# ACQUISITION OF ANTIGENS BY AIRWAY DENDRITIC CELLS. DO WE KNOW ENOUGH?

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#### ABSTRACT

The respiratory system is endowed with a number of structural and functional barriers that protect it against harmful and innocuous material from taking advantage of its vast surface area to gain access into the organism. These barriers include; 1) the surfactant system 2) a highly efficient mucociliary escalator system 3) a population of highly phagocytic macrophages and 4) an epithelium endowed with tight junctions. However, despite these barriers, pulmonary immune responses are easily generated by introduction of antigens into the airways. These responses are thought to be mediated via dendritic cells, which are located in the basal aspect of the epithelium, and the most potent antigen presenting cells in the lung. Although there is substantial information on the nature of interaction between dendritic cell and particles from *in vitro* experiments, there is little information on how the particles pass the epithelial barrier to reach the immunocompetent cells. An understanding of how these particles pass the epithelial barrier to reach the immunocompetent cells is important in the development of mucosal vaccines. Insights into how this may happen are discussed.

Key words: Immune Cells, Respiratory Tract

### INTRODUCTION

The surface epithelium of the respiratory tract is the most extensive surface that interfaces directly between man and his environment. In an average human weighing about 74 Kg, it may comprise up to 143 m<sup>2</sup> (Gehr *et al.*, 1978) in surface area. This surface is exposed to a large load of particulate matter that is dependent on the level of environmental pollution, inhalability of the pollutant and the breathing habit of the individual (Stahlhofen et al., 1994; Roth et al., 1997). It has been estimated that the human airways may be exposed to as much as 7 Kg of pollutants per year (Phalen 1984) although this may vary depending on the amount of particulate matter suspended in the inhaled air. To counter the danger posed by these particles, the lung is endowed with several structural and functional barriers that constitute its innate immunity against harmful and innocuous particles from reaching the rest of the body organs. These barriers include the following 1) the surfactant system (Schürch et al., 1990; Gehr et al., 1990), 2) a highly efficient mucociliary escalator system (Kilburn 3) a population of highly 1968), phagocytic macrophages (Geiser et al., 1990; Crystal 1991), and 4) a continuum of epithelium endowed with tight junctions (Breeze and Wheeldon 1977; Barry 1987). Nevertheless, numerous epidemiological studies have shown that particulate air pollution is associated with increased morbidity and mortality (Brunekreef 1997; Pope et al., 2009 Pope et al., 2014). Further, immune response related diseases are common in the lung, which suggests that breach of this barrier occurs. One factor that has been said to predispose to breaching of the barrier is particle-overload (Oberdöster et al., 1994). The phenomena of particle overload suggest that the lung barrier

system can only handle up to a given maximum particulate volume beyond which the particles will inevitably get into contact with the epithelium and become interstitialised (Marrow 1988; Oberdöster et al., 1994) where they may reach the dendritic cells. Furthermore, twenty years ago, Gehr and Schürch(1992) reported that upon inhalation, small particles are forcefully displaced by the surfactant lining film towards the airway epithelium forming a depression on the epithelial The displacement enables the cells. to interact with airwav particles macrophages and perhaps dendritic cells (Gehr et al., 1996) thereby determining their fate.

Dendritic cells are recognized as the main antigen presenting cells in the lung (Holt et al., 1990; Nicod 1997) and the nature of how they acquire antigens is a prerequisite in understanding the consequences of particle-lung interaction. Of particular importance is how the antigens can reach DC, which are the most competent antigen presenting cells in the lung (Holt et al., 1990; Nicod, 1997). The location of DCon the basal surface of the epithelium and lack of cells such as M cells in the airway epithelium suggests that there may be active and yet unresolved mechanisms of antigen delivery to the DC in the airways. Are the antigens phagocytosed and delivered byairway macrophages? Do thev penetrate through the airway epithelial cells, or do the antigens pass through the tight junction. Is there a possibility that the DC collect the particles by pushing cytoplasmic processes beyond the tight junctions into the airway lumen (Rescigno *et al.*, 2001; Vermaelen *et al*, 2001; Takano et al., 2005)? How does this antigen delivery change during epithelial injury and repair, which is a common feature of inflammatory airway disease, like asthma (Gaurav and Agrawal 2013)? These questions need urgent answers.

