glávits, J. R. Endres et al., “A toxicological evaluation of mango leaf extract (*Mangifera indica*) containing 60% mangiferin,” *Journal of Toxicology*, vol. 2019, Article ID 4763015, 14 pages, 2019.
- [98] J. K. Mutua, S. Imathi, and W. Owino, “Evaluation of the proximate composition, antioxidant potential, and antimicrobial activity of mango seed kernel extracts,” *Food Sciences and Nutrition*, vol. 5, no. 2, pp. 349–357, 2016.

- [99] Z. H. Mbwambo, M. J. Moshi, P. J. Masimba, M. C. Kapingu, and R. S. O. Nondo, "Antimicrobial activity and brine shrimp toxicity of extracts of *Terminalia brownii* roots and stem," *BMC Complementary and Alternative Medicine*, vol. 7, p. 9, 2007.
- [100] J. M. Omale, "Ethnobotany and antimicrobial properties of medicinal plants used in Emuhaya Sub-County, Vihiga County in Western Kenya," Masters thesis, University of Nairobi, Nairobi, Kenya, 2017.
- [101] M. O. Aduol and K. O. Ogila, "Antibacterial activity of dichloromethane, hexane, ethanol and methanol extracts of *Asparagus setaceus* Kunth and *Caesalpinia volkensii* harm," *Asian Journal of Pharmaceutical and Health Sciences*, vol. 2, no. 3, pp. 380–383, 2012.
- [102] M. M. Ndile, W. M. Mbinda, W. M. Mbinda, and M. P. Ngugi, "Caesalpinia volkensii: unexploited natural source of medicine," *The Journal of Phytopharmacology*, vol. 7, no. 3, pp. 288–291, 2018.
- [103] B. Amugune, G. Thoithi, J. Mwangi, L. Omosa, and I. Kibwage, "Antimicrobial activity and bioactive constituents of *Alectra sessiliflora* (vahl) Kuntze methanol extract," *The East and Central African Journal of Pharmaceutical Sciences*, vol. 16, no. 3, pp. 61–68, 2013.
- [104] J. M. Onyancha, A. W. Bibiane, A. M. Gervason, N. A. Lameck, and K. N. Japhet, "The antibacterial, antioxidant and phytochemical composition of *Combretum tanaense* (J. Clark) root extracts," *European Journal of Medicinal Plants*, vol. 23, no. 4, pp. 1–8, 2018.
- [105] M. J. Muema, "Phytochemical and antimicrobial investigation of *Ochna thomasiana* engl. & gilg," 2015, <https://ir-library.ku.ac.ke/handle/123456789/13322>.
- [106] Y.-C. Liang, C.-J. Lin, C.-Y. Yang et al., "Toxicity study of *Bidens pilosa* in animals," *Journal of Traditional and Complementary Medicine*, vol. 10, no. 2, pp. 150–157, 2019.
- [107] J. Cherotich, "Antimicrobial activity, acute toxicity and phytochemical composition of four medicinal plants traditionally used in sotik sub-county, Kenya," 2015, <https://erepository.uonbi.ac.ke/handle/11295/90127?show=full>.
- [108] S. Onoja, C. Ukwueze, M. Ezeja, and N. Udeh, "Antinociceptive and antioxidant effects of hydromethanolic extract of *Bridelia micrantha* stem bark," *Journal of Experimental and Integrative Medicine*, vol. 4, no. 4, p. 273, 2014.
- [109] A. W. Wanyama, J. A. Orwa, P. K. Njenga, and B. N. Irungu, "Evaluation of cytotoxicity, antimicrobial activities and minerals composition of *Vigna subterranea* (L.) verd. (*Bambara groundnut*) extracts," *African Journal of Health Sciences*, vol. 30, no. 2, 2017.
- [110] S. A. Oyebadejo and I. P. Solomon, "Acute and sub-acute toxicity study of *Citrus limon* (L) juice in Sprague dawley rats," *East African Scholars Journal of Biotechnology and Genetics*, vol. 1, no. 2, 2019.
- [111] M. M. Namadina, A. Nuhu, S. Yahuza et al., "Phytochemical screening, physicochemical properties and acute toxicity study of the leaves of *Ziziphus abyssinica* hochst. EX A. RICH. (RHAMNACEAE)," *Chemistry*, vol. 3, no. 3, pp. 102–108, 2019.
- [112] J. Naidu, R. Ismail, and S. Sasidharan, "Acute oral toxicity and brine shrimp lethality of methanol extract of *Mentha spicata* L (lamiaceae)," *Tropical Journal of Pharmaceutical Research*, vol. 13, no. 1, pp. 101–107, 2014.