### Dendritic cells

Dendritic cells (DC) are professional antigen presenting cells that play a critical role in generating primary and secondary immune responses against specific antigens (Steinman, 1991). Priming of naive T lymphocytes, proliferation and functional differentiation antigenof specific T cells as well as development of antigen specific T cell tolerance depends on an appropriate T cell - dendritic cell interaction (Banchereau et al., 2000). The high capacity of DC to stimulate T lymphocytes is attributed to (1) a high DC level of adhesion molecules that may favour Т cell receptor engagement (Steinman, 1991), (2) high expression levels of co-stimulatory molecules on the DC that facilitate T cell activation (Katayama et al., 1997; Quaratino et al., 2000) as well as (3) expression of high level of major histocompatibility complex molecules (Inaba et al., 1994; Cella et al., 1997).

DC were first isolated from murine spleen suspension and defined on morphological grounds (Steinman and Cohn, 1973). They were shown to display a distinctive dendritic morphology (Figure1). Subsequently, the DC were defined by functional criteria, notably the lack of endocytic activity invitro and in vivo, the inability to retain antigens or immune complexes on their surface and a low labelling index in vitro (1.5–2.5%) following administration of [<sup>3</sup>H] thymidine (Steinman Cohn, and 1974). Nonphagocytic DC were later isolated from rat peripheral lymph (Pugh et al., 1983) and human lung parenchyma (Nicod et al., 1987). The lack of endocytic activity was a paradox since antigen uptake and processing is expected to precede antigen presentation (Steinman and Swanson 1995).

### Life cycle of dendritic cells.

The origin and ontogeny of DC have been reviewed in detail (Hart, 1997; Granucci and Zanoni 2009). Evidence of bone marrow origin of dendritic cells was demonstrated from irradiation reconstitution experiments following bone marrow transplantation (Katz et al., 1979)(Figure 2). Furthermore, perturbation of the respiratory tract mucosa is knownto induce mobilization of respiratory epithelium. DC in the Inhalation of viruses, bacterial and soluble protein antigens result in recruitment of DC into the respiratory mucosa followed by DC migration to the lymph node (Mc Williamset al., 1996). Subsequently, the migratory immature dendriticcells undergo maturation and become professional antigen presenting cells (Banchereau and Steinman, 1998). Maturation changes DC in many ways that explain their potent antigen presenting capacity to naive T cells. These changes include: (1) a high lymphocytes expression of Т COstimulatory molecules such as B7-2 (Inaba et al., 1994), (2) production of IL-12 (Cella et al., 1996), (3) redistribution of MHC class II molecules from nonlysosomal vesicles to the cell surface (Pierra et al., 1997, Turley et al., 2000), (4) an upregulation of CCR7 that guide migration of DC into the T cell areas (Forster et al., 1999), (5) expression of DC survival molecule, TRANCE-R (Wong et al., 1997), (6) a high expression of adhesion molecules such as ICAM-1, ICAM-3 and LFA-3 (Banchereau and Steinman, 1998) and (7) downregulation of dextran uptake (Cochand et al., 1999). In addition, human DC havebeen shown to reduce their phagocytic capacity on maturation (Kiama et al., 2001).

### Dendritic cells in the epithelial tissue.

An interdigitating network of DC has been described in the intestinal and respiratory tract of rat and humans (Holt *et al.,* 1988; Pavli *et al.,* 1993). Studies from P. Holt's laboratory (Schon-Hegrad *et al.,* 1991; Holt *et al.,* 1994) and other laboratories (Nicod *et al.,* 1987; Gong *et al.,* 1992) have identified two main populations of DC in the lung that differ in location, phenotype and turnover. One population which exhibits a high turnover (2 to 3 days), and a more immature phenotype residing in the conducting airways just below the epithelium (Sertl *et al.,* 1986; Holt *et al.,* 1988) and the second population, that is fairly sparse, exhibiting a more advanced stage of differentiation, a slower turnover (7 days) located in the alveolar septa (Holt *et al.,* 1994).

Densities of DC in the rat are highest in the upper airways (600-800/mm<sup>2</sup>) and decrease with progression down the respiratory tree, reaching 75/mm<sup>2</sup> in the small airways of the peripheral lung (Schon-Hegrad *et al.,* 1991). The airway mucosal DC population is capable of up regulation in response to both acute and chronic inflammation (McWilliamset al., 1994). Targeting these cell population with vaccines presents high potential of inducing strong mucosal and systemic immunity. However, a good understanding of how the potential vectors reach the DC is a prerequisite in achieving desirable immunogenic responses.

## Function of Dendritic cells.

DC are professional antigen presenting cells (Steinman, 1991 van Spriel and de 2014). They Jong phagocytose, processand present immunogenic epitopes in the context of MHC class I and/or class molecules for recognition Π by T lymphocytes (Hart, 1997; Alberts 1998). Besides presenting the antigens, the DC endowed with an are arrav of costimulatory molecules whose interaction with complementary molecules on the T lymphocytes ensures optimal Т cell activation (Banchereau and Steinman, 1998). In addition to their antigen presenting function, DC serve a sentinel function. DC survey epithelial surfaces such as the skin, respiratory tract and gastrointestinal tract for agents that may present a threat to health. The sentinel position of the DC stands out after a challenge of the respiratory mucosa with Moraxella catarrhalis. Inhalation of M. catarrhalis organisms is accompanied by an amplification of active DC surveillance in the airways that results in an increase in the traffic of the DC between the airway

epithelium and the regional lymph nodes (McWilliamset al., 1994). **Besides** stimulating T lymphocytes, DC are now known to have effects on B cell growth and immunoglobulin secretion (Banchereau et al., 2000). DC activate and expand T-helper cells, which in turn induce B cell growth and antibody production (Briere et al., 1999). Human DC have been reported to skew isotope switching of CD40-activated naive B cells in presence of IL-10 and TGF- $\beta$  towards IgA secreting cells (Fayette et al., 1997). Immunoglobulin A is the major class of immunoglobulin present in the mucosa of the healthy respiratory tract and is thought to be the most important immunoglobulin for lung defence (Lamm, 1997). This suggests that DC are in control of mucosal immunity, since DC located in the airway epithelia could directly influence the isotype switch of B cells towards IgA. Other functions of the DC include induction of central and peripheral tolerance (Banchereau and Steinman, 1998), control of Th1/Th2 directed immune responses (Rissoan et al., 1999) and linking of innate and adaptive immunity via production of interferon alpha (Palucka and Banchereau, 1999).

Our studies and other reports have demonstrated that DC are efficiently phagocytic for a variety of particles such as, 1) polystyrene particles (Kiama et al, 2001; 2) puff ball spores, 3) biodegradable microspheres made of poly(lactid-coglycolid) acid (PLGA) (Walter et al., 2001), and 4) Salmonella typhimurium (Kiama et al., 2006; Dreher et al., 2001). The uptake of Salmonella and PLGA particles by DC is important because of their potential use in drug delivery and as vectors in delivery of DNA vaccines. This is because, Salmonella not only invade DC very efficiently but also induces the expression of costimulatory signals for T cell (Dreher et al. 2001 ) and hence can serve as a carrier of the vaccine. Salmonella was also found to induce formation of microvesicles after infection of DC (Kiama et al., 2006). The significance of these microvesicles

(Szakal *et al.*, 1988) in amplification of immunity on salmonella based vaccine remains to be explored (Obregon *et al.*, 2006). Whereas much is known on the interaction of particles with DC and with alveolar macrophages there is no sufficient data on how the antigens reach the DC (Vermaelen and Pauwels, 2005; Takano *et al.*, 2005).

## Surfactant barrier

The primary role of pulmonary surfactant is to reduce surface tension forces in the lung and to stabilize pulmonary alveoli (Schürch et al., 1976). However, it is now generally well recognized that surfactant may also serve the role of a barrier to inhaled antigens reaching the epithelium (Pison et al., 1994) and the dendritic cells. It does this through several mechanisms that include: 1) enhancing mucociliary clearance (De Sanctis et al., 1994), 2) improving phagocytosis of particles by alveolar macrophages (Pison et al., 1994), and 3) displacing small particles to the vicinity of macrophages and epithelial cells (Gehr et al., 1990; Schürch et al., 1990; Geiser et al., 2000). In view of the concomitant observation of surfactant deficiencies and poor airways clearance in lung diseases such as cystic fibrosis and asthma (Griese et al., 1997; Meyer et al., unravelling the role 2000), which surfactant may play as a barrier of particles to reach the epithelium and subsequently the dendritic cells would provide useful information on how to treat such diseases. Furthermore, emerging evidence indicate that surfactant proteins A and D modulates dendritic cell function and helper T cell polarization (Nayak et al, 2012, Schleh et al 2012)