- [113] B. M. Githaiga, E. M. Gathuru, E. M. Gathuru, P. N. Waithaka, and L. W. Kiarie, "Determination of antibacterial activity of essential oils from mint (*Mentha spicata*) leaves on selected pathogenic bacteria," *Journal of Drugs and Pharmaceutical Science*, vol. 2, no. 2, pp. 8–14, 2018.
- [114] S. N. Njeru, C. M. Josphat, G. M. Charles, and M. M. Charles, "Antibacterial activity of methanol root extract of *Indigofera lupatana* Baker F," *Eastern Journal of Medicine*, vol. 17, pp. 11–16, 2012.
- [115] R. N. Husna, A. Noriham, H. Nooraain, A. H. Azizah, and O. F. Amna, "Acute oral toxicity effects of *Momordica charantia* in Sprague dawley rats," *International Journal of Bioscience, Biochemistry and Bioinformatics*, vol. 3, no. 4, 2013.
- [116] D. S. B. Ongarora, "Phytochemical investigation and antimicrobial activity of *Blighia unijugata* bak (sapindaceae)," 2009, <https://erepository.uonbi.ac.ke/handle/11295/24499>.
- [117] N. M. Bléyéré, K. F. N'dia, K. L. Kouakou, K. J. Abo, and E. E. Ehilé, "Acute toxicity in mice and effects of a butanol extract from the leaves of *Blighia unijugata* bak. (Sapindaceae) on electrocardiogram of rabbits," *Scholars Academic Journal of Pharmacy*, vol. 2, no. 6, pp. 429–435, 2013.
- [118] M. Ondicho, J. Mutai, C. Rukunga, G. Oketch, and C. Bii, "Antimicrobial activity of the root, stem bark and seed extracts of *Moringa Oleifera* Lam." in *Proceedings of the 5th World Congress on Chemical, Biological and Radiological Terrorism*, Dubrovnik, Croatia, 2009.
- [119] G. A. Asare, B. Gyan, K. Bugyei et al., "Toxicity potentials of the nutraceutical *Moringa oleifera* at supra-supplementation levels," *Journal of Ethnopharmacology*, vol. 139, no. 1, pp. 265–272, 2012.
- [120] T. Chemweno, M. Lizzy, K. Richard, M. Angela, and B. Christine, "Antimicrobial activity and safety of *Maesa lanceolata* for the treatment and management of selected bacterial pathogens," *Journal of Advances in Microbiology*, vol. 8, no. 3, pp. 1–8, 2018.
- [121] K. R. Wamuyu, A. K. Machocho, and A. W. Wafula, "Antimicrobial and phytochemical screening of *Lannea schweinfurthii* (Engl.) Engl," *Biotechnologi*, vol. 17, pp. 1–13, 2020.
- [122] S. Dutta and D. Bhattacharyya, "Enzymatic, antimicrobial and toxicity studies of the aqueous extract of *Ananas comosus* (pineapple) crown leaf," *Journal of Ethnopharmacology*, vol. 150, no. 2, pp. 451–457, 2013.
- [123] M. Precious, K. John, and M. Naomi, *In Vitro Antimicrobial Activity of Nano-Encapsulated Bromelain against Bacteria Isolated from Milk of Dairy Goats with Sub-Clinical Mastitis in Thika East Sub-county, Kenya*, 2020.
- [124] D. M. Chalo, C. Lukhoba, D. S. Fidahussein, and J. M. Nguta, "Antimicrobial activity, toxicity and phytochemical screening of selected medicinal plants of Losho, Narok county, Kenya," *Biofarmasi (Rumphius Journal of Natural Product Biochemistry)*, vol. 15, pp. 29–43, 2017.
- [125] M. Ndikau, N. M. Noah, D. M. Andala, and E. Masika, "Green synthesis and characterization of silver nanoparticles using *Citrullus lanatus* fruit rind extract," *International Journal of Analytical Chemistry*, vol. 2017, Article ID 8108504, 9 pages, 2017.
- [126] S. Belemkar and P. N. Shendge, "Toxicity profiling of the ethanolic extract of *Citrullus lanatus* seed in rats: behavioral, biochemical and histopathological aspects," *Bioscience Reports*, vol. 41, no. 1, Article ID BSR20202345, 2021.
- [127] K. W. Ping, I. Darah, Y. Chen, S. Sreeramanan, and S. Sasidharan, "Acute and subchronic toxicity study of *Euphorbia hirta* L. methanol extract in rats," *BioMed Research International*, vol. 2013, Article ID 182064, 14 pages, 2013.

- [128] LP. Kibet, "Ethnobotanical Study, toxicity and phytochemical screening of selected medicinal plants of Tinderet district, Nandi county, Kenya," 2013, <https://erepository.uonbi.ac.ke/handle/11295/61411>.
- [129] M. Chagnon, "Inventaire pharmacologique general des plantes medicinales rwandaises," *Journal of Ethnopharmacology*, vol. 12, no. 3, pp. 239–251, 1984.