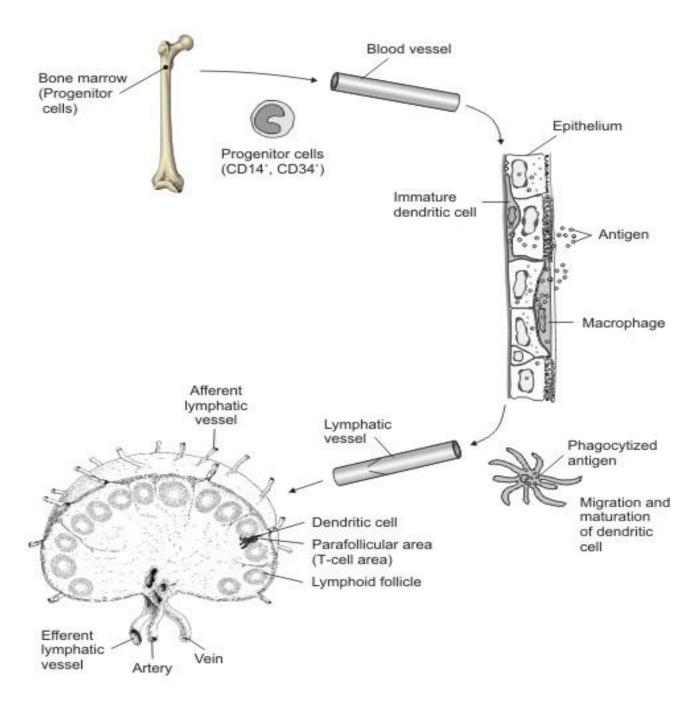
## Mucociliary barrier

The efficiency of the mucociliary barrier depends on among other factors, the morphological integrity of the cilia structure and mucus components (Werner *et al.,* 1996). Impairment of the mucociliary barrier, which is common in diseases such as cystic fibrosis and

asthma, predisposes to bacterial colonization of the airways (Werner *et al.,* 1996). Although impaired clearance is known to result in prolonged retention of deposited particles, its contribution to antigens gaining access to dendritic cells through the epithelium has not been

determined. Delayed clearance of particles deposited in the airways would predispose to a breach of airway epithelial barrier highlighting the risk of particles gaining access to dendritic cells.

# Migration of dendritic cells



**Figure 2:** A schematic illustration of the life cycle of Dendritic cell. The Dc precursors migrate from the bone marrow to the epithelium where they differentiate to immature

dendritic cells. The immature dendritic cells take up antigen and migrate to the regional lymph node in the process maturing to the professional antigen presenting cells.



**Figure 1:** An Electron micrograph of a dendritic cell displaying the long dendritic processes that are typical for this cell type.

### Macrophage Barrier

Airway macrophages are highly phagocytic and are rapidly recruited to the locations where particles have been deposited al., (Geiser et 1994). Macrophage overloading though, may interfere with the ability of these cells to phagocytise 1994). (Oberdoster et al., The phenomenon of particle-overload assumes that macrophages can only engulf up to a given maximum volume of particles (Morrow 1988; Oberdöster et al., 1994). Morrow (1988) suggested that alveolar macrophage functions begin to be impaired when on average 6 % of its volume is filled by phagocytosed particles. The capacity of pulmonary DC to serve as antigen presenting cells for heat killed Listeria after in vivo challenge is observed only when the dose of heat-killed Listeria exceeded 10<sup>9</sup> organisms per rat (MacLean et al., 1996). Moreover, elimination by alveolar macrophages in vivo is associated with increased pulmonary immune response to intra-tracheal administered heat-killed Listeria (Kradinet al., 1999) implying that macrophages serve a protective role. This suggests that particle overload plays a role in the breaching of the epithelial barrier.

### **Epithelial barrier**

A continuous layer of epithelial cells joined by tight junctions (Breeze and Wheeldon 1977) lines the pulmonary airways. The tight junctions completely prevent the diffusion of macromolecules through the intercellular spaces across the epithelium (Balda and Matter, 1998). Transport of the particles to the DC cells presupposes their passage across the epithelium, although the route they take has not been determined. Vermaelen and colleagues (2001)reported that fluorescein conjugated isothiocyanate (FITC)macromolecules are transported to the tracheal lymph nodes by airway DC after an intratracheal instillation. However, the mechanism in which the macromolecules passed through the epithelium to reach the DC was not provided. Takano and colleagues (Takano et al., 2005) showed that dendritic cells easily access antigens beyond epithelial tight junctions in human nasal mucosa, but of allergic rhinitis only.

Another *in vitro* model using mouse tracheal epithelial cells and mouse bone marrow dendritic cells showed impaired migration of metalloproteinase-9-deficient dendritic cells through tracheal epithelial tight junctions (Ichiyasu *et al.*, 2004). Furthermore, there is evidence that dendritic cells play an important role in the pathogenesis of allergic asthma, and there is an increased number of dendritic cells in the airway mucosa of patients with chronic obstructive pulmonary disease (Vermaelen and Pauwels, 2005).

In conclusion, once particles have evaded thesurfactant, mucociliary, macrophages and the epithelial barrier they come into contact with the professional antigen presenting cells, the dendritic cells. One or several mechanisms may be involved in breaching each barrier which could determine the outcome of the dendritic cell-particle interaction. Thus, concerted studies ought to be carried out to unravel how each barrier is breached perhaps identify whether there is a critical barrier and thus answer the questions. Are all the barriers equally important or is there one that is more redundant?. Or even the question. Do we know enough about the barriers?

### REFERENCES

- 1. Alberts ML, Pearce S F, Francisco L M, Sauter B, Roy P, Silverstein R L, Bhardwaj N. 1998. Immature dendritic cells phagocytose apoptotic cells via alphavbeta5 and CD36, and cross-present antigens to cytotoxic T lymphocytes. *J. Exp. Med.* 188:1359-1368.
- 2. Balda MS, Matter K. 1998. Tight junctions. J. Cell Sci. 111: 541-547.
- 3. Banchereau J, Steinman RM. 1998. Dendritic cells and the control of immunity. *Nature* 392: 245-252.
- 4. Banchereau J, Briere F, Caux C, Davoust J, Lebecque S, Liu Y J, Pulendran B, K. Palucka K.2000. Immunobiology of dendritic cells. *Annu. Rev. Immunol.* 18:767-811.
- 5. Breeze RG, Wheeldon EB. 1977. The cells of the pulmonary airways. Am. Rev. Respir. Dis. 116: 705-777.
- Briere F, Caux C, Dubois B, Fayette J, Vandenabeele S, Banchereau J. 1999. Interactions between dendritic cells and B lymphocytes. *In* M.T. Lotze and A.W. Thomson, eds. *Dendritic Cells. Biology and Clinical applications*. Academic press, San Diego, pp. 269-280.
- 7. Brunekreef B. 1997. Air pollution and life expectancy: is there a relation?Occupational and Environmental Medicine 54:781-784
- 8. Cella M, Engering A, Pinet V, Pieters J, Lanzavecchia A. 1997. Inflammatory stimuli induce accumulation of MHC class II complexes on dendritic cells. *Nature* 388:782-787.
- 9. Cella M, Scheidegger D, Palmer-Lehmann K, Lane P, Lanzavecchia A, Alber G.1996. Ligation of CD40 on dendritic cells triggers production of high levels of interleukin-12 and enhances T cell stimulatory capacity: T-T help via APC activation. *J. Exp. Med.* 184:747-52.
- 10. Cochand L, Isler P, Songeon F, Nicod LP.1999. Human dendritic cells have an immature phenotype with efficient mannose receptors. *Am J. Respir. Cell Mol. Biol.* 21: 547-554.
- 11. Crystal RG. 1991. Alveolar macrophages. In "The Lung: Scientific Foundation". (G. Crystal G, West JB, Barnes PJ, Cherniack NS, Weibel ER eds). Pp 527-538. Raven Press: New York.
- 12. De Sanctis GT, Tomkiewicz RP, Rubin BK, Schürch S, King M. 1994. Exogenous surfactant enhances mucociliary clearance in the anaesthetized dog. Eur. Respir. J. 7:1616-1621.