- [130] N. Kimutai, E. W. Njenga, P. Jeruto et al., "Antimicrobial activity and cytotoxicity of selected medicinal plants found in Nandi county, Kenya," *African Journal of Pharmacology and Therapeutics*, vol. 4, no. 3, pp. 86–91, 2015.
- [131] P. Kamdem Boniface, M. Singh, A. Kumar Maurya, and A. Pal, "Acute and sub-chronic toxicity of HPLC finger-printed extract of *Conyza sumatrensis* (Retz.) E.H. Walker in rodents," *Journal of Ethnopharmacology*, vol. 149, no. 3, pp. 833–837, 2013.
- [132] A. O. Maima, S. N. Ndwigah, G. N. Thoithi, F. N. Kamau, and O. I. Kibwage, "Antimicrobial properties of some medicinal plants of the Luo community of Kenya," *African Journal of Pharmacology and Therapeutics*, vol. 3, no. 4, pp. 112–115, 2014.
- [133] B. Olela, J. Mbaria, T. Wachira, and G. Moriasi, "Acute oral toxicity and anti-inflammatory and analgesic effects of aqueous and methanolic stem bark extracts of *Piliostigma thonningii* (schumach.)," *Evidence-Based Complementary and Alternative Medicine*, vol. 2020, Article ID 5651390, 10 pages, 2020.
- [134] W. Chitopo, I. Muchachaa, and R. Mangoyi, "Evaluation of the antimicrobial activity of *Erythrina abyssinica* leaf extract," *Journal of Microbial and Biochemical Technology*, vol. 11, no. 2, p. 413, 2019.
- [135] J. W. Mwangi, G. N. Thoithi, I. O. Kibwage et al., "Essential oil of *Rynchosia minima* DC. from Kenya: composition and antibacterial properties," *Journal of Essential Oil Research*, vol. 17, no. 2, pp. 230–231, 2005.
- [136] P. W. Mayeku, A. Hassanali, B. T. Kiremire, J. O. Odalo, and C. Hertweck, "Anti-bacterial activities and phytochemical screening of extracts of different parts of *Thalictrum rhynchocarpum*," *African Journal of Traditional, Complementary, and Alternative Medicines*, vol. 10, no. 5, pp. 341–344, 2013.
- [137] E. K. Ruttoh, P. K. Tarus, C. C. Bii, A. K. Machocho, L. K. Karimie, and P. O. Okemo, "Antibacterial activity of *Tabernaemontana stapfiana* britten (apocynaceae) extracts," *African Journal of Traditional, Complementary, and Alternative Medicines*, vol. 6, no. 2, pp. 186–194, 2009.
- [138] A. Mutuku, L. Mwamburi, L. Keter et al., "Evaluation of the antimicrobial activity and safety of *Rhus vulgaris* (Anacardiaceae) extracts," *BMC Complementary Medicine and Therapies*, vol. 20, no. 1, p. 272, 2020.
- [139] M. M. Kaigongi, C. W. Lukhoba, S. Yaouba, N. P. Makunga, J. Githomi, and A. Yenesew, "In vitro antimicrobial and antiproliferative activities of the root bark extract and isolated chemical constituents of *Zanthoxylum paracanthum* Kokwaro (Rutaceae)," *Plants*, vol. 9, no. 7, p. 920, 2020.
- [140] P. Thakurdesai, P. Deshpande, and V. Mohan, "Preclinical safety assessment of standardized extract of *Centella asiatica* (L.) urban leaves," *Toxicology International*, vol. 22, no. 1, pp. 10–20, 2015.
- [141] P. Waithaka, G. Eliud, and B. Githaiga, "Antimicrobial properties of *Aloe vera*, *Aloe volkensii* and *Aloe secundiflora* from egerton university," *Acta Scientific Microbiology*, vol. 1, pp. 6–10, 2018.
- [142] F. G. Mugweru and D. W Nyamai, "In vivo safety of aqueous extracts of *Maytemus putterlickoides*, *Senna spectabilis* and *Olinia usambarensis* on mice models," *Journal of Clinical Toxicology*, vol. 6, no. 3, 2016.
- [143] R. S. Odhiambo, G. P. Kareru, L. H. Kutima et al., "Antibacterial activity of ethanolic extracts of *Prosopis juliflora* against gram-negative bacteria," *European Journal of Experimental Biology*, vol. 5, no. 11, pp. 43–46, 2015.
- [144] V. D. A. da Silva, A. M. M. da Silva, J. H. C. E Silva, and S. L. Costa, "Neurotoxicity of *Prosopis juliflora*: from natural poisoning to mechanism of action of its piperidine alkaloids," *Neurotoxicity Research*, vol. 34, no. 4, pp. 878–888, 2018.