- 13. Dreher D, Cochand L, Kok M, Kiama S G, Gehr P, Pechére J C, Nicod LP. 2001. Genetic background of attenuated *Salmonella typhimurium* has profound influence on infection and cytokine patterns in human dendritic cells. *Journal of Leukocyte Biology*69:583-589.
- 14. Fayette J, Dubois B, Vandenabeele S, Bridon J M, Vanbervliet B, Durand I, Banchereau J, Caux C, Briere F. 1997. Human dendritic cells skew isotype switching of CD40-activated naive B cells towards IgA1 and IgA2. *J. Exp. Med.* 185:1909-18.
- 15. Forster R, Schubel A, Breitfeld D, Kremmer E, Renner-Muller I, Wolf E, Lipp M.1999. CCR7 coordinates the primary immune response by establishing functional microenvironments in secondary lymphoid organs. *Cell* 99:23-33.
- 16. Gaurav and D. K. Agrawal. 2013. Clinical view on the importance of dendritic cells in asthma. Expert. Rev. Clin. Immunol. 10: 899-919
- 17. Gehr P, Bachofen M, Weibel ER. 1978. The normal human lung: Ultrastructur and morphometric estimation of diffusion capacity. Respir Physiol 32, 121-140.
- 18. Gehr P, Schürch S. 1992. Surface forces displace particles deposited in airways toward the epithelium. News in Physiological Sciences 7:1-5.
- 19. Gehr P, Green FHY, Geiser M, Im Hof V, Lee MM, Schürch S.1996. Airway surfactant, a primary defense barrier: Mechanical and immunological aspects. J. Aerosol Med. 9: 163-181.
- 20. Gehr P, Schürch S, Berthiaume Y, Im Hof V, Geiser M.1990. Particle retention in airways by surfactant. J. Aerosol Med. 3: 27-43.
- 21. Geiser M, Baumann M, Cruz-Orive LM, Im Hof V, Gehr P.1990. Assessment of particle retention and clearance in the intrapulmonary conducting airways of hamster lungs with the fractionator. J. Microsc. 160: 75-88.
- 22. Geiser M, Baumann M, Cruz-Orive L M, Im Hof V, Gehr P.1994. The effect of particle inhalation on macrophage number and phagocytic activity in the intrapulmonary conducting airways of hamsters. Am. J. Respir. Cell and Mol. Biol. 10: 594-603.
- 23. Gong JL, McCarthy K M, Telford J, Tamatani T, Maiyasaka M, Schneeberger EE. 1992 Intraepithelial airway dendritic cells: A distinct subset of pulmonary dendritic cells obtained by microdissection. J. Exp. Med. 175: 797-807.
- 24. Granucci F, Zanoni I. 2009. The dendritic cell life cycle. Cell cycle 8: 3816-21
- 25. Griese M, Birrer P, Demirsoy A. 1997. Pulmonary surfactant in cystic fibrosis. Eur. Respir. J. 10:1983-1988.
- 26. Gumbiner BM. 1993. Breaking through the Tight Junction Barrier. The Journal of Cell Biology, 123, 1631-1633
- 27. Hart DJN. 1997. Dendritic cells: Unique leukocyte populations, which control the primary immune response. *Blood* 90: 3245-3287.
- Holt P G, Haining S, Nelson DJ, Sedwick JD. 1994. Origin and steady-state turnover of class II MHC-bearing dendritic cells in the epithelium of the conducting airways. J. Immunol. 153: 256-261.
- 29. Holt PG, Schon-Hegrad A, Mcmenamin PG. 1990. Dendritic cells in the respiratory tract. Int. Rev. Immunol. 6: 139-149.
- 30. Holt PG, Schon-Hegrad MA, Oliver J. 1988. MHC class II antigen-bearing dendritic cells in pulmonary tissues of the rat. Regulation of antigen presentation activity by endogenous macrophage populations. J. Exp. Med. 167:262-274.
- Ichiyasu H, McCormack JM, McCarthy KM, Dombkowski D, Preffer FI, Schneeberger EE.2004. Matrix metalloproteinase-9-deficient dendritic cells have impaired migration through tracheal epithelial tight junctions. Am J Respir Cell Mol Biol. 30:761-70. Epub 2003 Dec 4.
- 32. Inaba K, Witmer-Pack M, Inaba M, Hathcock KS, Sakuta H, Azuma M, Yagita H, Okumura K, Linsley P S, Ikehara S, Muramatsu S, Hodes RJ, Steinman RM. 1994.

The tissue distribution of the B7-2 costimulator in mice: abundant expression on dendritic cells in situ and during maturation in vitro. *J. Exp. Med.* 180:1849-1860.

- 33. Katayama I, Matsunaga T, Yokozeki H, Nishioka K. 1997. Blockade of costimulatory molecules B7-1 (CD80) and B7-2 (CD86) down-regulates induction of contact sensitivity by haptenated epidermal cells. *Br. J. Dermatol.* 136:846-852.
- 34. Katz S. I., Tamaki K., and D. H. Sachs (1979). Epidermal Langerhans cells are derived from cells originating in bone marrow. *Nature* 282:324-326.
- 35. Kiama, S. G., Cochand L., Karlsson L. M.,Nicod L. P., and P. Gehr(2001). Evaluation of phagocytic activity in human monocyte-derived dendritic cells. *Journal of Aerosol Medicine*14: 289-299.
- Kiama,S. G., D. Dreher , L Cochand, M. Kok,4 C. Obregon,5 L. Nicod and P. Gehr (2006) Host cell responses of Salmonella typhimurium infected human dendritic cells Immunology and Cell Biology (2006) 84, 475–481
- 37. Kilburn . H. (1968) A hypothesis for pulmonary clearance and its implications. Am. Rev. Respir. Dis. 98: 449-463.
- 38. Kradin R. L., Liu H. W., van Rooijen N., Springer K., Zhao L. H., and C. P. Leary (1999) Pulmonary immunity to Listeria is enhanced by elimination of alveolar macrophages. Am J Respir Crit Care Med 159:1967-74
- 39. Lamm M. E. (1997). Interaction of antigens and antibodies at mucosal surfaces. *Ann. Rev. Microbiol.* 51:311-340.
- 40. Lucas A. M. and L. C. Douglas (1934) Principals underlying ciliary activity in the respiratory tract. II. A comparison of nasal clearance in man, monkey and other mammals. Arch. Otolaryngol. Head Neck Surg. 20:518-541.
- 41. MacLean J. A., Xia W., Pinto C. E., Zhao L., Liu H. W., and R. L. Kradin ((1996) Sequestration of inhaled particulate antigens by lung phagocytes: A mechanism for the effective inhibition of pulmonary cell-mediated immunity. Am. J. Pathol. 148: 657-666.
- 42. McWilliam A. S., Nelson D., Thomas J. A., and P. G. Holt (1994) Rapid dendritic cells recruitment is a hallmark of the acute inflammatory response at mucosal surfaces. J. Exp. Med. 179: 1331-1336.
- 43. Meyer K. C., Sharma A., Brown R., Weatherly M., Moya F. R., Lewandoski J., and J. J. Zimmerman (2000) Function and composition of pulmonary surfactant and surfactant-derived fatty acid profiles are altered in young adults with cystic fibrosis. Chest 118:164-174.
- 44. Morrow P. E. (1988) Possible mechanisms to explain dust overloading of the lungs. Fundam. Appl. Toxicol. 10:369-84.
- 45. Nayak A, Dodagatta-Marri E, Tsolaki AG, Kishore U (2012). An insignt into the diverse roles of surfactant proteins, SP-A and SP-D in innate and adaptive immunity. Front Immunol. 3:131
- Nicod L. P. (1997) Function of human lung dendritic cells In "Lung Macrophages and Dendritic Cells in Health and Disease" (Lipscomb M. F., and Russels S. W. eds). Pp 311-334. Marcel Dekker, Inc: New York.
- 47. Nicod L. P., Lipscomb M. F., Weissler J. C., Lyons C. R., Albertson J., and G. B. Toews (1987). Mononuclear cells in human lung parenchyma. Characterization of a potent accessory cell not obtained by bronchoalveolar lavage. *Am. Rev. Respir. Dis.* 136:818-823.
- 48. Oberdoster, G., Ferin J., and B. E. Lehnert (1994) Correlation between particle size, in vitro particle persistence and lung injury. Environ. Health persp. 102(suppl. 5): 173-179.
- 49. Obregon C., Rothen-Rutishauser B., Kiama S.G., Gehr P., Nicod L. P. (2006). Exovesicles from human activated dendritic cells (DCS) fuse with resting DCS

allowing them to present allo-antigens. *American Journal of Pathology* 169:2127-2136

- 50. Palucka K., and J. Banchereau (1999). Linking innate and adaptive immunity: identification of precursor dendritic cells as the natural interferon-producing cells, their role in connecting two branches of the immune system. *Nature Med.* 5: 868-870.
- 51. Pavli P., Hume D. A., Van De Pol E., and W. F. Doe (1993). Dendritic cells, the major antigen-presenting cells of the human colonic lamina propria. *Immunol.* 78: 132-141
- 52. Phalen, Robert F. *Inhalation Studies: Foundations and Techniques*. Boca Raton, Fla: CRC Press, 1984.
- Pierre P., Turley S. J., Gatti E., Hull M., Meltzer J., Mirza A., Inaba K., Steinman R. M., and Mellman (1997). Developmental regulation of MHC class II transport in mouse dendritic cells. *Nature* 388:787-792.
- 54. Pison U., Max M., Neuendank A., Weissback S., and S. Pietschmann (1994) Host defense capacities of pulmonary surfactant: evidence of "non-surfactant" functions of the surfactant system. European Journal of Clin. Invest. 24: 586-599.
- 55. Pope C. A. 3<sup>rd</sup>, Ezzati M., Dockery D. W. (2009) Fine-particulate air pollution and life expectancy in the United States. N. Engl J. Med 360(4):376-86
- 56. Pope CA 3<sup>rd</sup> (2014). Particulate air pollution and lung function Am. J. Respir. Crit. Care Med. 190: 485-486Pugh C. W., MacPherson G. G., and H. W. Steer (1983). Characterization of nonlymphoid cells derived from rat peripheral lymph. *J. Exp. Med.* 157:1758-1779
- 57. Quaratino S., Duddy L. P., and M. Londei (2000). Fully competent dendritic cells as inducers of T cell anergy in autoimmunity. *Proc. Natl. Acad. Sci. USA* 97:10911-10916.
- 58. Rescigno M, Urbano M, Valzasina B, Francolini M, Rotta G, Bonasio R, Granucci F, Kraehenbuhl JP, Ricciardi-Castagnoli P (2001) Dendritic cells express tight junction proteins and penetrate gut epithelial monolayers to sample bacteria. Nat Immunol.;2(4):361-7.
- 59. Rissoan M. C., Soumelis V., Kadowaki N., Grouard G., Briere F., de Waal Malefyt R., and Y. J. Liu (1999). Reciprocal control of T helper cell and dendritic cell differentiation. *Science* 283:1183-1186.
- Roth, C., W.G.Kreyling, G.Scheuch, B.BuschandW.Stahlhofen (1997) Deposition and Clearance of Fine Particles in The Human Respiratory Tract. *Ann. occup. Hyg.*,Vol. 41, Supplement 1, pp. 503-508, 1997
- 61. Schleh C, Rothen-Rutishauser BM, Blank F, Lauenstein HD, Nassimi M, Krug N, Braun A, Erpenbeck VJ, Gehr P, Hohlfeld JM. (2012) Surfactant Protein D modulates allergen particle uptake and inflammatory response in a human epithelial airway model. Respir Res. 2012 Feb 1;13:8
- 62. Schon-Hegrad M. A., Oliver J., McMenamin P. G. and P. G. Holt (1991) Studies on the density, distribution, and surface phenotype of intraepithelial class II major histocompatibility complex antigen (Ia)-Bearing dendritic cells (DC) in the conducting airways. J. Exp. Med. 173: 1345-1356.
- 63. Schürch S., Gehr P., Im Hof V., Geiser M., and F. Green (1990) Surfactant displaces particles toward epithelium in airways and alveoli. Resp. Physiol. 80: 17-32.
- 64. Schürch S., Goerke J., and A. Clements (1976) Direct determination of surface tension in the lung. Proc. Nat. Aced. Sci. U. S. A. 73: 4698-4702.
- 65. Sertl K., Takemura T., Tschachler E., Ferrans V. J., Kaliner M. A., and E. M. Shevach (1986). Dendritic cells with antigen-presenting capability reside in airway epithelium, lung parenchyma, and visceral pleura. *J. Exp. Med.* 163:436-51.