- [145] H. T. Manekeng, A. T. Mbaveng, S. A. Ntyam Mendo, A. D. Agokeng, and V. Kuete, "Evaluation of acute and subacute toxicities of *Psidium guajava* methanolic bark extract: a botanical with *in vitro* antiproliferative potential," *Evidence-Based Complementary and Alternative Medicine*, vol. 2019, Article ID 8306986, 13 pages, 2019.
- [146] D. Yeo, B. N. Djyh, N. J. David, and R. Bouagnon, "Acute and subacute toxic study of aqueous leaf extract of *Combretum molle*," *Tropical Journal of Pharmaceutical Research*, vol. 11, no. 2, pp. 217–223, 2012.
- [147] G. Ibrahim, E. M. Abdurahman, H. Ibrahim, N. D. G. Ibrahim, and M. G. Magaji, "Toxicity and analgesic effects of *Vernonia amygdalina* Del. (Asteraceae) leaf extract on mice," *International Journal of Advanced Pharmaceutical and Biological Sciences*, vol. 1, no. 1, pp. 1–4, 2011.
- [148] S. W. Muhindi, C. M. Ngule, and F. Ramesh, "Phytochemical and antibacterial potential of *Vernonia adoensis* stem bark to curb cariogenic microorganisms," *American Journal of Phytomedicine and Clinical Therapeutics*, vol. 4, no. 01, pp. 019–027, 2016.
- [149] R. O. Okindo, "Study of antimicrobial, analgesic and toxic properties of *Vernonia hymenolepis* (A. Rich)," 2014, <https://erepository.uonbi.ac.ke/handle/11295/73596?show=full>.
- [150] I. Sandiga, C. N. Chacha, and M. P. Kanunah, *Traditional Medicine in Africa: The Existence and Use of Traditional Medicine in Kenya*, East African Educational Publishers Ltd, Nairobi, Kenya, 1995.
- [151] G. N. Njoroge and J. W. Kibunga, "Herbal medicine acceptance, sources and utilization for diarrhoea management in a cosmopolitan urban area (Thika, Kenya)," *African Journal of Ecology*, vol. 45, no. s1, pp. 65–70, 2007.
- [152] P. B. James, J. Wardle, A. Steel, and J. Adams, "Traditional, complementary and alternative medicine use in Sub-Saharan Africa: a systematic review," *BMJ Global Health*, vol. 3, no. 5, Article ID e000895, 2018.
- [153] N. Maina, J. M. Kagira, O. Achila, S. M. Karanja, and M. Ngotho, "Herbal medicines in Kenya: a review of the toxicity and quality control issues," *African Journal of Health Sciences*, vol. 24, no. 1, 2013.
- [154] P. G. Kareru, J. M. Keriko, G. M. Kenji, G. T. Thiong'o, A. N. Gachanja, and H. N. Mukiria, "Antimicrobial activities of skincare preparations from plant extracts," *African Journal of Traditional, Complementary, and Alternative Medicines*, vol. 7, no. 3, pp. 214–218, 2010.
- [155] M. Onyambu, "Identification and characterization of microbial contaminants of herbal medicines in Kenya," Research thesis, University of Nairobi, Nairobi, Kenya, 2011.
- [156] N. M. Sivasankaran and J. R. Selwin, "Synthesis and characterization of Co (II), Ni (II), Cu (II), and Zinc (II) complexes of tridentate Schiiff base derived from vanillin and DL- $\alpha$ -aminobutyric acid," *Spectrochimica Acta*, vol. 70, pp. 749–753, 2008.

- [157] J. A. Odhiambo, G. M. Siboe, C. W. Lukhoba, and S. F. Dossaji, "Antifungal activity of crude extracts of selected medicinal plants used in combinations in Lake Victoria Basin, Kenya," *Plant Products Research Journal*, vol. 13, 2011.
- [158] T. P. Tiwari, S. K. Bharti, H. D. Kaur, R. P. Dikshit, and G. S. Hoondal, "Synergistic antimicrobial activity of tea and antibiotics," *Indian Journal of Medical Research*, vol. 122, pp. 80–84, 2005.
- [159] S. Faizi, N. R. Mughal, R. A. Khan et al., "Evaluation of the antimicrobial property of *Polyalthia longifolia* var. pendula: isolation of a lactone as the active antibacterial agent from the ethanol extract of the stem," *Phytotherapy Research*, vol. 17, no. 10, pp. 1177–1181, 2003.
- [160] A. O. T. Ashafa, L. O. Orekoya, and M. T. Yakubu, "Toxicity profile of ethanolic extract of *Azadirachta indica* stem bark in male Wistar rats," *Asian Pacific Journal of Tropical Biomedicine*, vol. 2, pp. 811–817, 2012.
- [161] R. C. A. Dubey, *Textbook of Biotechnology*, R. Najar, Ed., S. Chand and Company Ltd., New Delhi, India, 2004.