- 66. Stahlhofen, W., Scheuch, G. and Bailey, M. R. (1994) Measurement of the tracheobronchial clearance of Particles after Aerosol Bolus Inhalation The Annals of Occupational HygieneVolume 38, Issue inhaled particles VII Pp. 189-196
- 67. Steinman R. M. (1991). The dendritic cell system and its role in immunogenicity. *Annu. Rev. Immunol.* 9: 271-296
- *68.* Steinman R. M., and Z. A Cohn (1973). Identification of a novel cell type in peripheral lymphoid organs of mice. I. morphological, quantitation, tissue distribution. *J. Exp. Med. 137:* 1142-1162.
- 69. Steinman R. M., and Z. A. Cohn (1974). Identification of a novel cell type in peripheral lymphoid organs of mice. II. Functional properties *in vitro. J. Exp. Med.* 139: 380-397.
- Takano K, Kojima T, Go M, Murata M, Ichimiya S, Himi T, Sawada N.(2005) HLA-DRand CD11c-positive dendritic cells penetrate beyond well-developed epithelial tight junctions in human nasal mucosa of allergic rhinitis. J Histochem Cytochem. May;53(5):611-9.
- 71. Turley S. J., Inaba K., Garrett W. S., Ebersold M., Unternaehrer J., Steinman R. M., and I Mellman (2000). Transport of peptide-MHC class II complexes in developing dendritic cells. *Science* 288:522-527
- 72. van Spriel AB<sup>1</sup>, de Jong EC Dendritic cell science: more than 40 years of history. (2014) J Leukoc Biol. 2013 Jan;93(1):33-8
- 73. Vermaelen K. Y., Carro-Muino I., Lambrecht B. N., and R. A. Pauwels (2001) Specific migratory dendritic cells rapidly transport antigen from airways to the thoracic lymph nodes. J. Exp. Med. 193: 51-60.
- 74. Walter E., Dreher D., Kok M., Thiele L, Kiama S. G., Gehr P., and H. P. Merkle (2001). Hydrophilic poly (DL-lactide-co-glycolide) microspheres for the delivery of DNA to human-derived macrophages and dendritic cells. *Journal of Controlled Release*76: 149-168.
- 75. Werner A., Salathe M., and T. G. O'Riordan (1996). Mucociliary Clearance in the airways Am. J. Crit. Care Med. 154: 1868-1902.
- 76. Wong B. R., Josien R., Lee S. Y., Sauter B., Li H. L., Steinman R. M., and Y. Choi (1997). TRANCE (tumor necrosis factor [TNF]-related activation-induced cytokine), a new TNF family member predominantly expressed in T cells, is a dendritic cell-specific survival factor. *J. Exp. Med.* 186:2075-80.